



#### **RESEARCH ARTICLE**

# Pollination ecology and flowering rhythm of *Gloriosa superba* L.: Behavioural insights into principal pollinators

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#### **OPEN ACCESS**

#### **ARTICLE HISTORY**

Received: 01 November 2024 Accepted: 25 March 2025 Available online Version 1.0: 28 April 2025



#### **Additional information**

**Peer review**: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

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#### **CITE THIS ARTICLE**

Priyanka R, Raj KT, Yash PS, Meena T, Mohammad AW, Kiran S, Bimal KS. Pollination ecology and flowering rhythm of *Gloriosa superba* L.: Behavioural insights into principal pollinators. Plant Science Today (Early Access). https://doi.org/10.14719/pst.5850

#### **Abstract**

Gloriosa superba L. (Glory lily), is an endangered medicinal plant valued for its high colchicine content. Understanding the floral biology and pollination ecology of G. superba is crucial for identifying factors limiting seed setting. This study was carried out at Dr. YS Parmar University of Horticulture and Forestry, Solan, Himachal Pradesh, India, during 2021-2022, focusing on G. superba's floral biology and pollinator activity from anthesis to senescence. The scarlet red flowers were born on pedicels, with tepals measuring 6.69 × 1.45 cm tepal and a mean weight of  $1.72 \pm 0.04$  mg. Pollen viability was highest on the one day after anthesis and stigma receptivity peaked on the day of anthesis. Among nine flower stages, nectar volume and nectar sugar concentrations were maximum at 6th and 7th stages, respectively. Three kinds of insect floral visitors viz., allotrophous, hemitrophous and eutrophous from 6 orders, 14 families and 16 genera were recorded. The Shannon, Simpson, dominance and Margalef richness indices during 2021 and 2022 were 2.58, 0.16, 0.83 and 1.3 and 2.15, 0.22, 0.83 and 1.22, respectively. Fruit set (86.45  $\pm$  0.21 %), pod length (7.24  $\pm$ 0.23 cm) and colchicine percentage were highest in hand pollination. However, test weight (2.02 ± 0.01 g), germination percentage (77.98 ± 0.26 %) and colchicine percentage in seed under honeybee pollination were statistically similar to blower-assisted pollination.

#### **Keywords**

foraging behaviour seed colchicine content; glory lily; insect diversity; nectar studies

#### Introduction

Gloriosa superba L. commonly known as Malabar glory lily, is a commercially valuable medicinal plant of the family Liliaceae. It contains the alkaloid colchicine in its seeds, stems, leaves and tubers (1, 2). The species is distributed across tropical regions, from the North-West Himalayas to Assam and the Deccan Peninsula, at elevations up to 2120 m (3-5). The tubers contain 2-5 times higher colchicine content than seeds compared to (6). Traditionally, its seeds and tubers were used in treating gout, rheumatism, cholera, typhus, leprosy, colic and skin diseases, haemorrhoids, impotence, gonorrhoea and chronic ulcers (7-10). During the 1980s, *G. superba* faced near-extinction due to indiscriminate harvesting and continuous over-exploitation of tubers from natural habitats by local population as well as pharmaceutical companies and has been declared as an 'endangered' species by IUCN Red Data Book (5, 11-13).

Despite this, low seed germination with poor viability is also one of the factors responsible for its diminishing population.

Studying floral biology, pollination ecology and pollinators' behaviour is crucial for crop improvement, as it provides insights into floral morphology, pollination mechanisms and the adaptation of flower visitors (14, 15). The plant-pollinator relationship plays a key role in community structure and can influence the spatial distribution, species richness and abundance (16-19). The flower of G. superba is characterized by its large size, vivid and gradient tepals, claw-like shape and nectar spurs. Due to its unique floral structure, both self- and cross-pollination is challenging, requiring external agents such as biotic (insects) or abiotic (wind) factors. Previous research on G. superba has primarily focused on pollen studies, butterfly pollinators, reproductive biology, seed set, colchicine content, seed characteristics and germination ability (20-24). However, little attention has been given to the foraging behaviour of insect visitors, particularly non-butterfly pollinators, across different times of the day. Understanding floral biology, pollination mechanism, diversity of pollinators and their foraging behaviour is extremely important for management of this species for pollination. Information is lacking on nectar characteristics, pollinator diversity, foraging behaviour and pollination efficiency of *G. superba* in the Northwestern Himalayan region and across India. In this study, we aimed to assess the diversity and foraging behaviour of key insect visitors on G. superba.

#### **Materials and Methods**

The research was conducted during July-November 2021 and July-November 2022 at Medicinal and Aromatic Farm, Department of Forest Products, Dr. YSPUHF, Nauni at an altitude of 1270 meters amsl (30°51'44.7444" N, 77°10'9.1488" E) Solan, Himachal Pradesh, India. The climate is subtemperate, with an average annual rainfall of 1250 mm and average annual temperature ranges from 11.0 to 25.9 °C. The floral events were determined by randomly selecting fully opened flowers (n=10) and floral characteristics namely measurement of flower spread, sepal length, petal length, tepal length, stamen length, pistil length, number of stamens, stamen basal gap and weight of flower were recorded in the laboratory. Stigma receptivity and pollen viability were determined using acetocarmine method (23, 25). The flowers selected for nectar collection were caged a day prior to collection to prevent nectar robbing by insects. From 800 to 1000 hours, nectar was directly extracted using micropipettes from the flower base into an Eppendorf tube and stored at 15° C for further analysis (26). Changes in nectar volume and nectar sugar concentration at different floral stages, quantitative analysis of dry nectar sugars and qualitative estimation of nectar sugars were conducted using HPLC-ELSD (27, 28).

Using a standard protocol, the diversity of insect floral visitors was recorded using scan sampling and sweep net capture methods (29, 30). All the specimens were identified by using published keys (31). The foraging behaviour such as foraging rate, foraging speed, loose pollen grains of important insect pollinators was also studied and these

parameters were recorded at different day intervals throughout the flowering period. Statistical analysis was conducted using mean values for nectar parameters, yield parameters, foraging rate, foraging speed and loose pollen grains. ANOVA was used to compare mean values. A Randomized Block Design (RBD) factorial was used to analyze foraging behaviour using OPSTAT.

#### **Results and Discussion**

The flowering period commences from second fortnight of July to September with ripening of pods in the month of October. There were nine flower stages were observed in G. superba ranging from stage-1 to stage-9, which included prepollination (stage 1 to stage 5), nectar accumulation (stage 4 to stage 8) and post pollination stages (after stage 8) (Fig. 1). The scarlet red flowers were solitary in position having average floral characters viz., the pedicel length, tepal length, tepal breadth, filament length, anther length, stamen basal gap and weight of flower were recorded as  $10.32 \pm 0.14$  cm,  $6.69 \pm 0.11$  cm,  $1.45 \pm 0.05$  cm,  $4.62 \pm 0.17$  cm,  $1.26 \pm 0.03$  cm,  $0.48 \pm 0.07$  mm and  $1.72 \pm 0.04$  g, respectively. The anther dehiscence was observed at the stage when longitudinal splitting of anthers started and average pollen viability ranged from 45.79 to 88.19 % and 46.67 to 88.61 % during year 2021 and 2022, respectively and significant maximum viable pollens were recorded on the first day after anthesis and pollen viability decreased thereafter (Fig. 2). The stigma remained receptive for four days i.e., one day prior to anthesis, on the day of anthesis, one day after anthesis and two days after anthesis and average stigma receptivity ranged from 39 to 80 % maximizing on the day of anthesis and minimizing two days after anthesis (Fig. 2). Similar observations on floral characters and pollen viability have been reported (26, 32, 33).

In G. superba, nectar production began after stage 4 and peaked at stage 6, then dropped with seed set until it halted entirely at stage 9. From stage 5 to stage 8, nectar secretion ranged between 4.13  $\pm$  0.28 and 33.04  $\pm$   $\mu$ l/flower in 2021 and between 3.78  $\pm$  0.33 and 32.10  $\pm$  0.54  $\mu$ l/flower in 2022. Significantly maximum and minimum nectar accumulation was observed at stage-6 and stage-8 respectively, and after stage 8, nectar accumulation ceased. Further, the sugar concentration of nectar ranged from 52.57±0.75 to 78.04 ± 0.33 % during 2021 and 52.14  $\pm$  0.72 to 77.91  $\pm$  0.36 % during 2022. Quantitative analysis of dry nectar sugars revealed that significant maximum dry nectar sugars were present at stage 6 (94.70±0.85 μg/flower) followed by stage 7 (86.07±0.28 μg/ flower) and stage 8 (81.07±0.34 µg/flower) during 2021 and similar trend was observed during 2022 also with 94.70±0.85,  $86.07\pm0.28$  and  $81.07\pm0.34$   $\mu$ g/flower at stage 6, 7 and 8 respectively. The qualitative analysis revealed that nectar comprises of sugars namely glucose, sucrose, fructose and raffinose. The present findings on nectar volume and nectar sugar concentrations of G. superba at Nauni (Solan) are higher than the previous observations (24, 34). The results may be variable due to the dry climatic conditions of their study area.

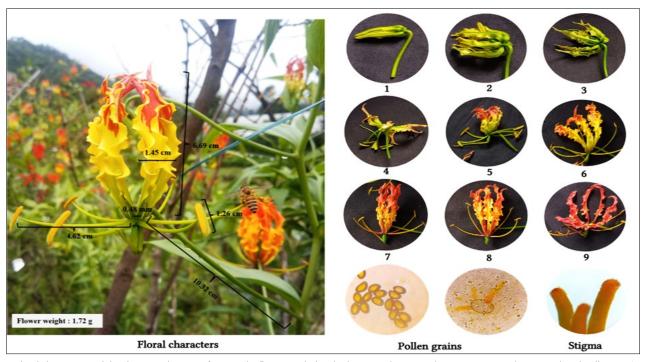


Fig. 1. Floral characters and developmental stages of G. superba flowers including bud opening (Stages 2-4), nectar secretion (Stages 4-7) and pollination/postpollination phases (Stages 8-9).

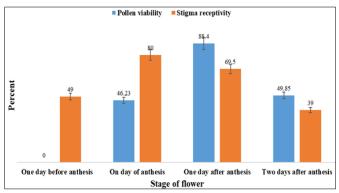


Fig. 2. Pollen viability and stigma receptivity at different floral stages in glory

The allotrophous, hemitropous and eutropous types of insect visitors were recorded on glory lily from 6 orders, 14 families and 16 genera. Among all, hymenopterans were most dominant insect visitors followed by dipterans, lepidopterans, coleopterans, orthopterans and hemipterans (Table 1, Fig. 3). Majority of the insects were reported during both years except

the insects of Vespidae and Pieridae families whose presence was not recorded during 2021 while during 2022, all insects were present except some lepidopterans such as P. bianor and E. core. The diversity indices viz., Shannon, Simpson, dominance and Margalef richness indices during 2021 and 2022 were 2.58, 0.16, 0.83 and 1.3 and 2.15, 0.22, 0.83 and 1.22 respectively. During both years, maximum relative abundance of insect visitors was recorded during morning and noon hours (800 to 1400 hours) and their abundance decreased thereafter (Table 2). Maximum B. haemorrhoidalis population was recorded during 800-1000 hours only. During 2021, no population of Vespa spp. was found visiting glory lily flowers but in 2022, considerable population of Vespa spp. was recorded. The change in incidence of wasps may occurred due to fluctuations in weather parameters.

Earlier studies of pollinator diversity in glory lily suggested that butterflies particularly from Pieridae and Papilionidae, families may act as best pollinators for glory lily due to their large size and wing pollination mechanism

Order	Family	Name of the species	2021	2022
Hymenoptera		Apis mellifera Linnaeus 1758	+	+
	Apidae	Apis cerana Fabricius 1793	+	+
		Bombus haemorrhoidalis Smith 1852	+	+
		Vespa magnifica Smith 1852	-	+
	Vespidae	Vespa velutina Lepeletier 1836	-	+
		Vespa spp.	-	+
	Encodeda.	Small ants (Lasius sp.)	+	+
	Formicidae	Large ants (Camponotus sp.)	+	+
Diptera	Syrphidae	Eristalis tenax Linnaeus 1758	+	+
	Muscidae	Musca spp.	+	+
	Calliphoridae	Calliphora sp.	+	+
	Tephritidae	Tephritidae Bactrocera zahadi Mahmood 1999		+
Coleoptera	Coccinellidae	llidae Coccinella septumpunctata Linnaeus 1758		+
	Meloidae	<i>Mylabris pustulata</i> Thunberg 1821	+	+
Lepidoptera	Pieridae	Pieris brassicae Linnaeus 1758	-	+
	Papilionidae	Papilio bianor Cramer 1777	+	-
	Nymphalidae	Euploea core Cramer 1780	+	-
Orthoptera	Pyrgomorphidae	Aularches miliaris Linnaeus 1758	+	+
Hemiptera	Pentatomidae	Erthesina fullo Thunberg 1783	+	+



Fig. 3. Diversity of insect species observed on Gloriosa superba L.

Table 2. Relative abundance of insect visitors on G. superba during 2021 and 2022

	Relative abundance of insect visitors (Number/m² in 5 minutes)								
Insects	2021				2022				
	0900h	1300h	1700h	Mean	0900h	1300h	1700h	Mean	
AC	6.53 (2.74)	4.23 (2.28)	1.20 (1.47)	3.99 (2.16)	7.10 (2.84)	4.63 (2.36)	2.77 (1.94)	4.83 (2.38)	
AM	7.70 (2.95)	5.57 (2.56)	2.03 (1.73)	5.10 (2.41)	7.23 (2.87)	5.97 (2.64)	3.17 (2.04)	5.46 (2.52)	
V	0.00 (1.00)	0.00 (1.00)	0.00 (1.00)	0.00 (1.00)	7.97 (2.99)	5.17 (2.48)	2.57 (1.89)	5.23 (2.45)	
ВН	0.67 (1.28)	0.17 (1.08)	0.00 (1.00)	0.28 (1.12)	3.53 (2.13)	0.00 (1.73)	1.47 (1.57)	1.67 (1.81)	
Α	11.60 (3.55)	4.73 (2.40)	2.60 (1.88)	6.31 (2.61)	12.13 (3.62)	7.43 (2.90)	4.03 (2.24)	7.87 (2.92)	
L	2.00 (1.72)	5.00 (2.44)	1.00 (1.38)	2.67 (1.85)	0.99 (1.14)	0.67 (1.24)	0.33 (1.14)	0.66 (1.17)	
0	3.67 (2.16)	1.33 (1.52)	3.00 (1.98)	2.67 (1.89)	2.67 (1.88)	1.67 (1.63)	2.00 (1.72)	2.11 (1.74)	
Mean	4.60 (2.21)	3.00 (1.91)	1.40 (1.49)	3.00 (1.86)	5.95 (2.51)	3.65 (2.14)	2.33 (1.79)	3.98 (2.14)	

CD  $_{(0.05)}$  (2021): Insects (0.19), Day hours (0.12), Insect $\times$  Day hours (0.33)

CD<sub>(0.05)</sub> (2022): Insects (0.19), Day hours (0.13), Insect× Day hours (0.34)

AC-Apis cerana; AM-Apis mellifera, V-Vespa sp. 1, A-ants, BH-Bombus haemorrhoidalis, L-Lepidopterans, O-other insects

Figures in the parentheses are  $\sqrt{x+1}$  transformed values

(34, 35). However, from India large sized bumble bees and sunbirds have also been reported as pollinators of G. superba (36). At Eastern Cape, South Africa butterflies (Eronia cleodora subspecies cleodora (Pieridae) and Papilio demodocus subspecies demodocus) were reported as the most common visitors to G. superba (34). In the present study, apart from insects of Pieridae and Papilionidae families, other insects from families Apidae, Vespidae and Nymphalidae were also reported as floral visitors to glory lily under Nauni (Solan) conditions during 2021 and 2022. Among the various pollinators, irrespective of day hours, B. haemorrhoidalis visited significantly maximum number of flowers followed by A. cerana, A. mellifera and P. bianor (Fig. 4) while during different day hours, irrespective of species foraging rate of all insects was significantly maximum during 1600-1800 hours (4.91 number of flowers visited per minute). In contrary, the foraging speed of all insects was significantly maximum during 800-1000 hours such as 14.79 seconds spent on one flower and irrespective of day hours, foraging speed of A. mellifera was significantly maximum followed by A. cerana, P. bianor, B. haemorrhoidalis and Vespa spp. (Fig. 4). Hence, three kinds of pollination in glory lily including melittophilus (honey bee), sphesophilus (wasps) and psychophilus (butterfly) was observed and these three insects may play an efficient role in glory lily pollination.

The effect of different pollination systems on the yield parameters of glory lily focusing on seed colchicine content along with fruit set, pod length, test weight and percentage of germination was analyzed. Maximum fruit set, pod length and test weight were obtained in hand pollination system and these yield parameters were statistically like the pollination system assisted with blower pollination (Fig. 5). However, the yield parameters, test weight and percent germination in pollination system assisted with honey bee pollination such as, 2.04  $\pm$  0.01g and 79.02  $\pm$  0.26 %, respectively were also statistically similar to hand and blower pollination treatments which represents that although bee pollination system not resulted in maximum fruit set and pod length but other crucial yield parameters like test weight and percentage germination in bee pollination were found to be significantly similar to blower and hand pollination system. About 96 %

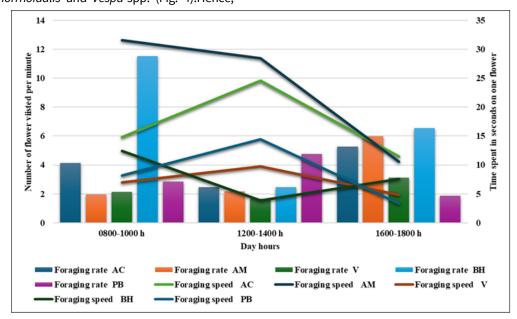


Fig. 4. Foraging behaviour viz., Foraging rate, Foraging speed and Loose pollen grains of important insect visitors on *G. superba* (\*AC- *A. cerana*; AM- *A. mellifera*, V- Vespa sp. 1, BH- B. haemorrhoidalis, PB- Papilio bianor).

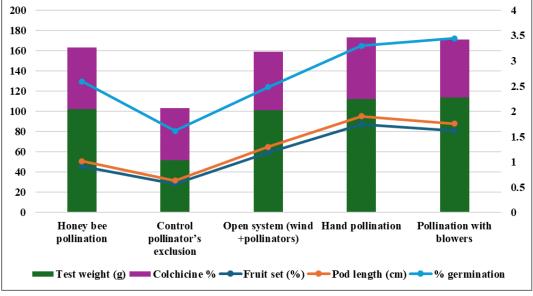


Fig. 5. Effect of different pollination systems on yield parameters of G. superba.

pod set, 8.4 cm pod length, 74 number of seeds per pod, 54 % germination were observed at Tamil Nadu, while 30 to 40 % seed set in self and hand-pollination treatments (21, 36). 80 80 % seed set in assisted pollination treatments and 66.66 % seed set in open pollination treatments (23). About 97 %, germination percentage after removal of sarcotesta along with other chemical treatments had been observed (37). In present study germination percentage after removal of sarcotesta without any other chemical treatments under laboratory ranged from 60 to 85 % which is like previous findings. The similar peaks of colchicoside and colchicine in HPLC chromatogram (Fig. 6) highlighted that the alkaloid including colchicoside and colchicine content present in whole seed of G. superba obtained from honey bee pollination and hand pollination system was similar, which showed the effectiveness of honey bee pollination for glory lily. The colchicine content in seed from assisted pollination treatments and open pollination treatment of present study are in line with results of previous research reported 1.16 % seed colchicine content in open pollination treatment (23). In present study, the effect of honey bee pollination on seed germination percentage, number of seeds per pod and test

weight of were significantly similar with hand and blower pollination treatments while the effect of honey bee pollination treatment on fruit set percentage and pod length was found to be statistically lower than hand and blower pollination treatments.

#### Conclusion

This study highlights the role of insect pollinators especially honey bees in enhancing the pollination, seed yield and colchicine content of *Gloriosa superba*. Honey bee pollination treatment was observed as best for seed germination percentage, number of seeds per pod and test weight which proved to be an effective and sustainable alternative to traditional pollination methods. Hence, it can be utilized for conservation, sustainable production and crop utilization. Integrating sustainable harvesting methods with biotechnological advancements like micropropagation and genetic improvement will help in long-term sustainability and use of this priceless medicinal plant.

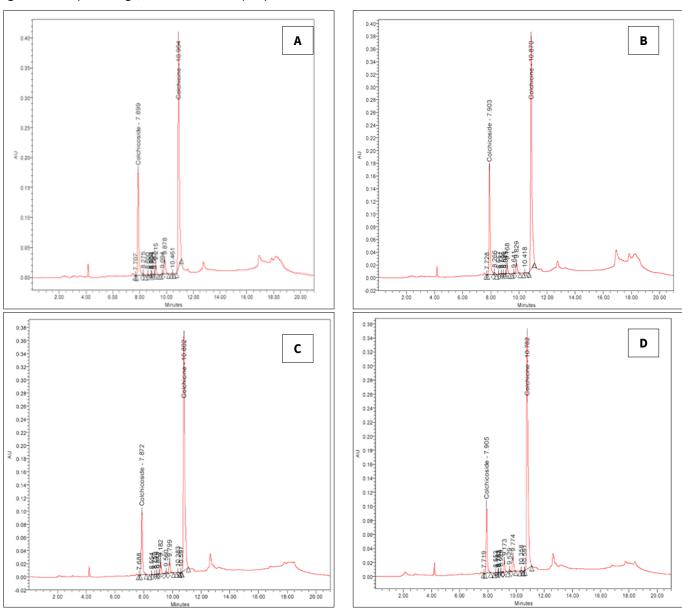


Fig. 6. HPLC chromatogram of alkaloid percentage (colchicoside and colchicine) in glory lily seeds obtained from A) Bee pollination system B) Hand pollination system C) Pollination with blower D) Open pollination.

#### **Acknowledgements**

The authors acknowledge Dr. YSPUHF, Nauni, Solan, HP, India-173230 for institutional support of this research and also thankful to the Department of Entomology and the Department of Forest Products for providing necessary facilities. The help and guidance provided by Dr. Hema Prashad and Dr. M P Singh is also acknowledged.

#### **Authors' contributions**

PR, RKT and YPS conceptualized and designed the research, PR conducted experiments, PR, RKT and MT analysed the data. PR wrote the manuscript and RKT, MT, MAW reviewed and corrected it. All authors read and approved the manuscript.

#### **Compliance with ethical standards**

Conflict of interest: None

Ethical issues: None

### References

- Satyavati GV, Raina MK, Sharma M, editors. Medicinal plants of India. New Delhi: Indian Council of Medical Research. 1976:433–34.
- Supari MR, Farooqi AA, Prasad TG. Influence of various pre-sowing treatments and growth regulators on seed germination in Gloriosa superba L. Indian J Forestry. 1993;16(2):123–26.
- Gupta BK. Production of colchicine from Gloriosa superba tubers in cultivation and utilization of medicinal plants. CSIR J. 1982:270–8.
- Ade R, Rai MK. Current advances in Gloriosa superba L. Biodiversitas. 2009;10(4):210–14. https://doi.org/10.13057/biodiv/d100409
- Sivakumar G, Krishnamurthy KV. Gloriosa superba L.: a very useful medicinal plant. In: Singh VK, Govil JN, Hashmi S, Singh G, editors. Recent progress in medicinal plants. Ethnomedicine and pharmacognosy, Part II. Texas: Series Sci Tech Pub. 2002;(7):465–82.
- Jana S, Shekhawat GS. Critical review on medicinally potent plant species: Gloriosa superba. Fitoterapia. 2011;82(3):293–301. https://doi.org/10.1016/j.fitote.2010.11.008
- Chopra RN, Nayar SL, Chopra IC. Glossary of Indian medicinal plants. New Delhi: Council of Scientific & Industrial Research. 1956:125–26.
- 8. Bellet P, Gaignault JC. *Gloriosa superba* L. and the production of colchicinic substances. Ann Pharm Fr. 1985;43(4):345–47.
- Farooqi AA, Kumarswamy BK, Bojappa KN, Pusalkar VR, Gupta R. Plantations of clinically important *Gloriosa superba*. Ind Hort. 1993;37(4):26–9.
- Lakshmi PT, Swathi S. Estimation of colchicine content in tuber, seed and leaves samples of *Gloriosa superba* using HPLC and their antibacterial studies on pathogenic strains. Int J Appl Res Nat Prod. 2015;5:34–41.
- 11. Badola HK, Pal M. Endangered medicinal plant species in Himachal Pradesh. Curr Sci. 2002;83:797–98.
- Yadav K, Aggarwal A, Singh N. Actions for ex situ conservation of Gloriosa superba L.-an endangered ornamental cum medicinal plant. J Crop Sci Biotechnol. 2012;15:297–303. https://doi.org/10.1007/s12892-012-0045-7
- 13. Padmapriya S, Rajamani K, Sathiyamurthy VA. Glory lily (*Gloriosa superba* L.): A review. Int J Curr Pharm Rev Res. 2015;7(1):43–9.

- Farooqi AA, Sreeramu BS, editors. Glory lily. In: Cultivation of medicinal and aromatic crops. Hyderabad: Universities Press Private Ltd. 2004:131–38.
- Chittka L, Raine NE. Recognition of flowers by pollinators. Curr Opin Plant Biol. 2006;9(4):428–35. https://doi.org/10.1016/ j.pbi.2006.05.002
- Wang H, Cao GX, Wang LL, Yang YP, Zhang ZQ, Duan YW. Evaluation of pollinator effectiveness based on pollen deposition and seed production in a gynodieocious alpine plant, Cyananthus delavayi. Ecol Evol. 2017;7(20):8156–60. https://doi.org/10.1002/ ece3.3391
- Kantsa A, Raguso RA, Dyer AG, Olesen JM, Tscheulin T, Petanidou T. Disentangling the role of floral sensory stimuli in pollination networks. Nat Commun. 2018;9(1):1041. https://doi.org/10.1038/ s41467-018-03448-w
- Danforth BN, Minckley RL, Neff JL. The solitary bees: Biology, evolution, conservation. Princeton: Princeton University Press. 2019. https://doi.org/10.1515/9780691189321
- Erikson E, Junker RR, Ali JG, McCartney N, Patch HM, Grozinger CM. Complex floral traits shape pollinator attraction to ornamental plants. Ann Bot. 2022;130(4):561–77. https:// doi.org/10.1093/aob/mcac082
- Mamatha M, Farooqi AA, Prasad TG. Studies on growth development and relationship between vegetative growth and yield in *Gloriosa superba* Linn. Acta Hortic. 1993;331:365–70. https://doi.org/10.17660/ActaHortic.1993.331.50
- 21. Martins DJ. Butterfly pollination of the dryland wildflower *Gloriosa minor*. J East Afr Nat Hist. 2015;103(1):25–30. https://doi.org/10.2982/028.103.0103
- 22. Selvarasu A, Kandhasamy R. Reproductive biology of *Gloriosa* superba. Open Access J Med Aromat Plants. 2012;3(2):5–11. https://epubs.icar.org.in/index.php/JMAP/article/view/22885
- 23. Dogra S. Studies on seed set and colchicine content in *Gloriosa* superba L. Master of Science [thesis]. Solan: Dr. YSPUHF. 2021.
- Poojitha M. Studies on seed characters and germination ability of glory lily (Gloriosa superba L.). Master of Science [thesis]. West Bengal: Uttar Banga Krishi Vishwavidyalaya. 2022.
- 25. Dafni A, Firmage D. Pollen viability and longevity: practical, ecological and evolutionary implications. In: Dafni A, Hesse M, Pacini E, editors. Pollen and pollination. 1st ed. Vienna: Springer. 2000:113–32. https://doi.org/10.1007/978-3-7091-6306-1\_6
- Amato B, Petit S. A review of the methods for storing floral nectars in the field. Plant Biol (Stuttg). 2017;19(4):497–503. https://doi.org/10.1111/plb.12565
- Roberts RB. Spectrophotometric analysis of sugars produced by plants and harvested by insects. J Apic Res. 1979;18:191–95. https://doi.org/10.1080/00218839.1979.11099966
- Lindqvist DN, Pedersen HÆ, Rasmussen LH. A novel technique for determination of the fructose, glucose and sucrose distribution in nectar from orchids by HPLC-ELSD. J Chromatogr B. 2018;1081– 82. https://doi.org/10.1016/j.jchromb.2018.02.019
- 29. Belavadi VV, Ganeshaiah KN. Insect pollination manual. New Delhi: Indian Council of Agricultural Research. 2013.
- Revanasidda, Belavadi VV. Floral biology and pollination in *Cucumis melo* L., a tropical andromonoecious cucurbit. J Asia-Pac Entomol. 2019;22(1):215–25. https://doi.org/10.1016/j.aspen.2019.01.001
- 31. Michener CD. The bees of the world. 2nd ed. Baltimore: The Johns Hopkins University Press. 2007.
- 32. Selvarasu A, Kandhasamy R. Floral biology and pollination behaviour of *Gloriosa rothschildiana*. J Pharmacogn Phytochem. 2019;8(2S):884–87.
- 33. Gupta LM, Raina R. Significance of sequential opening of flowers in *Gloriosa superba* L. Curr Sci. 2001;80(10):1266–67.

34. Daniels RJ, Johnson SD, Peter CI. Flower orientation in *Gloriosa* superba (Colchicaceae) promotes cross-pollination via butterfly wings. Ann Bot. 2020;125(7):1137–49. https://doi.org/10.1093/aob/mcaa048

- 35. Tercel MPTG, Veronesi F, Pope TW, Phylogenetic clustering of wingbeat frequency and flight-associated morphometrics across insect orders. Physiol Entomol. 2018;43(2):149–57. https://doi.org/10.1111/phen.12240
- 36. Venudevan B, Sundareswaran S, Vijayakumar A, Rajamani K. Studies on improving seed set and quality in glory lily through pollination methods. Madras Agric J. 2011;98:33–5. https://doi.org/10.29321/MAJ.10.100236
- 37. Le Roux LG, Robbertse PJ. Aspects relating to seed production in *Gloriosa superba* L. S Afr J Bot. 1997;63(4):191–97. https://doi.org/10.1016/S0254-6299(15)30743-2