



RESEARCH ARTICLE

# Effectiveness of CaCl<sub>2</sub> treatment on quality attributes of banana fruit during storage

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## Abstract

Banana is a delicious fruit with excellent nutrient components beneficial for human health. In post harvest, banana fruit is quickly ripe within few days at ambient temperature condition and its quality is seriously degraded afterwards. Extension of its stability during storage and distribution is very necessary to enhance its economic value. This research evaluated the effectiveness of CaCl<sub>2</sub> treatment on the physicochemical quality attributes, phytochemical and antioxidant activities and potential enzymes of banana fruit during storage. Banana fruits were dipped in 3% CaCl<sub>2</sub> solution for 5 min before draining on racks for 30 min at ambient temperature to remove excess calcium chloride solution. Another banana group is dipped in clean water, leaved on racks for 30 min as the control. Both these fruit groups were then stored at 4±0.5°C with 85-90% relative humidity for 28 days. In 7 day-interval, each group is taken to verify quality attributes such as decay index, firmness, extractable juice recovery, ascorbic acid, carotenoid, total soluble solid, phenolic content, total flavonoid content, DPPH radical scavenging rate, H<sub>2</sub>O<sub>2</sub> content, catalase (CAT) activity, superoxide dismutase (SOD) activity of banana fruit during storage. Our findings demonstrated that CaCl<sub>2</sub> exerted a positive influence in limiting decay (2.24±0.11 mark); slowing down texture firmness reduction (7.25±0.10 N); enhancing the extractable juice recovery (85.49±1.12%); controlling ascorbic acid (12.79±0.13 mg/100 g), total soluble solid (16.40±0.13 °Brix), phenolic retention (161.35±1.19 mg GAE/100 g), flavonoid content (64.41±0.25 mg QE/100 g) and H<sub>2</sub>O<sub>2</sub> accumulation (20.09±0.10 μmol/g); improving carotenoid (21.83±0.13 μg/100 g), catalase (15.28±0.12 U/g), superoxide dismutase activity (5.60±0.10 U/g), antioxidant capacity of banana fruit (46.20±0.15%) during chilling storage. CaCl<sub>2</sub> treatment would be an effective approach to extend shelf life of banana fruit in commercial distribution.

## Keywords

banana fruit, calcium chloride treatment, chilling storage, quality attribute

## Introduction

Calcium is an important mineral element in preserving post harvest quality of fruit and vegetable. It involved in forming the cell wall structure and cellular membrane by providing a linkage between calcium and pectin to form a complex of pectate-calcium (1). This application could delay senescence without any detrimental impact on consumer acceptance (2). Calcium treatment stabilized plant cell wall and membrane integrity, improved texture firmness, lowered membrane lipid catabolism, prolonged shelf-life of agricultural products (3). These functions of calcium effectively protected cell

wall from catalyzing enzymes (4). Calcium is a vital secondary delivery in crop indication inspiration participating in physiological pathways and the interaction to different tension (5). In low-temperature disorders, sparked temporary accumulation of calcium ion is significant by various calcium capturing proteins, thus facilitating numerous physiological reactions in the cell (6). Calcium treatment is demonstrated to be effective on different kinds of fruits and vegetables such as chili pepper (7), loquat (8, 9), apple (10), peach (11, 12), persimmon (13), papaya (14, 15), gooseberry (16), strawberry (17), mango (18), purple shallot (19), soursop (20), tomato (21) and watermelon (22).

Banana (*Musa* spp.) belonged to Musaceae family which is widely cultivated in the tropical and subtropical locations like Africa, Asia, the Pacific, Latin America and the Caribbean. Banana is believed to have originated in Southeast Asia. Banana is highly preferred due to its unique favor, aroma, texture and attractive appearance. Banana is ranked 4<sup>th</sup> in the list of the most preferred food-stuffs. Banana is low in fat and protein but rich in carbohydrates, dietary fibers, vitamins, minerals with numerous therapeutic advantages (23). Banana is considered as a medicinal fruit against various ailments such as diarrhea, diabetes, cancer, ulcer, hypertension and infection (24). Banana belonged to climactic fruit exhibiting a respiratory peak in natural ripening at 20 °C. The quality of the green banana went down quickly during post harvest as a result of degradation in appearance, texture and taste. It is highly perishable within 2-3 days in hot humid condition (25); therefore a controlled ripening in the fresh state is very important in post harvest and distribution. Ripening is correlated to different biochemical modifications including texture softening, discoloration, off-aroma, astringent disappearance (26).

There were several notable studies mentioned to the application of calcium treatment on banana fruit. Banana (*Musa accuminata*) fruit is submerged in 50, 100 and 200 mg/l CaCl<sub>2</sub> to verify the impact of the divalent cation Ca<sup>2+</sup> on organoleptic senescence, color change, fat content and moisture content. Calcium is effective in controlling the senescence in banana by slowing down the breakdown of chlorophyll. Moreover, this application also increased fat content of banana. Ca<sup>2+</sup> participated in reinforcing cell membranes, retaining cell wall hardness, slowing down chlorophyll a decomposing enzyme, retarding respiration and decomposition of membrane lipid in banana (27). CaCl<sub>2</sub> treatment resulted to a higher pulp to peel ratio and an extended stability on banana fruit (28). Calcium treatment effectively increased in protein content while controlled the reduction of chlorophyll content and photosynthetic intensity in banana fruit (29). CaCl<sub>2</sub> treatment of 20 mM greatly inactivated proliferation of anthracnose disease caused by *Colletotrichum* (30).

Horticultural harvest encountered different physical injuries (mishandling, collision), inappropriate storage conditions (high storage temperature, fluctuation in relative humidity, high ethylene accumulation) and physiological disorders (ripening, microbial invasion) causing significant quality and quantity losses (31). Horticultural prod-

ucts were highly perishable throughout series of steps from harvesting, preservation, delivery, trading and consumption (32). Post-harvest losses were a big challenge in agricultural production because they directly affected to income of farmer, processor and distributor. The objective of our study investigated the influence of CaCl<sub>2</sub> treatment on the decay index, firmness, extractable juice recovery, ascorbic acid content, carotenoid content, total soluble solid content, total phenolic content, total flavonoid content, DPPH radical scavenging activity, H<sub>2</sub>O<sub>2</sub> content, catalase (CAT) activity and superoxide dismutase (SOD) activity of banana fruit during storage.

## Materials and Methods

### Material

Banana fruits in maturity, uniformity and without defect were harvested in orchard in My Tu district, Soc Trang province, Vietnam.

Chemical reagents were all analytical grade. Standard β-carotene, hexane, Folin-Ciocalteu reagent, and 2, 6-dichlorophenol indophenols were purchased from Sigma Aldrich (USA). Thrichloroacetic acid, methanol were supplied from Fisher Scientific (USA). Standard quercetin, gallic acid, aluminum chloride, sodium acetate, sodium carbonate, potassium phosphate, potassium iodine, nitroblue tetrazolium were purchased from Merck (Germany).

### Methodology

Banana fruits (30 samples) were randomly separated into two different groups (control and CaCl<sub>2</sub> treatment). Control group (15 samples) is dipped in clean water for 5 min. Another group (15 samples) is dipped in 3% CaCl<sub>2</sub> solution for 5 min. 3 replications were prepared for each experiment. Both groups were drained on racks for 30 min at ambient temperature to remove excess water. They were then kept at 4±0.5°C with 85-90% relative humidity for 28 days. In 7 day-interval, samples were taken to define decay index, firmness, extractable juice recovery, ascorbic acid content, carotenoid content, total soluble solid content, total phenolic content, total flavonoid content, DPPH radical scavenging activity, H<sub>2</sub>O<sub>2</sub> content, catalase (CAT) activity and superoxide dismutase (SOD) activity.

Decay index is determined by visual appearance of fruit from completely fresh (1 mark), decay 5-10% (2 marks), decay 10-30% (3 marks), decay 30-50% (4 marks), decay surpass 50% (5 marks) (29). Firmness (N) is verified by texture penetrometer (Stable Micro Systems, model: TA.XTplusC). Extractable juice recovery (%) is estimated as the difference between the initial mass and the weight of the pellet after centrifugation divided by the initial mass (33). Ascorbic acid content (mg/100 g) is determined by using a 2,6-dichlorophenol indophenol visual titration method described by AOAC (34). Carotenoid content (µg/100 g) is examined by high-performance liquid chromatography method (Techno, model: HPLC 580) using a polymeric C30 column (22). Total soluble solid content (°Brix) is evaluated by hand-held refractometer (Atago, model: Master-53M) (26). Total phenolic content (mg

GAE/100 g) is quantified by using Folin–Ciocalteu assay (20). The optical density is read by the absorbance at 765 nm using a spectrophotometer (Mettler Toledo, model: UV5) (35). Total flavonoid content (mg GE/100 g) is quantified by the aluminium calorimetric method. The optical density is read by absorbance at 415 nm using a spectrophotometer (Mettler Toledo, model: UV5) (36). DPPH radical scavenging activity (%) is identified by the percentage of inactivation to radical DPPH of one prototype solution concentration (37). H<sub>2</sub>O<sub>2</sub> content (μmol/g) is identified by homogenizing 5 g of banana pulp tissue in ice bath with 50 ml 0.1% trichloroacetic acid. The macerate paste is centrifuged at 10000×g for 20 min and 10 ml of the supernatant is added to 10 ml 10 mM potassium phosphate buffer (pH 7.5) and 20 ml 1 M KI. The supernatant absorbance is measured at wavelength 390 nm. The H<sub>2</sub>O<sub>2</sub> content (μmol/g) is expressed on a standard curve (38). Catalase (CAT) activity (U/g) is determined as the amount of enzyme reacting with 1 μmol of H<sub>2</sub>O<sub>2</sub> per min and absorbance is read at wavelength 240 nm (39). Super-oxide dismutase (SOD) activity (U/g) is identified as the amount of enzyme inducing 50% inhibition of nitroblue tetrazolium reduction (39).

### Statistical analysis

The experiments were run in triplicate with different groups of samples. The data were presented as mean ± standard deviation. Statistical analysis is performed by the Statgraphics Centurion version XVI. The mean value and standard deviation of a set of data obtained by analysis of random samples estimating the population statistics. 95% of results would be expected to lie within the range we described the lower and upper bounds of this range as the 95% confidence limits of the results. The differences between the pickling samples were analyzed using a one-way analysis of variance (ANOVA). A significant value is set at a 95% confidence interval (P<0.05). If significant differences were found, then post hoc analysis is performed using Duncan's multiple range tests.

## Results and Discussion

### Effect of CaCl<sub>2</sub> post harvest treatment on decay index, firmness, extractable juice recovery of banana fruit during storage

Decay significantly happened on banana fruit at the 21<sup>st</sup> day and accelerated afterwards. CaCl<sub>2</sub> treatment minimized decay better than control. At the 28<sup>th</sup> day of storage, the decay index in CaCl<sub>2</sub>-dipped sample (2.24±0.11 mark) is lower than that in the control (3.71±0.13 mark) (Table 1).

**Table 1.** Decay index (mark) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	1.00±0.10 <sup>a</sup>	1.91± 0.12 <sup>a</sup>	2.45±0.13 <sup>a</sup>	3.03± 0.14 <sup>a</sup>	3.71±0.13 <sup>a</sup>
CaCl <sub>2</sub>	1.00±0.10 <sup>a</sup>	1.24± 0.13 <sup>b</sup>	1.63±0.11 <sup>b</sup>	1.98± 0.13 <sup>b</sup>	2.24±0.11 <sup>b</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly (α = P=0.05)

Anthracoze originated from *Colletotrichum gloeosporioides* is a major rotten disease in bananas. On green fruit, the fungus only created latent contamination. At the ripening stage, anthracnose behaviors occurred, grew and fruit

became decay immediately (30). Firmness in banana fruit revealed a decrease trend during storage. However, the CaCl<sub>2</sub>-dipped sample showed a gradual decrease in texture firmness (7.25±0.11 N) while the control presented a sharp

**Table 2.** Firmness (N) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	8.32± 0.14 <sup>a</sup>	7.83± 0.15 <sup>b</sup>	7.14± 0.13 <sup>b</sup>	6.28± 0.14 <sup>b</sup>	5.48± 0.12 <sup>b</sup>
CaCl <sub>2</sub>	8.32± 0.14 <sup>a</sup>	8.19± 0.13 <sup>a</sup>	8.02± 0.11 <sup>a</sup>	7.76± 0.13 <sup>a</sup>	7.25± 0.11 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly (α = P=0.05)

firmness reduction (5.48±0.12 N) at the 28<sup>th</sup> day of storage (Table 2). Extractable juice recovery of CaCl<sub>2</sub> dipped banana

**Table 3.** Extractable juice recovery (%) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	47.53 ±0.13 <sup>a</sup>	50.06 ±0.20 <sup>b</sup>	58.72±0.17 <sup>b</sup>	70.25 ±0.14 <sup>b</sup>	72.90±0.16 <sup>b</sup> x ± 2s
CaCl <sub>2</sub>	47.53±0.13 <sup>a</sup>	54.79 ±0.18 <sup>a</sup>	69.18±0.15 <sup>a</sup>	83.71 ±0.10 <sup>a</sup>	85.49±0.12 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly (α = P=0.05)

built up remarkably (85.49±1.12 %) while that of the control increased slightly (72.90±1.16) during storage (Table 3). These findings emphasized that CaCl<sub>2</sub> treatment effectively retarded decay, slowed down texture firmness reduction and improved the extractable juice recovery during chilling storage. We did not observe morphological or physiological changes.

CaCl<sub>2</sub> contributed to the control of the fruit respiration rate thus delayed the ripening and stabilized the texture firmness (4). Preservation at low temperature induced to the degradation of the center membrane and cellular depression causing a limitation in the tissue firmness (40). Calcium maintained texture firmness of fruit and vegetable by establishing the bridge between pectic components within the cell-wall (41). The application of calcium improved new ionic bridges among calcium and galacturonic acids, reinforced the cell wall and especially the center lamella accounted for combining cells together. From that, the durability of the cell wall structure and center lamella could be enhanced (42, 43). Texture firmness is also maintained when de-esterified pectic acid balance created cross-links between anionic carboxylic segments and divalent cations limiting pectin dissolution (44).

CaCl<sub>2</sub> treated strawberry had better firmness compared to the control (45). Firmness of kiwi fruit would be higher by dipping in 1% CaCl<sub>2</sub> (46). 2% CaCl<sub>2</sub> strongly retarded spore germination and proliferation of the pathogen *Rhizopus stolonifer* (47). 1% CaCl<sub>2</sub> submergence slowed down ripening, enhanced self-defense to decay and preserved cell wall integrity of strawberry (48). Calcium treatment is proven be effective in retarding fungus proliferation, lowering physiological disorder and respiration (49, 50). Mature-green banana dipped in 200 mg/l of calcium chloride induced a significant retention of weight with a better shelf life (27). Calcium treatment inhibited mycelial

development of the fungus *Fusarium oxysporum* f. sp. *cepae* that causing decay on onion (51). 1.5% CaCl<sub>2</sub> strongly retarded spore germination and proliferation of apple anthracnose lesions caused by *Colletotrichum acutatum* (52). 2% CaCl<sub>2</sub> treatment increased persimmon fruit firmness during 4 month storage at 0 °C (13). Calcium treatment showed a possibility to minimize the loss of cell integrity on peach (12). Calcium chloride is beneficial to lessen the internal discoloration of pineapple in cold storage (53). Calcium treatment is effective in inhibition of fungal development on guava fruit caused by *Alternaria* sp., *Aspergillus niger*, *Botrytis cinerea*, *Fusarium solani* (54); proliferation of *Lasiodiplodia theobromae* on mango (55). Calcium is proven to effectively inhibit the chilling injury on pear fruit (56). An incorporation of 50 °C of water dipping and 0.64% calcium chloride is efficient to maintain texture firmness of purple shallot (19). 1.5% CaCl<sub>2</sub> treatment on green matured tomato resulted low decay rate during 20 days of storage (57).

#### Effect of CaCl<sub>2</sub> post harvest treatment on ascorbic acid, carotenoid, total soluble solid of banana fruit during storage

There is a slight reduction of ascorbic acid on the CaCl<sub>2</sub>-treated banana (12.79±0.13 mg/100 g) while a remarkable decrease of ascorbic acid on the control (11.68±0.12 mg/100 g) from day 7<sup>th</sup> to the day 28<sup>th</sup> of storage (Table 4).

**Table 4.** Ascorbic acid (mg/100 g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	14.28±0.17 <sup>a</sup>	13.64±0.14 <sup>b</sup>	13.09±0.13 <sup>b</sup>	12.45±0.14 <sup>b</sup>	11.68±0.12 <sup>b</sup>
CaCl <sub>2</sub>	14.28±0.17 <sup>a</sup>	14.03±0.13 <sup>a</sup>	13.86±0.11 <sup>a</sup>	13.17±0.12 <sup>a</sup>	12.79±0.13 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

This might be due to CaCl<sub>2</sub> limited the oxidative degradation of ascorbic acid. Ascorbic acid participated in the respiration circle as well as photo-degradation therefore there is a downtrend of this content during storage (58). Ascorbic acid is one of the most vital vitamins available in fruit and vegetable. However it is very sensitive to oxidative decomposition (59). Carotenoid content built up on CaCl<sub>2</sub>-treated

**Table 5.** Carotenoid ( $\mu$ g/100 g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	11.3±0.15 <sup>a</sup>	13.82±0.14 <sup>b</sup>	15.5±0.13 <sup>b</sup>	18.20±0.14 <sup>b</sup>	19.0±0.12 <sup>b</sup>
CaCl <sub>2</sub>	11.3±0.15 <sup>a</sup>	14.54±0.13 <sup>a</sup>	17.9±0.11 <sup>a</sup>	20.46±0.12 <sup>a</sup>	21.8±0.13 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

banana (21.83±0.13  $\mu$ g/100 g) higher than that on the control (19.07±0.12  $\mu$ g/100 g) (Table 5). Extended storage induced to a higher loss of carotenoid content, followed by the evolution of ripening (60). The total soluble solid content increased during storage with a lower accumulation on the CaCl<sub>2</sub>-treated banana (16.40±0.13 °Brix) compared to the control (19.23±0.12 °Brix) (Table 6). During storage, polysaccharides such as starches and pectins were metabolized into simple sugar (61). Our findings proved that CaCl<sub>2</sub> treatment significantly controlled ascorbic acid degrada-

**Table 6.** Total soluble solid (°Brix) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	8.32±0.12 <sup>a</sup>	11.24±0.14 <sup>a</sup>	14.92±0.12 <sup>a</sup>	17.04±0.10 <sup>a</sup>	19.23±0.12 <sup>a</sup>
CaCl <sub>2</sub>	8.32±0.12 <sup>a</sup>	9.67±0.15 <sup>b</sup>	12.51±0.14 <sup>b</sup>	14.75±0.13 <sup>b</sup>	16.40±0.13 <sup>b</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

tion; improved carotenoid content; and inhibited total soluble solid increment during cool storage.

High ascorbic acid content in tomato is noticed by CaCl<sub>2</sub> treatment compared to the control (62). Calcium treatment effectively retained firmness of chili pepper (7). CaCl<sub>2</sub> treatment preserved the highest ascorbic acid, lycopene content on tomato during 14 days of ambient storage (63). 6% CaCl<sub>2</sub> remarkably minimized decay, while retained higher firmness and ascorbic acid of tomato fruit in a prolonged shelf life (64). 1.5% CaCl<sub>2</sub> treatment effectively preserved firmness and reduced decay severity on papaya fruit caused by anthracnose incidence (15). 2% CaCl<sub>2</sub> treatment significantly maintained the ascorbic acid content in raspberry and strawberry fruits (17). CaCl<sub>2</sub> treatment induced to a great reduction of anthracnose lesion circles in the infected papaya (65). 4 % CaCl<sub>2</sub> pre-harvest treatment resulted to higher total ascorbic acid content on citrus (66). Calcium treatment increased carotenoid content while maintained chlorophyll content, photosynthetic intensity in banana fruit (29). Tomato fruit treated by 6% CaCl<sub>2</sub> had an improvement of physicochemical attributes (67). An incorporation of 2.5% CaCl<sub>2</sub> and 0.6 ppm 1-MCP created a synergistic effect to prolong watermelon stability with minimal weight loss and decay rate while firmness, total soluble solid, carotenoid and ascorbic acid contents could be retained in an efficient manner (22).

#### Effect of CaCl<sub>2</sub> post harvest treatment on total phenolic content, total flavonoid content, DPPH radical scavenging rate of banana fruit during storage

There is a trend of reduction total phenolic content during storage with a remarkable decrease on the control (140.19±1.37 mg GAE/100 g) and slight falling on the CaCl<sub>2</sub>-

**Table 7.** Total phenolic content (mg GAE/100 g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	184.5±1.21 <sup>a</sup>	174.18±1.30 <sup>b</sup>	163.27±1.25 <sup>b</sup>	151.34±1.41 <sup>b</sup>	140.19±1.37 <sup>b</sup>
CaCl <sub>2</sub>	184.5±1.21 <sup>a</sup>	179.32±1.17 <sup>a</sup>	172.56±1.09 <sup>a</sup>	167.20±1.20 <sup>a</sup>	161.35±1.19 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

treated banana (161.35±1.19 mg GAE/100 g) at the 28<sup>th</sup> day of storage (Table 7). Similarly, the total flavonoid content also decreased during storage with a sharp reduction on the control (57.63±0.33 mg QE/100 g) while the CaCl<sub>2</sub>-treated banana showed a better value (64.41±0.25 mg QE/100 g) (Table 8). DPPH of both the control and CaCl<sub>2</sub>-treated banana fruit showed a minor improvement (44.39±0.12% and 46.20±0.15%, respectively) during stor-



**Table 8.** Total flavonoid content (mg QE/100 g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	75.19±0.49 <sup>a</sup>	71.38±0.37 <sup>b</sup>	68.51±0.40 <sup>b</sup>	63.17±0.29 <sup>b</sup>	57.63±0.33 <sup>b</sup>
CaCl <sub>2</sub>	75.19±0.49 <sup>a</sup>	73.57±0.28 <sup>a</sup>	71.63±0.31 <sup>a</sup>	68.94±0.27 <sup>a</sup>	64.41±0.25 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

age (Table 9). These findings expressed that CaCl<sub>2</sub> treatment efficiently minimized the reduction of phenolic constituents as well as better retention of antioxidant capacity on banana fruit during preservation.

**Table 9.** DPPH (%) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	42.3±0.15 <sup>a</sup>	42.81±0.13 <sup>b</sup>	43.3±0.16 <sup>b</sup>	43.91±0.14 <sup>b</sup>	44.3±0.12 <sup>b</sup>
CaCl <sub>2</sub>	42.3±0.15 <sup>a</sup>	43.25±0.10 <sup>a</sup>	44.0±0.12 <sup>a</sup>	45.18±0.13 <sup>a</sup>	46.2±0.15 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

2% CaCl<sub>2</sub> continuously maintained DPPH radical scavenging capacity until the last day (68). 0.2 M CaCl<sub>2</sub> treatment caused a reduction of tocopherol content and an accumulation of total phenolic content in lettuce (69). 80 mM CaCl<sub>2</sub> treatment retained high total phenol on cherry fruit (70). 2% CaCl<sub>2</sub> treatment increased antioxidant activities of persimmon fruit during 4 month storage at 0°C (13). 2% calcium chloride treatment showed high total antioxidant activity and total phenolic content on papaya fruit during storage (15). The raspberry and strawberry fruits treated with 2% CaCl<sub>2</sub> showed a positive retention of the total phenolic content during 8 days of storage at 0°C and relative humidity of 95% (17). 4% CaCl<sub>2</sub> pre-harvest treatment resulted to higher total phenol, total flavonoid and antioxidant capacity on citrus (65). Hydrothermal-calcium chloride treatment resulted in higher L-ascorbic acid, total phenol content and powerful antioxidant ability on pepper (71). A combination of 50 °C of water dipping and 0.64% of calcium chloride is useful to retain bioactive components such as total phenolic, total flavonoid and quercetin of purple shallot (19).

### Effect of CaCl<sub>2</sub> post harvest treatment on H<sub>2</sub>O<sub>2</sub> content, catalase (CAT) activity, superoxide dismutase (SOD) activity of banana fruit during storage

H<sub>2</sub>O<sub>2</sub> content of banana fruit increased on both the control and treated sample with a sharp increase on the control (25.24±0.16 µmol/g) and a minor accumulation on the CaCl<sub>2</sub>-treated one (20.09±0.10 µmol/g) (Table 10). Hydrogen peroxide production during storage at high level is a negative

**Table 10.** H<sub>2</sub>O<sub>2</sub> content (µmol/g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	17.04±0.15 <sup>a</sup>	20.49±0.13 <sup>a</sup>	38.26±0.14 <sup>a</sup>	31.17±0.12 <sup>a</sup>	25.24±0.16 <sup>a</sup>
CaCl <sub>2</sub>	17.04±0.15 <sup>a</sup>	18.87±0.12 <sup>b</sup>	27.30±0.12 <sup>b</sup>	23.41±0.13 <sup>b</sup>	20.09±0.10 <sup>b</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

symptom of membrane damage and quality deterioration (72). Catalase activity significantly accumulated on the CaCl<sub>2</sub>-treated banana (15.28±0.12 U/g) higher than that on

**Table 11.** Catalase activity (U/g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	13.2±0.12 <sup>a</sup>	13.57±0.12 <sup>a</sup>	14.6±0.12 <sup>a</sup>	14.29±0.10 <sup>a</sup>	13.9±0.13 <sup>a</sup>
CaCl <sub>2</sub>	13.2±0.12 <sup>a</sup>	14.10±0.11 <sup>b</sup>	19.4±0.13 <sup>b</sup>	17.72±0.10 <sup>b</sup>	15.2±0.12 <sup>b</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

the control (13.91±0.13 U/g) (Table 11). Catalase activity is correlated to oxidative resistance and delayed ripening (73). High catalase activity is beneficial to eliminate reactive oxygen species, particularly O<sub>2</sub><sup>-</sup> released by biotransformation. Overload O<sub>2</sub><sup>-</sup> could speed up the membrane oxidative rancidity and senescence (74). Superoxide dismutase activity is increased during storage. CaCl<sub>2</sub>-treated banana had a high SOD value (5.60±0.10 U/g) compared to the control (4.97±0.13 U/g) (Table 12). These results expressed that CaCl<sub>2</sub> treatment effectively controlled H<sub>2</sub>O<sub>2</sub> accumulation while improved catalase and superoxide dismutase activity during cool storage.

**Table 12.** Superoxide dismutase activity (U/g) of banana fruit by the control and 3% CaCl<sub>2</sub> treatment during 28 days of storage

Storage (days)	0	7	14	21	28
Control	3.65±0.13 <sup>a</sup>	4.01±0.13 <sup>b</sup>	6.28±0.10 <sup>b</sup>	5.61±0.12 <sup>b</sup>	4.97±0.13 <sup>b</sup>
CaCl <sub>2</sub>	3.65±0.13 <sup>a</sup>	4.43±0.12 <sup>a</sup>	7.51±0.11 <sup>a</sup>	6.39±0.13 <sup>a</sup>	5.60±0.11 <sup>a</sup>

Figures are the mean of three replications; Figures in row followed by the same letter/s are not differed significantly ( $\alpha = P=0.05$ )

CaCl<sub>2</sub> treatment efficiently suppressed chilling injury and H<sub>2</sub>O<sub>2</sub> production; maintained firmness and extractable juice recovery; improved DPPH radical scavenging activity, catalase activity, superoxide dismutase activity of loquat fruit (9). Destabilization of the cell membrane is the main factor correlated to decay on fruit. Moreover, over production of hydrogen peroxide also induced fruit quality degradation (75). Superoxide dismutase and catalase were responsible for alleviation of fruit decay (76). Superoxide dismutase altered the redundant O<sub>2</sub><sup>-</sup> into hydrogen peroxide, while catalase triggered the hydrogen peroxide into water and oxygen (77). Calcium chloride treatment showed a significant influence on ripening physiology of banana fruit (78). Calcium treatment could retain a higher catalase activity in loquat (79) and apricot (80) during cold storage. Calcium treatment enhanced firmness and catalase activity of apple during 5 months of cold storage (81). Calcium chloride treatment enhanced superoxide dismutase and catalase activities contributing to better resistance to decay (82). Apricot treated by 0.5% CaCl<sub>2</sub> at 20 °C for 5 min showed an increase in catalase activity (83). 2% CaCl<sub>2</sub> treatment increased catalase activity of persimmon fruit during 4 month storage at 0°C (13). Hydrothermal-calcium chloride treatment showed lower peroxidase, polyphenol oxidase, and phenylalanine ammonia lyase activities with exception of higher catalase activity in the pepper during 32 days of

storage at 8 °C (70).

## Conclusion

Banana is one of the most available, delicious and nutritional fruits. Calcium chloride is used as a firming agent to extend shelf-life, physicochemical attributes such as decay rate, firmness, extractable juice recovery, ascorbic acid, carotenoid, total soluble solid of banana fruit. 3% CaCl<sub>2</sub> post-harvest treatment is especially efficient to control the degradation of bioactive components (total phenolic and flavonoid contents) and antioxidant power (DPPH). CaCl<sub>2</sub> treatment significantly retarded H<sub>2</sub>O<sub>2</sub> accumulation while improved catalase and superoxide dismutase activity of banana fruit during storage. CaCl<sub>2</sub> submergence is a practical method to prolong the stability and nutritional quality of banana fruit. High retention of total phenolic and flavonoid contents contributed to better antioxidant power for banana fruit. Low H<sub>2</sub>O<sub>2</sub> content while high and superoxide dismutase activity would be beneficial for banana fruit during storage. Findings of this research greatly contributed to the income improvement for farmer, processor and distributor to market banana fruit on trade.

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## Authors' contributions

Nguyen Phuoc Minh arranged the experiments and also wrote the manuscript.

## Compliance with ethical standards

**Conflict of interest:** The author strongly confirmed that this research is conducted with no conflict of interest.

**Ethical issues:** None.

## References

- Hocking B, Tyerman SD, Burton RA, Gilliam M. Fruit calcium: Transport and physiology. *Frontiers in Plant Science*. 2016;7:569. <https://doi.org/10.3389/fpls.2016.00569>
- Lester GE, Grusak MA. Post harvest application of calcium and magnesium to honeydew and netted muskmelons: Effects on tissue ion concentrations, quality and senescence. *Journal of American Society Horticulture Science*. 2004;124:545-52. <https://doi.org/10.21273/JASHS.124.5.545>
- Picchioni GA, Watada AE, Conway WS, Whitaker BD, Sams CE. post harvest calcium infiltration delays membrane lipid catabolism in apple fruit. *Journal of Agriculture and Food Chemistry*. 1998;46:2452-57. <https://ucanr.edu/datastoreFiles/234-1737.pdf>
- White PJ, Broadley MR. Calcium in plants. *Annals of Botany*. 2003;92:487-511.
- Ranty B, Aldon D, Cotellet V, Galaud JP, Thuleau P, Mazars C. Calcium sensors as key hubs in plant responses to biotic and abiotic stresses. *Frontiers in Plant Science*. 2016;7:327. <https://doi.org/10.3389/fpls.2016.00327>
- Yuan P, Yang T, Poovaiah BW. Calcium signaling-mediated plant response to cold stress. *International Journal of Molecule Science*. 2018;19:3896. <https://dx.doi.org/10.3390%2Fijms19123896>
- Tunde-Akintunde TY. Effect of pretreatment on drying time and quality of chilli pepper. *Journal of Food Processing and Preservation*. 2010;34(4):595-608. <https://doi.org/10.1111/j.1745-4549.2008.00360.x>
- Attig A, Nadeem AA, Azhar H. Effect of calcium chloride treatments on quality characteristics of loquat fruit during storage. *Pakistan Journal of Botany*. 2010;42(1):181-88.
- Hou Y, Li Z, Zheng Y, Jin P. Effects of CaCl<sub>2</sub> treatment alleviates chilling injury of loquat fruit (*Eriobotrya japonica*) by modulating ROS homeostasis. *Foods*. 2021;10:1662. <https://doi.org/10.3390/foods10071662>
- Chardonnet CO, Charron CS, Sams CE, Conway WS. Chemical changes in the cortical tissue and cell walls of calcium infiltrated 'Golden Delicious' apples during storage. *post harvest Biology and Technology*. 2003;28:97-111. <https://ucanr.edu/datastoreFiles/234-1666.pdf>
- Sohail M, Ayub M, Khalil SA, Zeb A, Ullah F, Afridi SR, Ullah R. Effect of calcium chloride treatment on post harvest quality of peach fruit during cold storage. *International Food Research Journal*. 2015;22(6):2225-29. [http://www.ifrj.upm.edu.my/22%20\(06\)%202015/\(7\).pdf](http://www.ifrj.upm.edu.my/22%20(06)%202015/(7).pdf)
- Techakanon C, arrett DM. The effect of calcium chloride and calcium lactate pretreatment concentration on peach cell integrity after high-pressure processing. *International Journal of Food Science and Technology*. 2017;52:635-43. <http://www.fruitandvegetable.ucdavis.edu/files/263061.pdf>
- Bagheri M, Esna-Ashari M, Ershadi A. Effect of post harvest calcium chloride treatment on the storage life and quality of persimmon fruits (*Diospyros kaki* Thunb.) cv. 'Karaj'. *International Journal of Horticultural Science and Technology*. 2015;2(1):15-26. <https://dx.doi.org/10.22059/ijhst.2015.54260>
- Mahmud TMM, Al Eryani-Raqeeb A, Syed Omar SR, Mohamed Zaki AR, Abdul-Rahman AE. Effects of different concentrations and applications of calcium on storage life and physicochemical characteristics of papaya (*Carica papaya* L.). *American Journal of Agricultural and Biological Sciences*. 2008;3(3):526-33. <https://doi.org/10.3844/ajabssp.2008.526.533>
- Madani B, Mirshekari A, Yahia E. Effect of calcium chloride treatments on calcium content, anthracnose severity and antioxidant activity in papaya fruit during ambient storage. *Journal of the Science of Food and Agriculture*. 2016;96(9):2963-68. <https://doi.org/10.1002/jsfa.7462>
- Reyes-Medina JA, Pinzon EH, Alvarez-Herrera JG. Effect of calcium chloride and refrigeration on the quality and organoleptic characteristics of cape gooseberry (*Physalis peruviana* L.). *Acta Agronomy*. 2017;66(1):15-20. <http://dx.doi.org/10.15446/acag.v66n1.50610>
- Tamar T, Levan G, Merab J, Louise W. Potential antioxidant retention and quality maintenance in raspberries and strawberries treated with calcium chloride and stored under refrigeration. *Brazilian Journal of Food Technology*. 2017;20:e2016089. <http://dx.doi.org/10.1590/1981-6723.8916>
- Anna B, Hosea M, Lucy C, Theodosy M, Abdul K, George T, Jaspas S, Gopinadhan P, Alan S, Jayasankar S, Maulid M. Evaluation of post-harvest losses and shelf life of fresh mango (*Mangifera indica* L.) in Eastern zone of Tanzania. *International Journal of Fruit Science*. 2020;20(4):855-70. <https://dx.doi.org/10.1080/15538362.2019.1697411>
- Thuy NM, Tuyen NTM, Tai NV. Combination of mild heat and calcium chloride treatment on the texture and bioactive compounds of purple shallot. *Food Research*. 2020;4(5):1681-87. <https://www.myfoodresearch.com/uploads/8/4/8/5/84855864/>

- [\\_37\\_\\_fr-2020-245\\_thuy.pdf](#)
20. Santos EXD, Repolho RPJ, Sanches AG. The preservative effect of bee wax and calcium chloride coating on the quality and firmness of graviolas (*Annona muricata* L.). *MOJ Food Process Technology*. 2020;8(2):32–38. <https://doi.org/10.15406/mojft.2020.08.00239>
  21. Makonnen T, Fikreyohannes G, Kebede W. Impacts of harvesting stages and pre-storage treatments on shelf life and quality of tomato (*Solanum lycopersicum* L.), *Cogent Food and Agriculture*. 2021;7(1):1863620. <https://doi.org/10.1080/23311932.2020.1863620>
  22. Nguyen MP. Synergistic effect of calcium chloride and 1-Methylcyclopropene on storage of watermelon (*Citrullus lanatus* (Thunb.) Matsum. and Nakai). *Plant Science Today*. 2021;8(1):118–22. <https://horizonpublishing.com/journals/index.php/PST/article/view/960>
  23. Happi ET, Wathelet B, Paquot M. Changements texturaux et biochimiques des fruits du bananier au cours de la maturation. *Biotechnology Agronomy Society*. 2008;12(1):89–98.
  24. Jyothirmayi N, Rao NM. Banana medicinal uses. *Journal of Medical Science Technology*. 2015;4(2):152–60.
  25. Simran A, Saleem S, Rakesh G, Naseer A. Effects of anti-browning pretreatments on browning of banana pulp. *International Journal of Current Microbiology and Applied Sciences*. 2018;7(4):242–49. <https://doi.org/10.20546/ijcmas.2018.704.027>
  26. Maduwanthi SDT, Marapana RAUJ. Induced ripening agents and their effect on fruit quality of banana. *International Journal of Food Science*. 2019;2520179. <https://doi.org/10.1155/2019/2520179>
  27. Aghofack JN, Yambou TN. Effects of Calcium chloride and magnesium sulphate treatments on the shelf-life of climacteric banana and non-climacteric pineapple fruits. *Cameroon Journal of Experimental Biology*. 2006;1(1):34–38. <https://doi.org/10.4314/cajeb.v1i1.37923>
  28. Sakariyawo O, Mengestie B, Getachew Y, Kidane S. Post harvest application of calcium and sodium chloride formulations on ripening, shelf life and quality of banana (*Musa* spp cv. *Grand naine*) under jimma condition. *Nigerian Journal of Horticultural Science*. 2009;14(1):28–31. <https://doi.org/10.4314/njhs.v14i1.62162>
  29. Phounzong-Tafre E, Kouete JO, Aghofack-Nguemezi J. Effects of calcium chloride treatment on the photosynthetic capacity and intensity of banana fruit during ripening. *Journal of Advances in Biology and Biotechnology*. 2019;21(4):1–9. <https://journaljabb.com/index.php/JABB/article/view/30098/56478>
  30. Le TT, Vo THD, Nguyen TML, Vo TK, Trieu PL. Effects of calcium chloride treatment on suppression of fruit anthracnose disease caused by *Colletotrichum gloeosporioides*. *AGU International Journal of Sciences*. 2019;7(3):75–83. <https://apps.agu.edu/vn/qlkh/uploads/1575014361-maket-9-part-c-e2019c01-vo-thi-huong-duong-bai-hoan-chinhpdf.pdf>
  31. Musasa ST, Mvumi BM, Manditsera FA, Chinhanga J, Musiyandaka S, Chigwedere C. Post-harvest orange losses and small-scale farmers' perceptions on the loss causes in the fruit value chain: A case study of Rusitu Valley, Zimbabwe. *Food Science Quality Management*. 2013;18:1–8. <https://www.iiste.org/Journals/index.php/FSQM/article/view/7173>
  32. Murthy DS, Gajanana TM, Sudha M, Dakshinamoorthy V. Marketing and post-harvest losses in fruit: Its implications on availability and economy. *Marketing*. 2009;64(2):259–75.
  33. Cao S, Yang Z, Cai Y, Zheng Y. Fatty acid composition and antioxidant system in relation to susceptibility of loquat fruit to chilling injury. *Food Chemistry*. 2011;127:1777–83. <https://doi.org/10.1016/j.foodchem.2011.02.059>
  34. AOAC. Official method of analysis. 18<sup>th</sup> ed. Washington DC: Association of Official Analytical Chemists. 2005.
  35. Nizar S, Elhadi MM, Algaili MA, Hozeifa MH, Mohamed O. Determination of total phenolic content and antioxidant activity of roselle (*Hibiscus sabdariffa* L.) calyx ethanolic extract. *Standard Research Journal of Pharmacy and Pharmacology*. 2014;1:034–039.
  36. Formagio ASN, Ramos DD, Vieira MC, Ramalho SR, Silva MM, Zárate NAH, Foglio MA, Carvalho JE. Phenolic compounds of *Hibiscus sabdariffa* and influence of organic residues on its antioxidant and antitumoral properties. *Brazilian Journal of Biology*. 2015;75:69–76. <https://doi.org/10.1590/1519-6984.07413>
  37. Valentao P, Fernandes E, Carvalho F, Andrade PB, Seabra RM, Bastos ML. Antioxidant activity of *Centarium erythraea* infusion evidenced by its superoxide radical scavenging and xanthine oxidase inhibitory activity. *J Agric Food Chem*. 2001;49:3476–79. <https://doi.org/10.1021/jf001145s>
  38. Velikova V, Yordanov I, Edreva A. Oxidative stress and some antioxidant systems in acid rain-treated bean plants: Protective role of exogenous polyamines. *Plant Science*. 2000;151:59–66. [https://doi.org/10.1016/S0168-9452\(99\)00197-1](https://doi.org/10.1016/S0168-9452(99)00197-1)
  39. Zhang Y, Jin P, Huang YP, Shan TM, Wang L, Li YY, Zheng YH. Effect of hot water combined with glycine betaine alleviates chilling injury in cold-stored loquat fruit. *post harvest Biology and Technology*. 2016;118:141–47. <http://dx.doi.org/10.1016/j.postharvbio.2016.04.010>
  40. Siddiqui S, Bangerth F. The effect of calcium infiltration on structural changes in cell walls of stored apples. *Journal of Horticultural Science*. 1996;71(5):703–08. <https://doi.org/10.1080/14620316.1996.11515450>
  41. Demarty M, Morvan C, Thellier M. Ca and the cell wall. *Plant Cell Environment*. 1984;7:441–48.
  42. Munoz PH, Almenar E, Valle VD, Velez D, Gavara R. Effect of chitosan coating combined with post harvest calcium treatment on strawberry (*Fragaria ananassa*) quality during refrigerated storage. *Journal of Food Chemistry*. 2008;110:428–35. <https://doi.org/10.1016/j.foodchem.2008.02.020>
  43. Angeletti P, Castagnasso H, Miceli E, Terminiello L, Concellón A, Chaves A, Vicente A. Effect of preharvest calcium applications on post harvest quality, softening and cell wall degradation of two blueberry (*Vaccinium corymbosum*) varieties. *post harvest Biology and Technology*. 2010;58:98–103. <http://dx.doi.org/10.1016/j.postharvbio.2010.05.015>
  44. Krall SM, McFeeters RF. Pectin hydrolysis; effect of temperature, degree of methylation, pH, and calcium on hydrolysis rates. *Journal of Agriculture and Food Chemistry*. 1998;46:1311–15. <https://doi.org/10.1021/jf970473y>
  45. Rosen JC, Kader AA. post harvest physiology and quality maintenance of sliced pear and strawberry fruits. *Journal of Food Science*. 1989;54:656–59. <https://doi.org/10.1111/j.1365-2621.1989.tb04675.x>
  46. Agar IT, Massantini R, Hess-Pierce B, Kader AA. post harvest CO<sub>2</sub> and ethylene production and quality maintenance of fresh cut kiwifruit slices. *Journal of Food Science*. 1999;64:433–40.
  47. Tian SP, Fan Q, Xu Y, Jiang AL. Effects of calcium on biocontrol activity of yeast antagonists against the post harvest fungal pathogen *Rhizopus stolonifer*. *Plant Pathology*. 2002;51:352–58. <https://doi.org/10.1046/j.1365-3059.2002.00711.x>
  48. Lara I, Garcia P, Vendrell M. Modifications in cell wall composition after cold storage of calcium-treated strawberry fruit. *post harvest Biology Technology*. 2004;34:331–39. <https://doi.org/10.1016/j.postharvbio.2004.05.018>
  49. Ramirez J, Galvis J, Fischer G. Maduración poscosecha de la feijoa (*Acca sellowiana* Berg) tratada con CaCl<sub>2</sub> en tres temperaturas de almacenamiento. *Agron Colomb*. 2005;23(1):117–27.



50. Hernandez-Munos P, Almenar E, Ocio M, Gavara R. Effect of calcium dips and chitosan coatings on post harvest life of strawberries (*Fragaria x ananassa*). post harvest Biology and Technology. 2006;39(3):247-53. <http://dx.doi.org/10.1016/j.postharvbio.2005.11.006>
51. Turkkan M. Antifungal effects of various salts against *Fusarium oxysporum* f.sp. *capae*, the causal agent of *Fusarium* basal rot of onion. Journal of Agricultural Sciences. 2013;19, 178-87. <http://dergiler.ankara.edu.tr/dergiler/15/1814/19158.pdf>
52. Stosic S, Stojanovic S, Milosavjevic A, Polovac EP, Zivkovic S. Effect of calcium salts on post harvest fungal pathogens *in vitro*. Plant Protection. 2014;65(1):40-46. <http://dx.doi.org/10.5937/zasbilj1401040S>
53. Youryon P, Supapvanich S, Kongtrakool P, Wongs-Aree C. Calcium chloride and calcium gluconate peduncle infiltrations alleviate the internal browning of queen pineapple in refrigerated storage. Horticulture and Environment Biotechnology. 2018;59:205-13. <https://doi.org/10.1007/s13580-018-0028-9>
54. Hassanein RA, Salem EA, Zahran AA. Efficacy of coupling gamma irradiation with calcium chloride and lemongrass oil in maintaining guava fruit quality and inhibiting fungal growth during cold storage. Folia Horticulture. 2018;30(1):67-78. <https://doi.org/10.2478/fhort-2018-0007>
55. Nur Fatimma A, Munirah MS, Sharifah Siti Maryam SAR, Najihah A, Nur Ain Izzati MZ. Efficacy of *Allium sativum* extract as post-harvest treatment of fruit rot of mango. Plant Pathology and Quarantine. 2018;8(2):144-52. [https://plantpathologyquarantine.org/pdf/PPQ\\_8\\_2\\_6.pdf](https://plantpathologyquarantine.org/pdf/PPQ_8_2_6.pdf)
56. Zhang L, Wang J, Zhou B, Li G, Liu Y, Xia XL, Xiao ZG, Fei L, Ji SJ. Calcium inhibited peel browning by regulating enzymes in membrane metabolism of 'Nanguo' pears during post-ripeness after refrigerated storage. Science Horticulture. 2019;244:15-21. <http://dx.doi.org/10.1016/j.scienta.2018.09.030>
57. Rahel D, Neela S, Solomon WF. Effect of different concentrations of the gibberellic acid and calcium chloride dipping on quality and shelf-life of kochoro variety tomato. Philippine Journal of Science. 2021;150(1):335-49. [https://philjournalsci.dost.gov.ph/images/pdf/pjs\\_pdf/vol150no1/effect\\_of\\_different\\_concentrations\\_of\\_the\\_gibberellic\\_acid\\_pdf](https://philjournalsci.dost.gov.ph/images/pdf/pjs_pdf/vol150no1/effect_of_different_concentrations_of_the_gibberellic_acid_pdf)
58. Davey MW, Montagu MV, Inze D, Sanmartin M, Kanellis A, Smirnoff N, Benzie IJJ, Strain JJ, Favell D, Fletcher J. Plant L-ascorbic acid: chemistry, function, metabolism, bioavailability and effects of processing. Journal of the Science of Food and Agriculture. 2000;80:825-60. [https://doi.org/10.1002/\(SICI\)1097-0010\(20000515\)80:7%3C825::AID-JSFA598%3E3.0.CO;2-6](https://doi.org/10.1002/(SICI)1097-0010(20000515)80:7%3C825::AID-JSFA598%3E3.0.CO;2-6)
59. Veltman RH, Kho RMA, Van-Schaik CR, Sanders MG, Oosterhaven J. Ascorbic acid and tissue browning in pears under controlled atmosphere conditions. Post Harvest Biology and Technology. 2000;19:129-37. [https://doi.org/10.1016/S0925-5214\(00\)00095-8](https://doi.org/10.1016/S0925-5214(00)00095-8)
60. Msogoya TJ, Majubwa RO, Maerere AP. Effects of harvesting stages on yield and nutritional quality of African eggplant (*Solanum aethiopicum* L.) fruits. Journal of Applied Bioscience. 2014;78(1):6590-99. <http://dx.doi.org/10.4314/jab.v78i1.1>
61. Hussain PR, Dar MA, Meena RS, Mir MA, Shafi F, Wani AM. Changes in quality of apple (*Malus domestica*) cultivars due to gamma irradiation and storage conditions. Journal of Food Science and Technology. 2008;45:444-49.
62. Subbiah S, Perumal GR. Post harvest treatment of mangoes with calcium. Tropical Science. 1996;36:14-17.
63. Maedeh G, Neena J, Elnaz S. Effect of post harvest treatment on stored cherry tomatoes. Journal of Nutrition and Food Sciences. 2012;2:8. <https://doi.org/10.4172/2155-9600.1000157>
64. Eric A, Ibok O, Patrick K. Effect of maturity stage and post harvest calcium chloride treatment on the quality and storage life of tomatoes (*Lycopersicon esculentum* Mill). Journal of Post Harvest Technology. 2015;3(3):74-81. <http://hdl.handle.net/123456789/9737>
65. Ayon-Reyna LE, Lopez-Valenzuela JA, Delgado-Vargas F, Lopez-Lopez ME, Molina-Corral FJ, Carrillo-Lopez A, Vega-Garcia MO. Effect of the combination hot water – calcium chloride on the *in vitro* growth of *Colletotrichum gloeosporioides* and the post harvest quality of infected papaya. The Plant Pathology Journal. 2017;33:572-81. <https://dx.doi.org/10.5423%2FPPJ.OA.01.2017.0004>
66. Moghadam JF, Hashempour A. Effect of pre-harvest calcium chloride spraying on maintaining fruit bioactive compounds and antioxidant capacity of three citrus cultivars during storage. Iranian Journal of Horticultural Science and Technology. 2018;19(4):485-96. <http://journal-irshs.ir/article-1-110-en.pdf>
67. Fayez S, Tawfiq Q. Effect of calcium chloride post harvest treatment in combination with plant natural substance coating on fruit quality and storability of tomato (*Solanum lycopersicum*) fruits during cold storage. Journal of Applied Botany and Food Quality. 2021;94:100-07. <https://doi.org/10.5073/JABFQ.2021.094.012>
68. Antunes MDC, Dandlen S, Cavaco AM, Miguel G. Effects of post harvest application of 1-MCP and postcutting dip treatment on the quality and nutritional properties of fresh-cut kiwifruit. Journal of Agriculture and Food Chemistry. 2010;58:6173-81. <https://dx.doi.org/10.1021/jf904540m>
69. Irena P, Katarzyna O. Effect of foliar calcium chloride treatment on the level of chlorogenic acid,  $\beta$ -carotene, lutein and tocopherols in lettuce (*Lactuca sativa* L.). Acta Agrobotanica. 2011;64(1):65-72. <https://doi.org/10.5586/aa.2011.008>
70. Aghdam MS, Dokhanieh AY, Hassanpour H, Fard JR. Enhancement of antioxidant capacity of cornelian cherry (*Cornus mas*) fruit by post harvest calcium treatment. Science Horticulture. 2013;161:160-64. <https://doi.org/10.1016/j.scienta.2013.07.006>
71. Haishan X, Shenghua D, Hui Z, Youjin Y, Fangming D, Rongrong W. Quality attributes and related enzyme activities in peppers during storage: effect of hydrothermal and calcium chloride treatment. International Journal of Food Properties. 2019;22(1):1475-91. <https://doi.org/10.1080/10942912.2019.1653909>
72. Li ZY, Wang L, Xie B, Hu SQ, Zheng YH, Jin P. Effects of exogenous calcium and calcium chelant on cold tolerance of post harvest loquat fruit. Science Horticulture. 2020;269:109391. <https://doi.org/10.1016/j.scienta.2020.109391>
73. Huang RH, Xia RX, Hu LM, Lu YM, Wang MY. Antioxidant activity and oxygen-scavenging system in orange pulp during fruit ripening and maturation. Science Horticulture. 2007;113:166-72. <https://doi.org/10.1016/j.scienta.2007.03.010>
74. Gao HY, Zeng Q, Ren ZN, Li PZ, Xu XX. Effect of exogenous  $\gamma$ -aminobutyric acid treatment on the enzymatic browning of fresh-cut potato during storage. Journal of Food Science Technology. 2018;55:5035-44. <https://doi.org/10.1007/s13197-018-3442-1>
75. Habibi F, Ramezani A, Guillen F, Martinez-Romero D, Serrano M, Valero D. Susceptibility of blood orange cultivars to chilling injury based on antioxidant system, physiological and biochemical responses at different storage temperatures. Foods. 2020;9:1609. <https://doi.org/10.3390/foods9111609>
76. Yao M, Ge W, Zhou Q, Zhou X, Luo M, Zhao Y, Wei B, Ji S. Exogenous glutathione alleviates chilling injury in post harvest bell pepper by modulating the ascorbate-glutathione (AsA-GSH) cycle. Food Chemistry. 2021;352:129458. <https://doi.org/10.1016/j.foodchem.2021.129458>
77. Mittler R. Oxidative stress, antioxidants and stress tolerance. Trends in Plant Science. 2002;7:405-10. [https://doi.org/10.1016/S1360-1385\(02\)02312-9](https://doi.org/10.1016/S1360-1385(02)02312-9)
78. Antenette NP, Anjani MK. post harvest calcium chloride treat-



- ments do not help to increase shelf-life of bananas. *Fruits*. 2002;57:87–94. <https://dx.doi.org/10.1051/fruits:2002008>
79. Akhtar A, Abbasi NA, Hussain A. Effect of calcium chloride treatments on quality characteristic of loquat fruit during storage. *Pakistan Journal of Botany*. 2010;42(1):181-88. [http://www.pakbs.org/pjbot/PDFs/42\(1\)/PJB42\(1\)181.pdf](http://www.pakbs.org/pjbot/PDFs/42(1)/PJB42(1)181.pdf)
80. Ali S, Masud T, Abbasi KS, Mahmood T, Hussain I. Influence of CaCl<sub>2</sub> on biochemical composition, antioxidant and enzymatic activity of apricot at ambient storage. *Pakistan Journal of Nutrition*. 2013;12(5):476-83. <https://dx.doi.org/10.3923/pjn.2013.476.483>
81. Elham S, Vali R, Yavar S. Effect of calcium chloride (CaCl<sub>2</sub>) on post harvest quality of apple fruits. *African Journal of Agricultural Research*. 2011;6(22):5139-43. <https://doi.org/10.5897/AJAR11.1189>
82. Shi H, Ye T, Ba OZ, Liu X, Chan Z. Comparative proteomic and metabolomic analyses reveal mechanisms of improved cold stress tolerance in bermudagrass (*Cynodon dactylon* (L.) Pers.) by exogenous calcium. *Journal of Integrative Plant Biology*. 2014;56:1064–79. <https://doi.org/10.1111/jipb.12167>
83. Wu B, Guo Q, Wang GX, Peng XY, Wang JD, Che FB. Effects of different post harvest treatments on the physiology and quality of ‘xiaobai’ apricots at room temperature. *Journal of Food Science Technology*. 2015;52:2247–55. <https://doi.org/10.1007/s13197-014-1288-8>

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