



RESEARCH ARTICLE

Ethnobotanical survey and *in vitro* quality assessment of *Dendrobium thysiflorum* B. S. Williams from the Ultapani Forest Range, Assam, India

Sangita Das¹, Sanjib Baruah² & Arvind Kumar Goyal^{1*}

¹Department of Biotechnology, Bodoland University, Kokrajhar 783 370, Bodoland Territorial Region (BTR), Assam, India

²Department of Botany, Bodoland University, Kokrajhar 783 370, Bodoland Territorial Region (BTR), Assam, India

*Correspondence email - arvindgoyal210883@gmail.com

Received: 17 August 2025; Accepted: 22 December 2025; Available online: Version 1.0: 05 February 2026; Version 2.0: 28 February 2026

Cite this article: Das S, Baruah S, Goyal AK. Ethnobotanical survey and *in vitro* quality assessment of *Dendrobium thysiflorum* B. S. Williams from the Ultapani Forest Range, Assam, India. *Plant Science Today*. 2026; 13(1): 1-16. <https://doi.org/10.14719/pst.11303>

Abstract

Dendrobium thysiflorum B. S. Williams (Orchidaceae), locally known as 'Garudi Baha/ Khejari Baha' by the Santhal community of the Ultapani Forest Range, Bodoland Territorial Region (BTR), Assam. The species occurs anthesis between March and May, with flowers reaching full bloom and attracting pollinators. This study investigated the ethnobotanical uses, macroscopic and microscopic pharmacognostic characteristics, qualitative and quantitative phytochemical composition, heavy metal content, antioxidant activity and volatile compound profile of *D. thysiflorum*. Ethnobotanical surveys conducted among the Santhal community revealed the traditional use of leaves and pseudobulbs as a medicine for the treatment of various gynecological and parasitic ailments. The examination of powdered plant material showed satisfactory organoleptic properties (colour, aroma/odour, flavour/taste and texture) for both leaves and pseudobulbs. Microscopic analysis identified fibers in both leaves and pseudobulbs, parenchyma cells and stomata in leaves and acicular crystals and spiral vessels in pseudobulbs. Aqueous extracts of leaves and pseudobulbs underwent qualitative phytochemical screening, revealing the presence of alkaloids, carbohydrates, glycosides, terpenoids and steroids in both tissues. The heavy metal analysis indicated the absence of bismuth, cadmium and lead in the samples. Quantitative phytochemical analysis determined the total polyphenol content (TPC) to be 27.62 ± 0.03 mg gallic acid equivalents (GAE)/g dry weight in leaves and 17.57 ± 0.02 mg GAE/g dry weight in pseudobulbs. The total flavonoid content (TFC) was measured as 120.36 ± 0.05 mg quercetin equivalents (QE)/g dry weight in leaves and 87.50 ± 0.04 mg QE/g dry weight in pseudobulbs. The antioxidant activity was further evaluated using 2,2-diphenyl-1-picrylhydrazyl (DPPH) and ferric ion reducing antioxidant potential (FRAP) assays, demonstrating higher activity for DPPH assay compared to the standard. Gas chromatography-mass spectrometry (GC-MS) analysis identified 88 compounds in leaves and 59 compounds in pseudobulbs. These findings contribute to the establishment of pharmacognostic and phytochemical standards for *D. thysiflorum*, aiding in species identification, quality control and standardization of herbal formulations.

Keywords: antioxidant; *Dendrobium thysiflorum*; ethnobotany; GC-MS analysis, phytochemical analysis; traditional community healers

Introduction

Plants are one such living organism that has contributed to the human world unaccountably, starting from timber, firewood, fibers, dyes, pesticides, oils, rubber, medicines and many more. In both western and eastern cultures, as well as in highly industrialized and underdeveloped nations, there is evidence of the usage of plants for medical purposes dating back as far as 60000 years (1). Approximately 70000 plant species provide benefits to people today, yet only 7000 of those species are grown in cultivation (2). The age of discovery and conquest brought about a resurgence of interest in the therapeutic potential of plants in Western societies. For instance, the native Indians' use of plants as medicine inspired the Spaniards, who between 1531 and 1536 conquered Mexico and Peru (1).

Orchids are one of the most diverse families of angiosperms, including approximately 25000 to 30000 species classified into 750 to 800 genera (3). Recently, it was stated that there were 28000 currently identified species, 800 subspecies and approximately 763

genera (4). They are mostly grown for their gorgeous blossoms and are well-known for their economic significance rather than for their potential medical benefits (5). Orchid habitat is most abundant in India. According to the first Orchid census of India conducted and released by Botanical Survey of India (BSI) in 2019, India is fortunate to have 1256 orchid species or taxa that belong to 155 genera, 388 of which were reported to be endemic (6–8). The genus *Dendrobium* has 800-1400 species in the world (9). This genus extends from Korea and Japan across Southeast Asia, the Himalayas in the west, the Philippines in the east and Indonesia in the south, Australia, New Zealand and the Pacific Islands (10). The largest genus is *Dendrobium*, which has 58 taxa, 51 of which are monotypic genera that are found in the various regions (11).

In the Northeastern part of India, known for its abundant natural riches, the state of Assam is located at the base of the Himalayas. Due to the high temperatures, high levels of rainfall and humidity in the area, there are huge evergreen and semi-evergreen deciduous forest covers. And as a result, has developed into an ideal

habitat for the growth of many different varieties of orchids. Kokrajhar district in lower Assam has extensive, large-scale forest cover, just like the other districts in Assam. Additionally, this forest area was reported to have several kinds of terrestrial and epiphytic orchids (12, 13).

Ultapani Forest Range has a substantial amount of evergreen and semi-deciduous forest cover. Due to its outstanding biodiversity, this area is progressively growing in popularity amongst tourists. It is one of the 4 territorial ranges under Haltugaon Forest Division under Kokrajhar district. The geographical area of the Ultapani forest range is 224.64 sq. km., located from 26°39'18.1" N to 26°52'13.5" N latitude and from 90°14'04.7" E to 90°21'28.0" E longitude (14).

Dendrobium thysiflorum B. S. Williams is one such exquisite orchid found in this forest region. This species belongs to the family Orchidaceae and it referred to as "Garudi Baha" or "Khejari Baha" among the Santhal folks living in Ultapani Forest Range, BTR, Assam. It is an epiphytic orchid having aching cane stem like pseudobulb of about 5-30 × 1-2.5 cm. The phenology of the vegetative phase of *D. thysiflorum* revealed that the new shoots and leaves emerge during May to June, whereas senescence is initiated in the second year of growth, specifically from June and during November to February, no growth is observed. The plant blooms in the month of March to May.

In addition to being an ornamental plant *D. thysiflorum* is used by a group of people in Ultapani Forest Range for its medicinal properties. However, no scientific study has been done till date on *D. thysiflorum* of this region to specify its medicinal value that is being practiced by the Santhal community traditional healers. The present study was carried out to reveal the ethnobotanical studies, pharmacognostic and phytochemical data (qualitative, quantitative), heavy metal analysis, antioxidant activity and gas chromatography-mass spectrometry (GC-MS) and morphological characterization for the identification of this species of orchid, to ensure quality and purity and standardization of the herbal drug.

Materials and Methods

Plant material collection and identification

A systematic field survey was conducted in the Ultapani Forest Range, Kokrajhar, Assam, India (Global positioning system (GPS) coordinates: N26°46'21.3", E90°18'29.4"), from 2019 to 2022. Plant specimens were collected directly from their native habitat to ensure authenticity and ecological relevance (Table 1). The voucher specimens were prepared according to standard procedures (15). The preliminary identification was performed by consulting relevant literature and comparing specimens with those deposited at the Department of Botany, Bodoland University. *D. thysiflorum* was further verified and authenticated by the Botanical Survey of India (BSI), Howrah (specimen number BUSD-01, dated November 30, 2021). The taxonomic classification of *D. thysiflorum* is as follows:

Kingdom: Plantae

Class: Monocotyledons

Order: Asparagales

Family: Orchidaceae

Genus: *Dendrobium*

Species: *D. thysiflorum*

Preparation of aqueous extracts

Leaves and pseudobulbs of *D. thysiflorum* were washed, air-dried at room temperature and pulverized using an electric grinder. 5 g of each powdered samples were subjected to extraction using 50 mL of double-distilled water (1:10 w/v) for 24 hr at room temperature. The extracts were filtered through Whatman filter paper No. 1 and stored at 4 °C in airtight glass bottles.

Macroscopic examination

Macroscopic features of the plant material were documented through visual and physical examination of whole plants and individual parts against a black background (8).

Ethnobotanical survey

Ethnobotanical information was gathered through semi-structured interviews and questionnaires administered to local residents and traditional healers, focusing on the indigenous knowledge and medicinal uses of *D. thysiflorum*.

Pharmacognostic analysis

Organoleptic evaluation

Organoleptic properties (colour, aroma/odour, flavour/taste, texture) of the powdered leaves and pseudobulbs were assessed according to standard procedures (16).

Microscopic examination

Powdered leaf and pseudobulb samples were mounted in water and safranin and examined under a binocular microscope (LaboMed Vision 2000) for fragmented characteristics (17, 18).

Phytochemical screening

Qualitative tests

Aqueous extracts were subjected to qualitative tests for alkaloids, carbohydrates, flavonoids, glycosides, phenols, saponins, tannins, terpenoids, steroids and phlobatannins using established methods (19-23).

Heavy metal analysis

The presence of cadmium, bismuth and lead in the extracts was evaluated using standard protocols (24). Each heavy metal was tested using 2 independent procedures:

- **Bismuth:** Reaction with H₂S (brown precipitate) and NH₄OH (white precipitate).
- **Cadmium:** Reaction with NH₄OH (white precipitate) and potassium ferrocyanide (white precipitate).
- **Lead:** Reaction with dilute HCl (white precipitate) and dilute potassium iodide (yellow precipitate).

Table 1. Details on the parts of the orchid that are used, the vernacular name and the Global positioning system (GPS) coordinates of the collection location

Botanical name	Parts used	Santhal name	Common English name	GPS coordinates
<i>Dendrobium thysiflorum</i> B S Williams	Leaves and/or pseudobulbs	Garudi Baha or Khejari Baha	Pinecone-like Raceme Dendrobium	N26°46'21.3", E90°18'29.4"

Quantitative tests

Total polyphenol content (TPC)

TPC was determined using the Folin-Ciocalteu reagent method, with minor modifications (25). Absorbance was measured at 730 nm and the results were expressed as milligrams of gallic acid equivalents (GAE) per gram of sample in dry weight.

Total flavonoid content (TFC)

TFC was quantified using the aluminum chloride method with quercetin as a standard (26). Absorbance was measured at 510 nm and the results were expressed as milligrams of quercetin equivalents (QE) per gram of sample in dry weight.

Antioxidant activity

2, 2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity: The extracts and standard were evaluated for their antioxidant activity by measuring their ability to scavenge the stable, DPPH free radical using a standard protocol (23). A 0.006 % w/v solution of DPPH was prepared using a 95 % solvent. The extract and a 95 % freshly prepared DPPH solution were added in test tubes. Subsequently, successive dilutions ranging from 100 to 1000 µg were made in each test tube until the total volume reached 2 mL. After an incubation period of 30 min in the dark, the degree of discoloration was evaluated at a wavelength of 540 nm using a Thermo UV1 spectrophotometer (manufactured by Thermo Electron Corporation in England, UK). Measurements were conducted on a minimum of 3 occasions. Ascorbic acid served as the standard and was diluted in distilled deionized water (DDW) to create the first solution at a concentration of 1 mg/mL. To serve as a control sample, an equivalent volume was prepared without any extract and a blank was created using a solvent consisting of 95 % concentration. The equation provided was utilized to compute the percentage of DPPH-free radical scavenging.

$$S = \frac{A_0 - A_1}{A_0} \times 100$$

Where, A_0 is the absorbance of the control and A_1 is the absorbance in the presence of the sample.

In order to determine if the test chemicals reduced absorption, we compared them to the positive controls. Using the dosage inhibition curve, the IC_{50} value was determined.

Ferric ion reducing antioxidant potential (FRAP) assay: The reducing capacity of the extract was assessed following the method described by Oyaizu, with some modifications (27). The extract was mixed with different quantities ranging from 25 to 2500 µg in 1 mL of DDW. This mixture was then blended with 2.5 mL of phosphate buffer (0.2 M, pH 6.6) and 2.5 mL of potassium ferricyanide ($K_3Fe(CN)_6$) (1 %). The solution was subjected to incubation at a temperature of 50 °C for a duration of 20 min. The mixture was thereafter subjected to centrifugation at a speed of 3000 revolutions per minute for a duration of 10 min, with the addition of 2.5 mL of trichloroacetic acid (10 %). The top layer solution (2.5 mL) was combined with DDW (2.5 mL) and $FeCl_3$ (0.5 mL, 0.1 %) and the absorbance was determined at a wavelength of 700 nm. The increased absorbance of the reaction mixture showed a greater capacity for reducing substances. Ascorbic acid was used as a reference standard. Phosphate buffer (pH 6.6) was used as a control. The mean absorbance of the final reaction mixture in 2 parallel experiments was computed, together with its standard deviation.

GC-MS/MS analysis

Phytoconstituent profiling of the aqueous extracts was performed following the standard method using an Agilent 7890B GC and 7000D series Triple Quadrupole mass spectrometer, equipped with an Elite-5MS capillary column (28). The NIST 14 MS Library was used for compound identification.

Statistical analysis

All quantitative data are presented as mean \pm SEM (n=3). Statistical comparisons were performed using GraphPad Prism, version 5.0.

Results and Discussion

Macroscopic morphology

It is an epiphytic orchid, was observed to grow to a height of 20–60 cm. It exhibited clustered pseudobulbs arising from a short rhizome. The pseudobulbs were cane-like, pendent, cylindrical, stout, longitudinally ridged, olive-green to yellowish-brown and measured 5–30 \times 1–2.5 cm.

The upper portion of the pseudobulbs bore 1–4 distichous, scarcely ovate to elliptic, emerald green leaves (11.5–17 \times 3.5–6.5 cm) with obtuse, leathery textures and sheathed bases. Pendulous inflorescences (10–27 cm long) emerged laterally from the apical part of defoliated stems, bearing 4–18 white flowers (approximately 1.5–2 cm across, expanding to 3.5 cm in diameter). The dorsal sepal was white, narrowly ovate and obtuse (2–2.3 \times 1.7–1.8 cm). Lateral sepals were white, slightly oblique (2.3–2.8 \times 1.7–1.8 cm). Petals were widely elliptic to suborbicular, shortly clawed at the base and as long as the dorsal sepal, with irregularly finely denticulate margins and rounded apices. Sepals and petals displayed a light purple-violet tint dorsally at the base. The lip was bright yellow with white edges, slightly triangular to suborbicular, briefly clawed (2.2–2.7 \times 2–2.2 cm). The column and foot were yellow and the anther cap was white (Fig. 1).

Distribution: Native to the Himalayan mountains of India, Northeast, Laos, Myanmar, Burma and Thailand; rare in Vietnam and Assam.

Flowering and fruiting: March–May

Habitat: Epiphytic in semi-deciduous forests.

Status in Assam: Rare

Ecology/occurrence: Epiphytic in semi-deciduous forests. New shoots and leaves emerge during May–June, with senescence occurring in the second year of growth, specifically from June and a period of no growth observed from November–February. GPS coordinates: N26°46'21.3", E90°18'29.4".

Specimen examined: Assam: Kokrajhar, Ultrapani forest range, Collection no: 01, 13-03-2020, *Sangita Das*, herbarium information: submitted to CAL, herbarium accession no.: BUSD-01.

Ethnobotanical survey

Ethnobotanical data collected through interviews and questionnaires with Santhal community traditional healers revealed the use of *D. thyrsiflorum* leaves and pseudobulbs for various medicinal purposes (Table 2). Preparations involving *D. thyrsiflorum* with or without *Hibiscus rosa-sinensis* L. and *Shorea robusta* C F Gaertn. were used to treat menstrual problems, abnormal white discharge, otalgia, otorrhea and intestinal worms.



Fig. 1. *Dendrobium thysiflorum*. **a)** Habit and whole plant, **b)** Full grown inflorescence **c)** Emerald green leaves, **d)** Pseudobulb **e)** Immature inflorescence **f)** Whole flower **g)** Dissected flower **h. I)** Sepals; **h. II)** Dorsal sepal; **h. III)** Lateral sepal, **i. I)** Petals; **i. II)** Petals as long as dorsal sepal **j)** Lip.

Table 2. Ethnobotanical information collected from Santhal community traditional healers

Scientific name	Parts used	Other ingredient/ supplementary ingredients	Medicinal uses	Processing	Dosage	Herbalist	Preventive measure
<i>Dendrobium thysiflorum</i> B S Williams		<i>Hibiscus rosa-sinensis</i> L. flowers (20%)	Tonic or tablets to treat menstrual problem viz. irregular period; proper blood flow and pre-menopausal issues related to uterine bleeding	Juiced or crushed and made into tablets	2 tablespoon/ 2 tablets per day	TH	Intake of egg, fish, meat or any other allergic food items is prohibited during medication.
	Leaves and/or Pseudobulbs	Bark of <i>Shorea robusta</i> C.F.Gaertn. (10 g) and water (10 L)	Treats abnormal white discharge/ leukorrhoea	Decoction	1 cup of tonic per day	TH	
		NR	Recovers otalgia	Juiced	2 drops per day	TH, BM	
		NR	Recovers otorrhea	Juiced	2 drops per day	TH	
		NR	Treat worms in intestine	Juiced	2 drops per day	BM	

Herbalists: Mr. Tibru Hembram (TH), Mr. Basu Murmu (BM)

Pharmacognostic analysis

Organoleptic evaluation

Organoleptic analysis of pulverized leaves and pseudobulbs revealed distinct characteristics. Leaves exhibited a granular texture, paddy smell, tastelessness and olive-green colour. Pseudobulbs displayed a spongy granular texture, sweet aroma, tastelessness and wheat-brown colour (Table 3).

Microscopic study

Microscopic analysis of the pulverized plant parts revealed distinct anatomical features in *D. thyrsiflorum*. Fibers were observed in both the leaves and pseudobulbs. Parenchyma cells and stomata were exclusively present in the leaves, while acicular crystals and spiral vessels were identified in the pseudobulbs (Fig. 2).

Parenchyma cells, fundamental to plant metabolism, are involved in various processes, including cellular repair, photosynthesis, gas exchange, energy storage and waste elimination (29, 30). Stomata facilitate gaseous exchange and fibers provide mechanical support within plant tissues (31, 16). Acicular crystals contribute to elasticity and offer support against crushing pressure (32, 33). Spiral vessels, the smallest components facilitating plant movement, play a crucial role in the development of reproductive organs (34, 35).

Table 3. Organoleptic parameters of pulverized plant parts

Parameters	Leaves of <i>D. thyrsiflorum</i>	Pseudobulbs of <i>D. thyrsiflorum</i>
Texture	Granular	Spongy granular
Aroma	Paddy smell	Sweet
Taste	Tasteless	Tasteless
Colour	Olive green	Wheat brown

Qualitative tests

Qualitative phytochemical screening of aqueous extracts from pulverized *D. thyrsiflorum* plant parts revealed distinct compositional differences between leaves and pseudobulbs (Table 4). Both leaves and pseudobulbs exhibited the presence of alkaloids, carbohydrates, glycosides, terpenoids and steroids. However, flavonoids, phenols, saponins, tannins and phlobatannins were detected exclusively in the leaves.

The observed phytochemical profile suggests potential therapeutic applications, aligning with established literature. Alkaloids, for instance, are known for their diverse pharmacological activities, including anti-allergic, anti-inflammatory, antimicrobial, anticonvulsant and anticancer effects (36–39). Carbohydrates, particularly reducing sugars, contribute to energy provision and may offer cardioprotective benefits (40–42). Flavonoids, found only in the leaves, are recognized for their antioxidant properties, mitigating oxidative damage (43–45). Glycosides exhibit a broad spectrum of therapeutic potential, including analgesic, anti-inflammatory, cardiogenic, antimicrobial, antiviral and anticancer activities (46). Phenols act as antioxidants and possess anti-inflammatory properties, potentially beneficial in conditions like inflammatory bowel disease, rheumatoid arthritis and skin disorders (47). Saponins have been associated with lipid-lowering, cholesterol-reducing and hypoglycemic effects, along with potential applications in hypercalciuria, lead poisoning prevention, dental caries reduction and antiplatelet aggregation (48). Tannins display a range of bioactivities, including anti-inflammatory, antiviral, antioxidant, antimicrobial and anticancer properties and are also implicated in anti-diabetic, anti-wrinkle, anti-cardiac and

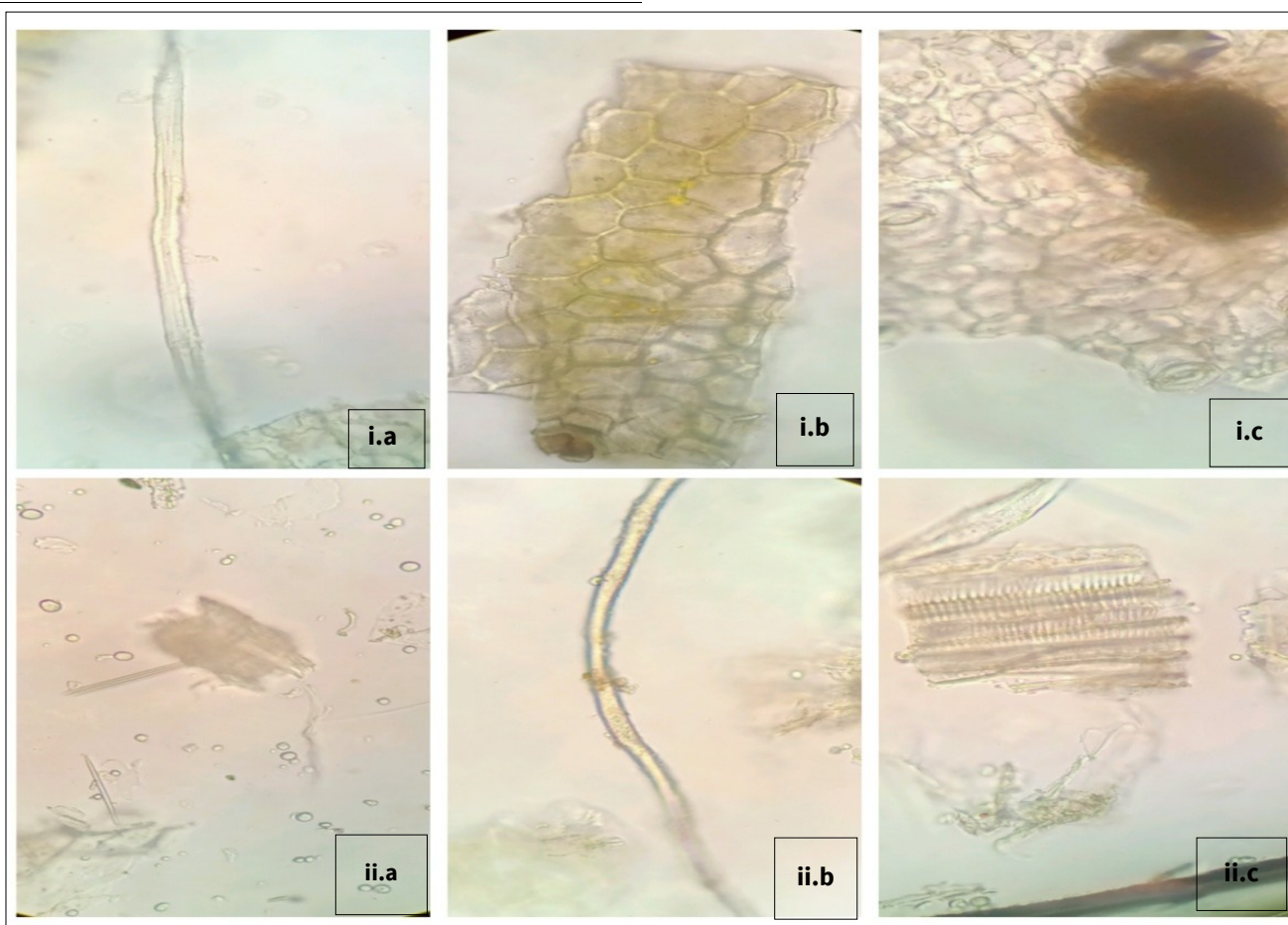


Fig. 2. Microscopy of powdered *D. thyrsiflorum*. (i) Leaves (40X): a) Fibre; b) Parenchyma; c) Stomata; (ii) Pseudobulbs (40X): a) Acicular crystal; b) Fibre; c) Spiral vessel.

Table 4. Phytochemical screening of leaves and pseudobulbs of *D. thyriflorum* aqueous extract (Qualitative tests)

Constituent	Chemical test	Leaves of <i>D. thyriflorum</i>	Pseudobulbs of <i>D. thyriflorum</i>
Alkaloids	Mayer's reagent test	+	+
	Wagner's reagent test	+	+
Carbohydrates	Molisch's test	+	+
	Fehling's test	+	+
Flavonoids	Shinoda test	+	-
	Pew's test	+	-
	Alkaline reagent test	+	-
Glycosides	Keller-kiliani test	+	+
	Ellagic acid test	+	-
Phenols	Ferric chloride test	+	-
	Foam test	+	-
Tannins	Ferric chloride test	+	-
Terpenoids	Bromine water test	+	-
	Salkowski test	+	+
Steroids	Salkowski test	+	+
Phlobatannins	Hydrochloric acid test	+	-

“+” sign and “-” sign indicate the presence and absence of constituent in the sample respectively

anti-diarrheal applications (49). Terpenoids demonstrate anticancer, anti-inflammatory, antimicrobial, antiviral, antimalarial, immunomodulatory, neuroprotective and anti-allergic activities (50). Steroids, essential for various physiological processes, play critical roles in development, growth, energy metabolism, homeostasis and reproduction (51). Finally, phlobatannins, present only in the leaves, have been reported to possess diuretic properties (52).

Heavy metal analysis

Heavy metal analysis indicated the absence of lead, cadmium and bismuth in both leaves and pseudobulbs, suggesting the safety of these plant parts for medicinal use (Table 5).

Table 5. Heavy metal test for *D. thyriflorum* leaves and pseudobulbs aqueous extract

Experiment	Observation	Results
Test for Bismuth		
H ₂ S gas + sample solution	No dark brown precipitate	Bismuth absent
NH ₄ OH + sample solution	No white precipitate	Bismuth absent
Test for Cadmium		
NH ₄ OH + sample solution	No white precipitate	Cadmium absent
Potassium ferrocyanide + sample solution	No white precipitate	Cadmium absent
Test for Lead		
Dilute HCl + sample solution	No white precipitate	Lead absent
KI + sample solution.	No yellow precipitate	Lead absent

Quantitative tests

Determination of total polyphenol content (TPC)

Polyphenolic compounds are recognized contributors to antioxidative activity. The TPC of *D. thyriflorum* leaves and pseudobulbs was determined to be 27.62 ± 0.03 mg gallic acid equivalents (GAE) per gram dry weight (mg/g) and 17.57 ± 0.02 mg GAE/g dry weight, respectively.

Research on *D. thyriflorum* is rare, but several studies have been reported on *Dendrobium* species. For instance, an ethanol extracts of *D. crepidatum* exhibited a TPC of 78.11 ± 0.72 µg GAE/mg extract (53), another study revealed that a TPC of 54.47 ± 0.12 mg GAE/g DW was found in the methanolic stem extract of *in vitro*-raised

D. nobile, while only 3.25 ± 0.20 mg GAE/g DW was found in the chloroform leaf extract of the same species, highlighting the impact of solvent and plant part on phenolic yield (54).

Determination of total flavonoid content (TFC)

The TFC of aqueous extracts from *D. thyriflorum* leaves and pseudobulbs was measured as 120.36 ± 0.05 mg QE per gram dry weight (mg/g) and 87.50 ± 0.04 mg QE/g dry weight, respectively.

Compared to other studies, direct comparative study was rare, however, examining a few *Dendrobium* species reveals that significant amounts of flavonoids were found in *D. crepidatum* extracts, with the hexane fraction having TFC of 82.62 ± 1.13 µg QE/mg (53). Another study found that the TFC of *D. heterocarpum* tissue-culture regenerates was 38.38 mg QE per gram dry weight (mg QE/g DW), whereas young leaves contained 27.27 mg QE/g DW (55).

Determination of antioxidant activity via DPPH scavenging assay

The antioxidant activity of *D. thyriflorum* leaves and pseudobulbs aqueous extracts were evaluated using the DPPH radical scavenging assay, providing an assessment of reducing power and total antioxidant capacity. The extracts demonstrated significant antioxidant effectiveness when compared to ascorbic acid, the reference standard. The DPPH assay is based on the principle that antioxidants can reduce the stable free radical DPPH, resulting in a loss of its characteristic deep purple colour and a corresponding decrease in absorbance at 540 nm. This change in absorbance is quantitatively proportional to the antioxidant capacity. The comparative antioxidant activity of the extracts and ascorbic acid across varying concentrations, demonstrating a gradual increase in activity with increasing concentration (Fig. 3). The half-maximal inhibition concentrations (IC₅₀) for *D. thyriflorum* leaves and pseudobulbs were determined to be 10.22 and 6.98, respectively.

In other studies, it was found that, at 800 µg/mL, the ethanol stem-extract of *D. crepidatum* scavenged DPPH free radicals with approximately 94.69 % activity, with an IC₅₀ value of approximately 73.90 µg/mL (53).

FRAP assay

The reducing capabilities of the *D. thyriflorum* leaf and pseudobulb extracts compared to ascorbic acid standard (Fig. 4). The reducing power, which is measured by the conversion of Fe³⁺ to Fe²⁺, increased with rising extract concentration for both leaf and pseudobulb

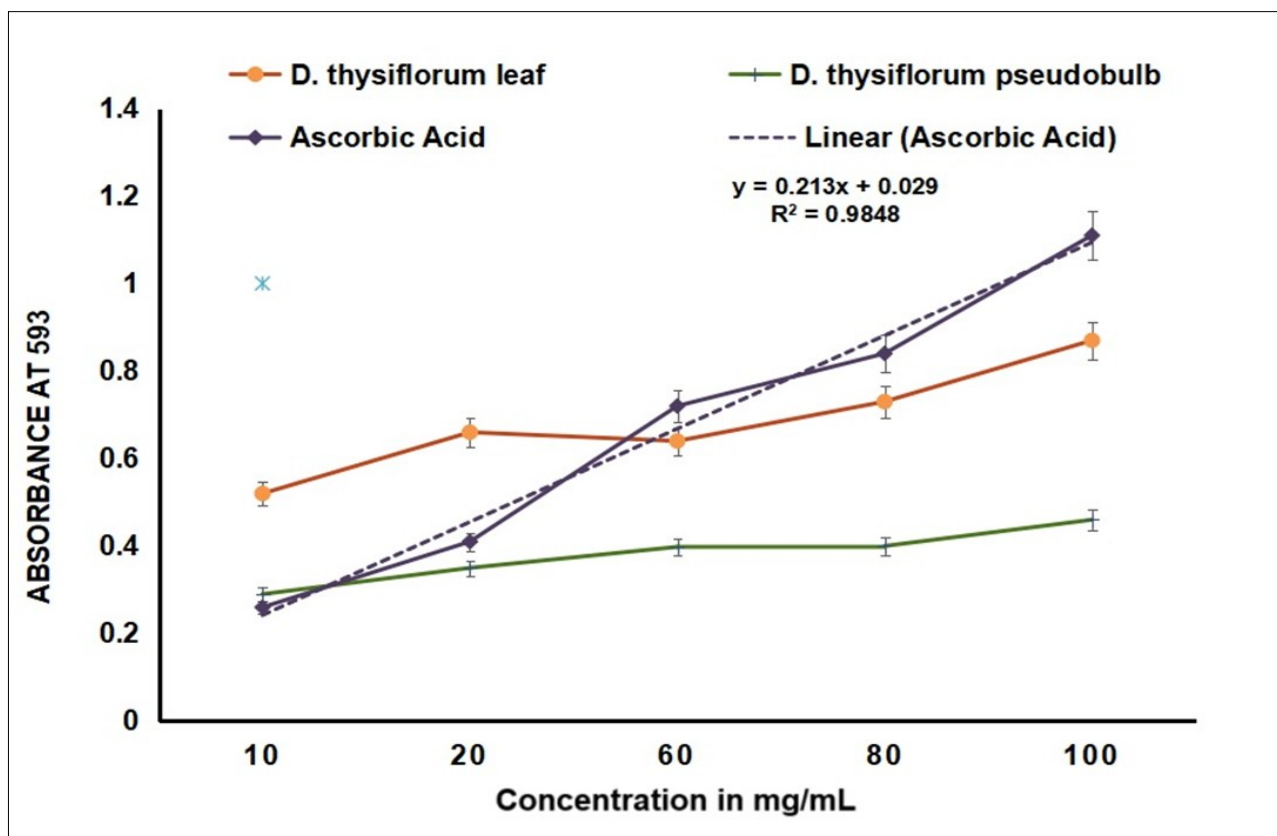


Fig. 3. DPPH Scavenging activity (n=3; mean±SE) of the leaves and pseudobulbs of *D. thysiflorum*.

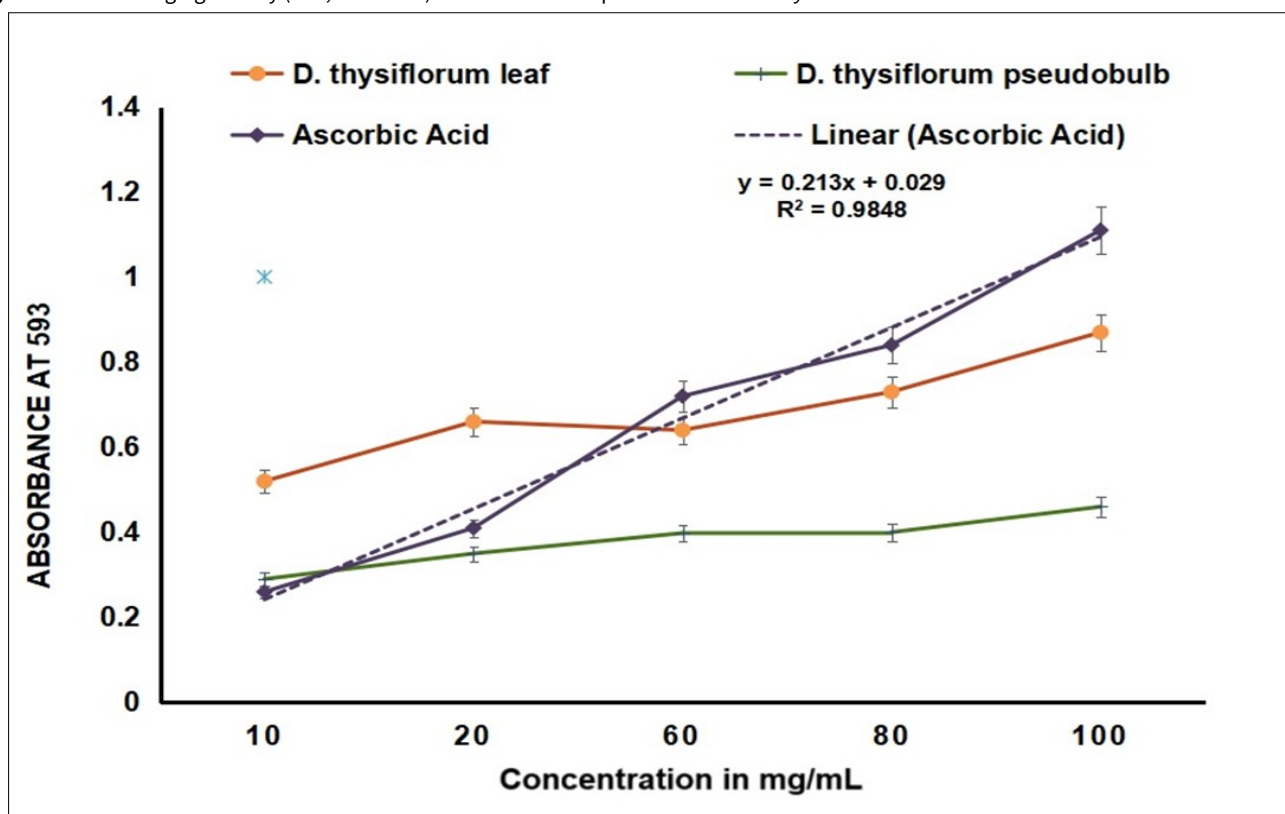


Fig. 4. Ferric reducing-antioxidant power (FRAP) assay (n=3; mean±SE) of the aqueous extract of the leaves and pseudobulbs of *D. thysiflorum* in comparison with a standard.

samples.

Specifically, the leaf extract exhibited a higher reducing power than the ascorbic acid standard at lower concentration (10 mg/mL and 20 mg/mL). However, at 60 µg/mL, the reducing power of the leaf extract plateaued or decreased relative to ascorbic acid. Conversely, the pseudobulb extract exhibited consistently

lower reducing power than ascorbic acid across all the tested concentrations.

Regarding other species, earlier research on the ethanolic stem, leaf and flower extracts of *D. sulcatum* showed FRAP value of 0.50 ± 0.04 mmol Fe^{2+} /g extract, 0.35 ± 0.05 and 0.12 ± 0.02 , respectively (56).

GC-MS/MS profiling

GC-MS analysis was performed on the leaves and pseudobulbs of *D. thysiflorum*. The resulting chromatogram is presented in Fig. 5 and Fig. 6 respectively and the identified 88 and 59 phytochemicals of each sample are summarized in Table 6 and Table 7 respectively.

An ethnobotanical survey among the Santhal community revealed the traditional use of *D. thysiflorum* leaves and pseudobulbs for treating various ailments. To validate these traditional claims, a scientific investigation was conducted involving qualitative and quantitative phytochemical analyses. Qualitative tests confirmed the presence of diverse compounds in both pulverized plant parts. Subsequently, GC-MS analysis identified 88 compounds in the leaves and 59 in the pseudobulbs. A comparative analysis was performed, correlating the identified compounds with existing literature. Secondary sources reported that several identified compounds, including hexylene glycol, cyclotrisiloxane (hexamethyl-), 1H-indole, 2-methoxy-4-vinyl-phenol, benzaldehyde (3-hydroxy-4-methoxy-), phenol (3,4-dimethoxy-), 2,4-Di-tert-butylphenol, n-hexadecanoic acid, scoparone, hexadecanoic acid (2-hydroxy-1-(hydroxymethyl)ethyl ester), acetic acid, 2(5H)-furanone, 2,5-Dimethyl-4-hydroxy-3(2H)-furanone and phenol (2-methoxy-), possess anti-infectious properties, including antimicrobial, antibacterial, antiviral, anti-inflammatory and antifungal activities (58, 59, 61–63, 65–72, 74–76, 78, 79, 85–88). The presence of these compounds in *D. thysiflorum* leaves and pseudobulbs may explain the plant's efficacy in treating ear ailments, such as otalgia and otorrhea, as well as obesity, as reported by the Santhal traditional healers.

Furthermore, cyclotrisiloxane (hexamethyl-), benzaldehyde (3-hydroxy-4-methoxy-), 2,4-Di-tert-butylphenol, 4-ethenyl-2,6-

dimethoxy-phenol, n-Hexadecanoic acid, scoparone, hexadecanoic acid (2-hydroxy-1-(hydroxymethyl)ethyl ester), phenol (2-methoxy-), 2,4-Di-tert-butylphenol and hexadecanoic acid (methyl ester) have been documented to possess antioxidant activities (60, 62, 63, 67–69, 71–76, 78, 88, 89). Consequently, the presence of these compounds in the extracts may contribute to the observed DPPH radical scavenging activity.

Conclusion

Our investigation has documented the traditional use of *D. thysiflorum* by Santhal healers residing within the Ultapani forest range for the treatment of diverse ailments. To establish a foundation for its medicinal applications, a comprehensive pharmacognostic analysis of the leaves and pseudobulbs of *D. thysiflorum* was conducted, encompassing macroscopic, microscopic, organoleptic, phytochemical and heavy metal assessments. The leaves and pseudobulbs exhibited significant antioxidant activity, as evidenced by their ability to scavenge DPPH radicals and reduce ferric ions, comparable to standard antioxidants (ascorbic acid, quercetin and gallic acid). Furthermore, substantial quantities of total polyphenols and flavonoids, known for their antioxidant properties, were detected. While many potential antioxidant and therapeutic compounds were identified, further isolation, purification and targeted bioassays are required to pinpoint the specific molecules responsible for the observed effects. GC-MS profiling revealed the presence of 88 and 59 compounds in the leaves and pseudobulbs respectively, suggesting a rich source of natural antioxidants. Importantly, the absence of detectable heavy metals further supports the potential therapeutic utility of this

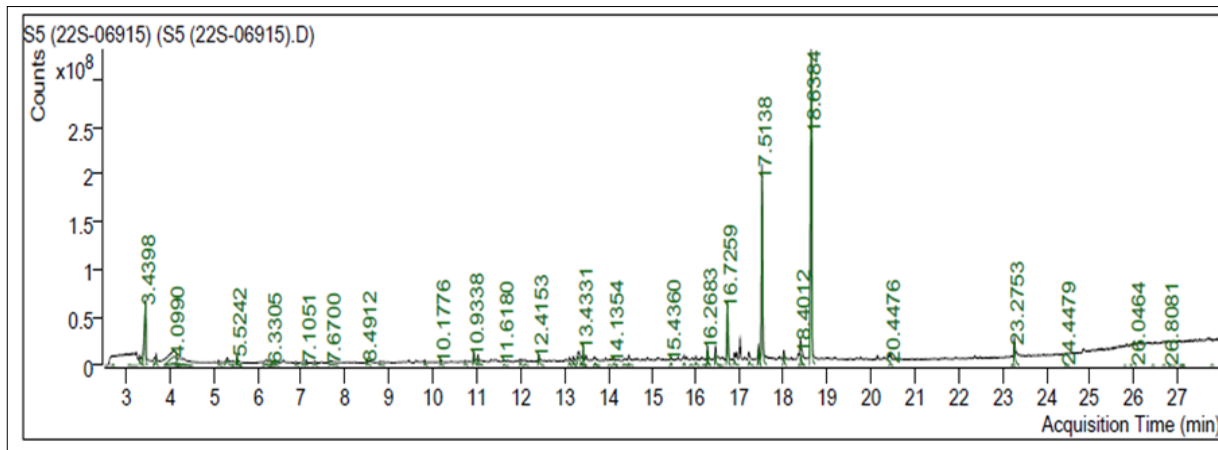


Fig. 5. Gas chromatography-mass spectroscopy (GC-MS) analysis of leaves of *Dendrobium thysiflorum* B. S. Williams.

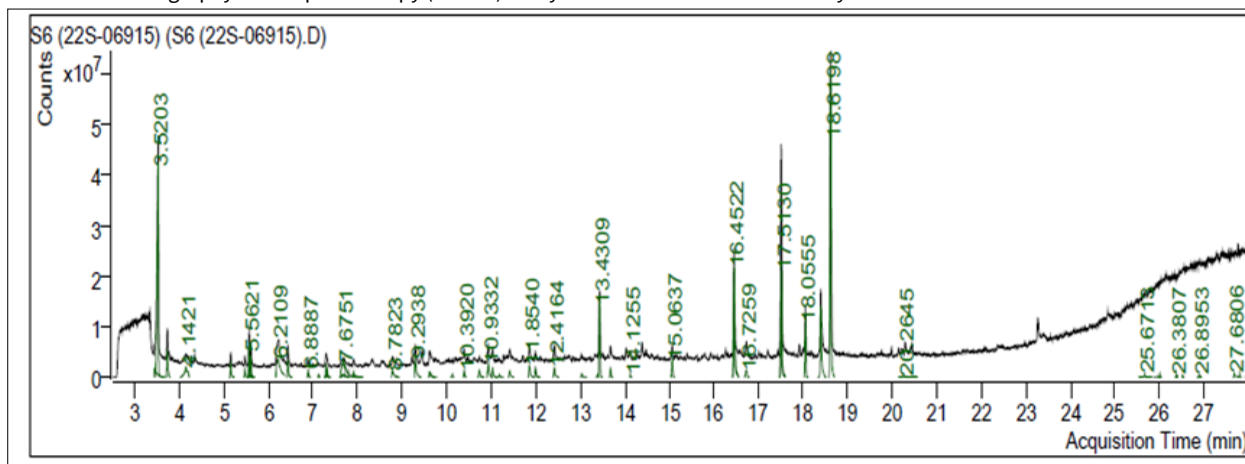


Fig. 6. Gas chromatography-mass spectroscopy (GC-MS) analysis of pseudobulbs of *D. thysiflorum* B. S. Williams.

Table 6. Results of GC-MS for Leaves of *Dendrobium thyrsiflorum* B. S. Williams

Component RT	Compound name	Formula	Area	Medicinal value	References
2.6887	2-(ethylthio)-5,6,7,8-tetrahydro-benzothio(2,3-d)(1,3) oxazin-4-one (1)	C ₁₂ H ₁₃ NO ₂ S ₂	685475	NR	
3.0673	trans-(carveylbenzyl) oxide	C ₁₇ H ₂₂ O	1734248	NR	
3.3060	Methylamine, N, N-dimethyl-	C ₃ H ₉ N	11359790	NR	
3.4398	Methyl formate	C ₂ H ₄ O ₂	149700260	NR	
3.6365	methyl-(3R)-(-)-3-ethyl-5-oxopentanoate	C ₈ H ₁₄ O ₃	3152372	NR	
3.6805	Propanoic acid	C ₃ H ₆ O ₂	13476842	1. Lowers fatty acids content in liver and plasma 2. Reduces food intake 3. Exerts immunosuppressive actions 4. Improves tissue insulin sensitivity	(57)
4.0990	6-oxabicyclo (3.1.0) hexane-2,4-diol	C ₅ H ₈ O ₃	102422183	NR	
4.0994	Butanoyl(tert-butyl)dimethyl silane	C ₁₀ H ₂₂ OSi	25082251	NR	
4.2100	1,1-dimethyl-4-tert-butylsemicarbazide	C ₇ H ₁₇ N ₃ O	9008720	NR	
4.2201	((1R,4S,5R)-5-((E)-5-(tertButyldimethylsilyloxy)-3-methyl-3-pentenyl)-1-methyl-4-(2-methyl (1,3)dioxolan-2-yl)-2-cyclopentenyl) methanol	C ₂₃ H ₄₂ O ₄ Si	6323813	NR	
4.2766	(1R,2R,9R,10R,13S)-(E)-7-Benzenesulfonyl-1,5,9-trimethyl-13-(2-methyl (1,3)dioxolan-2-yl)-4-oxatricyclo (10.3.0.0(3,5)) pentadeca-8,14-diene	C ₂₇ H ₃₆ O ₅ S	3482635	NR	
4.3010	Propanediamide, 2-amino-	C ₃ H ₇ N ₃ O ₂	8819330	NR	
5.1045	(1R,2R)-1-Methanol-2-acetonitrile-4-cyclohexene	C ₉ H ₁₃ NO	3552669	NR	
5.3039	2-Pentenoic acid, 4-methyl-	C ₆ H ₁₀ O ₂	19464380	NR	
5.4315	Methyl 3-hydroxy-2-(p-toluenesulfonyloxy)methyl-10-tetrahydropyran-1-yl decanoate	C ₂₄ H ₃₈ O ₈ S	1354302	NR	
5.5242	Hexylene glycol	C ₆ H ₁₄ O ₂	11737899	1. Used as topical corticosteroids 2. Has antimicrobial properties	(58, 59)
5.5242	2-Methyl-2,4-pentanediol	C ₆ H ₁₄ O ₂	11741559	NR	
5.5656	(Thiomethyl)(4'-nitrophenyl)((N-methyl)bromopyrrolidinyl)isothioacylamidrazone	C ₁₄ H ₁₉ BrN ₄ O ₂ S	1680393	NR	
6.1692	Propanedioic-1,3-13C2 acid	C ₃ H ₄ O ₄	5690219	NR	
6.2051	(4R,5R)-3-Methylene-4-vinyl-5-n-pentyl-gamma-butyrolactone	C ₁₂ H ₁₈ O ₂	4514457	NR	
6.3305	2-Butenoic acid, 2,3-dimethyl-	C ₆ H ₁₀ O ₂	5284152	NR	
6.3312	Ethane-1,1-d2, 2,2,2-trifluoro-	C ₂ H ₂ F ₃	880713	NR	
6.4058	Propane, 2-methyl-	C ₄ H ₁₀	3655378	NR	
6.8701	2-Cyclopenten-1-one, 2-hydroxy-3-methyl-	C ₆ H ₈ O ₂	1805620	NR	
7.0311	1-Phenylpropane-1,2-diol	C ₉ H ₁₂ O ₂	1574299	NR	
7.1051	3-hydroxy-4,4-dimethyl-2-hydroxyfuran-2-one	C ₆ H ₁₀ O ₃	3732451	NR	
7.3011	Cyclotrisiloxane, hexamethyl-	C ₆ H ₁₈ O ₃ Si ₃	3683998	1. It has antioxidant, antibacterial, antidiabetic and antimicrobial activities 2. It is essential for scavenging free radicals	(60–64)
7.6700	(+)-3(5)-((methyl (3'-pinanylmethyl) amino) methyl) pyrazole	C ₁₆ H ₂₇ N ₃	6658720	NR	
8.4912	L-Homoserine lactone, N, N-dimethyl-	C ₆ H ₁₁ NO ₂	8516387	NR	
8.7855	2-Hydroxy-1-phenyl-3-(phenylsulfonyl)propan-1-one	C ₁₅ H ₁₄ O ₄ S	3134002	NR	
9.8129	1H-Pyrrole-2,5-dione, 3-ethyl-4-methyl-	C ₇ H ₉ NO ₂	3789491	NR	
10.1776	(5R,6R)-6-((3aR,5S,6aS)-2,3,3a,4,5,6a-hexahydrofuro(2,3-b)furan-5-yl)-5,6-dimethyl-1-cyclohex-2-enone	C ₁₄ H ₂₀ O ₃	4464988	NR	

10.7326	1H-Indole	C ₈ H ₇ N	4331171	1. It help to maintain the human intestine's biological barrier 2. Has anti-inflammatory activities 3. Used to treat inflammatory bowel disease, hemorrhagic colitis and colorectal cancer 4. Used to treat diabetes mellitus, inflammation in the central nervous system and vascular regulation	(65)
10.9338	2-methoxy-4-vinyl-phenol	C ₉ H ₁₀ O ₂	20375623	1. Possess anti-inflammatory qualities 2. Has anti-cancer qualities in relation to pancreatic cancer cell lines, Panc-1 as well as SNU-213	(66)
11.0269	1,3-Benzodioxol-5-ol	C ₇ H ₆ O ₃	17865555	NR	
11.6180	Phenol, 2-methoxy-5-(1-propenyl)-, (E)-	C ₁₀ H ₁₂ O ₂	2029140	NR	
11.9943	2H-1-Benzopyran-2-one, 3,4-dihydro-	C ₉ H ₈ O ₂	5864258	NR	
12.1088	Benzaldehyde, 3-hydroxy-4-methoxy-	C ₈ H ₈ O ₃	2980149	1. They act as plant metabolite 2. Used as an anti-inflammatory, antioxidant and anticonvulsant agent	(67-69)
12.4153	Phenol, 3,4-dimethoxy-	C ₈ H ₁₀ O ₃	17328838	1. It exhibits induction of apoptosis for cancer chemoprevention, cardio protection 2. Photoprotection of the skin 3. It has anti-inflammatory and anti-atherogenic activity	(70)
13.1197	N-(3,4-dimethoxyphenyl) hydroxylamine	C ₈ H ₁₁ NO ₃	5387306	NR	
13.2096	3-Hydroxy-4-methoxyacetophenone	C ₉ H ₁₀ O ₃	7882834	NR	
13.3290	beta-D-Glucopyranose, 1,6-anhydro-	C ₆ H ₁₀ O ₅	38533797	NR	
13.3770	(3R*,5S*,6R*)-3-Isopropyl-5-mesitylamino-6-methyl-6-phenyl-1,2,4-trioxane	C ₂₂ H ₂₉ NO ₃	3348702	NR	
13.4331	2,4-Di-tert-butylphenol	C ₁₄ H ₂₂ O	22999644	1. It has anti-inflammatory, antioxidant and antifungal properties 2. Neurodegenerative ailments are treated using it as a neuroprotective substance	(71, 72)
13.7014	1,3,3a,4,5,6,7,8,9,9a-Decahydro-4,9-epoxybenzo(f)isobenzouran-1,3-dione	C ₁₂ H ₁₂ O ₄	1790109	NR	
13.7066	2-methyl-1-(1,3,5-trimethyl-4-pyrazolyl)-1-propanone	C ₁₀ H ₁₆ N ₂ O	4545777	NR	
14.0282	5,6,7,8-Tetrahydro-7,7-dimethyl-6,8-methanoisquinolin-3-amine	C ₁₂ H ₁₆ N ₂	892878	NR	
14.1354	4-ethenyl-2,6-dimethoxy-phenol	C ₁₀ H ₁₂ O ₃	4874989	1. It has antioxidative and antimutagenic properties	(73)
14.1815	(1E)-2-Methylcyclohexanone oxime	C ₇ H ₁₃ NO	1237260	NR	
14.3622	Ethanone, 1-(1a,2,3,5,6a,6b-hexahydro-3,3,6-atrimethyloxireno (g) benzofuran-5-yl)-	C ₁₃ H ₁₈ O ₃	2284944	NR	
14.4253	Ethanone, 1-(1a,2,3,5,6a,6b-hexahydro-3,3,6-atrimethyloxireno (g) benzofuran-5-yl)-	C ₁₃ H ₁₈ O ₃	1420409	NR	
14.4776	Ethyl 2-(1-(N-Ethoxycarbonylamino)-4-tbutylcyclohexyl) prop-2-enoate isomer	C ₁₈ H ₃₁ NO ₄	4756548	NR	
15.4360	2-Cyclohexen-1-one, 3,5,5-trimethyl-4-(3-oxobutyl)-	C ₁₃ H ₂₀ O ₂	4495369	NR	
15.7331	trans-4,5-dihydro-4,4-dimethyl-2-(6-methyl-5-(phenylmethyl)-1,3-cyclohexadien-1-yl) oxazole	C ₁₉ H ₂₃ NO	4247809	NR	
15.8967	(E)-3-(7,7-difluoro-5-bicyclo(4.1.0) hepta-1,3,5-trienyl)-2-propenoic acid ethyl ester	C ₁₂ H ₁₀ F ₂ O ₂	1835194	NR	
16.0034	2-Propanone, 1-hydroxy-3-(4-hydroxy-3-methoxyphenyl)-	C ₁₀ H ₁₂ O ₄	6822476	NR	
16.1778	1H-Cyclopenta(1,3) cyclopropa(1,2) benzen 6(7H)-one, 3a-(acetyloxy) hexahydro-3b methyl-, (3a.alpha.,3b.alpha.,7aR*)-	C ₁₃ H ₁₈ O ₃	1533535	NR	
16.2683	6-Hydroxy-4,4,7a-trimethyl-5,6,7,7-atetrahydrobenzofuran-2(4H)-one	C ₁₁ H ₁₆ O ₃	30232075	NR	
16.2711	1-Oxaspiro (5.5) undecan-8-one, (+.-)-	C ₁₀ H ₁₆ O ₂	10094768	NR	
16.4517	8-Methoxytricyclo (6.2.1.0(1,5)) undec-5-en-7-one	C ₁₂ H ₁₆ O ₂	22279273	NR	
16.5366	2-(4-Nitrophenyl)-2-(4-methylphenoxy)-3,3-	C ₁₈ H ₁₉ NO ₄	931654	NR	
16.7259	2-Cyclohexen-1-one, 4-hydroxy-3,5,5-trimethyl 4-(3-oxo-1-butenyl)-	C ₁₃ H ₁₈ O ₃	105272345	NR	
16.8842	(E)-2-acetyl-3-octenoic acid methyl ester	C ₁₁ H ₁₈ O ₃	13132926	NR	
17.2217	3-(3,4-Dimethoxyphenyl) propane-1,2-diol	C ₁₁ H ₁₆ O ₄	9184606	NR	
17.4325	Acetic acid, 10,11-dihydroxy-3,7,11-trimethyldodeca-2,6-dienyl ester	C ₁₇ H ₃₀ O ₄	25644430	NR	
17.5138	(1,3) dioxolo(4,5-g) chromen-6-one	C ₁₀ H ₆ O ₄	374812281	NR	
18.0131	7-Oxabicyclo (4.1.0) heptan-3-ol, 6-(3-hydroxy1-butenyl)-1,5,5-trimethyl	C ₁₃ H ₂₂ O ₃	20447079	NR	

18.4012	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	27694138	1. It has antioxidant and antibacterial properties	(74)
18.4285	1,1-dipropyl-1-germacyclohexan-4-one	C ₁₁ H ₂₂ GeO	4110577	NR	
18.6384	Scoparone	C ₁₁ H ₁₀ O ₄	667413188	1. It is a potent and useful ingredient in the herbal treatment <i>Artemisia capillaris</i> Thunb, which has been used to treat cholestasis, jaundice and hepatic dysfunction 2. It has anti-inflammatory, antioxidant, anti-apoptotic, anti-fibrotic activities and hypolipidemic effects 3. Used to treat cholestatic liver disease (CLD), a broad category of hepatobiliary disorders with varied aetiologies that share common characteristics such as periductal fibrosis, ductal reactivity and inflammatory response 4. Used to treat erectile dysfunction (ED)	(75-77)
20.4476	5-Iodo-2-methylbenzofuran	C ₉ H ₇ IO	4628379	NR	
23.2251	2-Acetyl-3-cyano-2,3-dimethylcyclobutane-1-carboxylic acid	C ₁₁ H ₁₅ NO ₃	879412	NR	
23.2753	Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl) ethyl ester	C ₁₉ H ₃₈ O ₄	48315450	1. Has antioxidant, anti-inflammatory and anthelmintic properties	(78)
24.4479	cis-6a,7a,8,9,10,11a,11b-Octahydro-4-methoxybenzo(4,5) furo (2,3-c)(1) benzopyran-6-one	C ₁₆ H ₁₆ O ₄	1765024	NR	
24.4481	4-Methyl-9-phenyl-1-nitro-3,8,10-triaza-7-oxatricyclo(4.3.1.0(4,10)dodeca-2,8-diene	C ₁₃ H ₁₂ N ₄ O ₃	1449610	NR	
25.7993	2H-Pyran-6-carboxylic acid, 3-((4-(4-heptylcyclohexyl) benzoyl) oxy)-3,4-dihydro-2-methyl-4-oxo-, methyl ester, (2S-(2.alpha.,3.beta.(trans)))-	C ₂₈ H ₃₈ O ₆	263173	NR	
25.9210	3-(p-Ethoxyphenyl)-4-(5-(o-chlorophenyl)-.delta.(2)-1,2,4-oxadiazol-3-yl) sydnone	C ₁₈ H ₁₃ ClN ₄ O ₄	756472	NR	
26.0464	2-(4-Bromophenyl)-5,6-dihydropyrazolo (5,1-a) isoquinoline	C ₁₇ H ₁₃ BrN ₂	465197	NR	
26.4389	tri(propan-2-yl)-((4-(2-tri (propan-2-yl) silylethynyl) phenyl) methoxy) silane	C ₂₇ H ₄₈ OSi ₂	568402	NR	
26.6966	2,8-Bis(dimethylamino)-4,6-bis(propylamino)-5-phenyl-5H-thiopyrano(2,3-d:6,5-d') dipyrimidine	C ₂₅ H ₃₄ N ₈ S	727727	NR	
26.8081	3-Methyl-5(Z)-(E)-(3-phenylprop-2-ynylidene)-4-(prop-1-en-2-yl)-5,6-dihydropyran-2-one	C ₁₈ H ₁₆ O ₂	473464	NR	
26.8489	4-Formyl-3-(p-nitrophenyl) sydnone	C ₉ H ₅ N ₃ O ₅	843677	NR	
27.0983	2-(2'-Nitro-1-(4"-methoxyphenyl) ethyl)-benzofuran	C ₁₇ H ₁₅ NO ₄	761705	NR	
27.1005	N-Methyl-4'-methoxy benzene sulfena NRide-d5	C ₁₄ H ₁₀ D ₅ NOS	892805	NR	
27.1456	5-(di(propan-2-yloxy) phosphoryl-(methylthio)methyl)-2,4-diphenylthiazole	C ₂₃ H ₂₈ NO ₃ PS ₂	411226	NR	
27.7938	Ethyl 3-methyl-4-triphenylphosphoranylidene-4,5-dihydropyrazol-5-one-1-carboxylate	C ₂₅ H ₂₃ N ₂ O ₃ P	742480	NR	
27.9895	3-(N-(2-Hydroxycarbonylmethyl)-carboxamide)-4-methyl-9H-carbazole	C ₁₆ H ₁₄ N ₂ O ₃	524240	NR	
27.9895	(E)-beta-(2-Hydroxyphenyl ethylene) benzeneethanol-D2	C ₁₅ H ₁₂ D ₂ O ₂	666738	NR	

*NR= Not Reported

Table 7. Results of GC-MS for pseudobulbs of *Dendrobium thyrsiflorum* B S Williams

Component RT	Compound name	Formula	Area	Medicinal value	References
3.4407	Formamide, N-(2-methylpropyl)-	C ₅ H ₁₁ NO	7168491	NR	
3.5203	Acetic acid	C ₂ H ₄ O ₂	84691845	1. Used to treat cancer, diabetes, obesity and cardiovascular diseases 2. Used to treat microbial infections 3. Used to treat hypertension, hyperlipidemia 4. Enhance glycemic control	(79-84)
3.7251	2-Propanone, 1-hydroxy-	C ₃ H ₆ O ₂	8487824	NR	
4.1421	6-oxabicyclo (3.1.0) hexane-2,4-diol	C ₅ H ₈ O ₃	9587146	NR	
5.1476	6-Methylhexahydrocycloprop(a)pentalen-3a,6-diol	C ₁₀ H ₁₆ O ₂	4182521	NR	
5.4677	2(5H)-Furanone	C ₄ H ₄ O ₂	2617535	1. Has the potential to stop different harmful bacteria from forming biofilms	(85, 86)
5.5621	2-Methyl-2,4-pentenediol	C ₆ H ₁₄ O ₂	7146689	NR	
5.5649	Hexane, 2,3-dimethyl-	C ₈ H ₁₈	2678126	NR	
5.5650	Acetic acid, mercapto-, 3-methylbutyl ester	C ₇ H ₁₄ O ₂ S	2525333	NR	
5.6013	2-Cyclopenten-1-one, 2-hydroxy-	C ₅ H ₆ O ₂	6466340	NR	
6.2109	Methane, nitro-	CH ₃ NO ₂	22037565	NR	
6.4311	2-Hydroxy-gamma-butyrolactone	C ₄ H ₆ O ₃	5638605	NR	
6.8887	2-Cyclopenten-1-one, 2-hydroxy-3-methyl-	C ₆ H ₈ O ₂	2169166	NR	
7.1240	4,4-Dideuteriomethoxy cyclohexane	C ₇ H ₁₂ D ₂ O	929875	NR	
7.2902	2,5-Dimethyl-4-hydroxy-3(2H)-furanone	C ₆ H ₈ O ₃	4077703	1. Has antimicrobial properties 2. Possess the capacity to act as an anti-infective agent against microbial infections in humans	(87)
7.3106	Cyclotrisiloxane, hexamethyl-	C ₆ H ₁₈ O ₃ Si ₃	2956558	1. It has antioxidant, antibacterial, antidiabetic and antimicrobial activities 2. It is essential for scavenging free radicals	(60-64)
7.3123	2-chloranyl-7-methoxy-3-methyl-quinoline	C ₁₁ H ₁₀ ClNO	2412151	NR	
7.6598	Benzene, ((fluorophenylmethyl)sulfinyl)-	C ₁₃ H ₁₁ FOS	3444637	NR	
7.6751	Phenol, 2-methoxy-	C ₇ H ₈ O ₂	24525290	1. Possesses the capacity to treat cardiovascular, metabolic and cancerous disorders 2. These compounds assist avoid vascular issues because of their antioxidant and anti-inflammatory properties. 3. Improve general health	(88)

7.7072	Cyclopentadeca-1,2-diene	C ₁₅ H ₂₆	2245463	NR
7.9062	1,3-Propanediamine	C ₃ H ₁₀ N ₂	2118238	NR
8.7823	O-(1-Carboxyethyl) benzaldoxime	C ₁₀ H ₁₁ NO ₃	4487880	NR
9.2938	Catechol	C ₆ H ₆ O ₂	9569857	NR
9.6144	2-(N-ethyl) imino-3-pentanone	C ₇ H ₁₃ NO	3211743	NR
9.6193	4,7-Ethanoisobenzofuran-1,3-dione, 3a,4,7,7atetrahydro-4-methyl-7-(1-methylethyl)- , (3a.alpha.,4.alpha.,7.beta.,7a.alpha.)-	C ₁₄ H ₁₈ O ₃	2788478	NR
10.1285	1,4-Cyclohexadiene, 1,3,3-trimethyl-6-(2,4,4-trimethyl-2,5-cyclohexadien-1-ylidene)-	C ₁₈ H ₂₄	1052527	NR
10.3920	2-(Deuteriomethylene)bornane	C ₁₁ H ₁₆ D ₂	2262264	NR
10.7280	m-Aminophenylacetylene	C ₈ H ₇ N	3200991	NR
10.9332	Ethanone, 1-(2-hydroxy-5-methylphenyl)-	C ₉ H ₁₀ O ₂	6531238	NR
11.0283	4-Ethyl-5-methoxypyridazine	C ₇ H ₁₀ N ₂ O	3517340	NR
11.1806	1-hydroxy-4,4-dimethyl-3-phenyl-2-pentanone	C ₁₃ H ₁₈ O ₂	1071920	NR
11.4129	2-methoxy-3-methyl-1,4-benzenediol	C ₈ H ₁₀ O ₃	2799822	NR
11.8540	1-(Acetoxymethyl)-3-isopropyl-2,2-	C ₁₂ H ₂₂ O ₂	5554845	NR
11.9914	3,4-Dihydro-1H-isochromen-1-one	C ₉ H ₈ O ₂	4065648	NR
12.4164	1-(5-fluoranyl-2-oxidanyl-phenyl) ethanone	C ₈ H ₇ FO ₂	4420441	NR
13.0322	8-fluoranyl-3-methyl-isochromen-1-one	C ₁₀ H ₇ FO ₂	1461593	NR
13.3868	(3R*,5S*,6R*)-3-Isopropyl-5-mesitylamino-6-methyl-6-phenyl-1,2,4-trioxane	C ₂₂ H ₂₉ NO ₃	919705	NR
13.4309	2,4-Di-tert-butylphenol	C ₁₄ H ₂₂ O	21829246	1. It has anti-inflammatory, antioxidant and antifungal properties 2. Neurodegenerative ailments are treated using it as a neuroprotective substance (71, 72)
13.6787	Benzoic acid, 4-ethoxy-, ethyl ester	C ₁₁ H ₁₄ O ₃	2752578	NR
14.1255	2-Cyclohexen-1-ol, 2-(phenylethynyl)-, acetate	C ₁₆ H ₁₆ O ₂	700660	NR
15.0637	1(2H)-Naphthalenone, 3,4-dihydro-6-methoxy-2,2-dimethyl	C ₁₃ H ₁₆ O ₂	5659907	NR
16.4522	2H-1-benzopyran-2-one, 3,4-dihydro-6-hydroxy-5,7-dimethyl	C ₁₁ H ₁₂ O ₃	42803690	NR
16.4525	Isopropyl N,P-diisopropylphosphonamidate	C ₉ H ₂₂ NO ₂ P	28528457	NR
16.7259	(4E)-6-Methylhepta-4,6-dien-2-one	C ₈ H ₁₂ O	3473638	NR
17.5041	1,3-Dimethyl-2-oxo-1,2-dihydro-5-lambda.(5)-pyrido(1,2-a) pyrimidin-5-ylum-4-olate	C ₁₀ H ₁₀ N ₂ O ₂	27022210	NR
17.5130	(+)-Malbranicin	C ₁₁ H ₁₂ O ₄	38652771	NR
18.0555	Hexadecanoic acid, methyl ester	C ₁₇ H ₃₄ O ₂	16408615	1. Has antioxidant and anticancer activities 2. Has the potential to treat human colon cancer cells (89)
18.3984	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	29153153	1. Has antioxidant and antibacterial properties (74)
18.6198	Scoparone	C ₁₁ H ₁₀ O ₄	104989115	1. It is a potent and useful ingredient in the herbal treatment Artemisia capillaris Thunb, which has been used to treat cholestasis, jaundice and hepatic dysfunction 2. It has anti-inflammatory, antioxidant, anti-apoptotic, anti-fibrotic activities and hypolipidemic effects 3. Used to treat cholestatic liver disease (CLD), a broad category of hepatobiliary disorders with varied aetiologies that share common characteristics such as periductal fibrosis, ductal reactivity and inflammatory response 4. Used to treat erectile dysfunction (ED) (75-77)
20.2645	syn-(Ra*,1'S*)-N,N-Diethyl-2-(1'-ethoxybut-3'-enyl)-1-naphthamide	C ₂₁ H ₂₇ NO ₂	3643238	
25.6713	i-Propyl-(1,3-diphenylprop-2-ynyl)ether	C ₁₈ H ₁₈ O	2232809	
25.9768	6-((9'-Anthracenyl) methyl) (ethyl) aminomethyl)-2-pyridinamine	C ₂₃ H ₂₃ N ₃	535801	
26.0239	3,4-Dimethyl-11-(3,4-di(carbomethoxy)-5-(cyclohexylimino)dihydrofuran-2-yl) indenoquinoxaline	C ₃₀ H ₂₉ N ₃ O ₅	367496	
26.3807	1-methyl-3,4-bis(trifluoromethylthio)pyrazole	C ₆ H ₄ F ₆ N ₂ S ₂	624033	
26.5163	(2Z)-2-((6E,7E)-1-keto-6-(2-keto-2-methoxyethylidene)-7-trimethylsilyl-3,4-dihydro-2H-2-benzazecin-5-ylidene) acetic acid methyl ester	C ₂₂ H ₂₇ N ₅ O ₅ Si	282399	
26.8953	4,12,20-Trideoxyphorbol-13-(2',3'-dimethyl)-butyrate	C ₂₆ H ₃₈ O ₄	865622	
27.6806	5,11,13,14-Tetracarboxymethoxytetracyclo(7.3.2.0(2,8).0(10,11)) tetradeca-2(8),3,6,13-tetraene	C ₂₂ H ₂₂ O ₈	1562368	
27.8083	2-ethyl-3-ferrocenyl-N-phenylpyrrole	C ₂₂ H ₂₁ FeN	1029494	
27.9891	(E)-Ethyl 2-benzyl-3-(4'-phenylphenyl) but-2-enoate	C ₂₅ H ₂₄ O ₂	685251	

*NR= Not reported

species. Notably, several compounds identified through GC-MS have been previously reported to possess activities relevant to the treatment of various disorders, including diabetes, cardiovascular diseases, cancer, cholestatic liver disease, obesity and erectile dysfunction. We propose that further investigation into the untapped therapeutic potential of *D. thyrsoiflorum* is warranted, aiming to facilitate the development and discovery of valuable herbal products.

Acknowledgements

All the authors are thankful to the Higher Education Department, Government of Assam for financial assistance vide letter no. AHE.493/2017/110 under the scheme “Tejasvi Navadhitamastu Edu Infra Fund: Astadash Mutukar Unnoyonee Mala” and Department of Biotechnology, Ministry of Science and Technology, Government of India for the project grant vide letter No. BT/IN/Indo-US/Foldscope/39/2015 under the scheme “Proposal for use of Foldscope as a research tool”.

Authors' contributions

AKG conceptualized and designed the study. SD, SB, AKG carried out the research work and acquired the data. SD and AKG analyzed the data and wrote the first draft of the manuscript. All authors read and approved the final manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interests to declare.

Ethical issues: None

References

- Gossell-Williams M, Simon RE, West ME. The past and present use of plants for medicines. *West Indian Med J.* 2006;55(4):217–18.
- Selvi S, Polat R, Çakılcıoğlu U, Celep F, Dirmenci T, Ertuğ ZF. An ethnobotanical review on medicinal plants of the Lamiaceae family in Turkey. *Turk J Bot.* 2022;46(4):283–332. <https://doi.org/10.55730/1300-008X.2712>
- Dressler RL. *Phylogeny and classification of the orchid family.* Cambridge: Cambridge University Press; 1993.
- Biswas S, Singh D. *A manual on orchid education.* Pakyong: ICAR–National Research Centre for Orchids; 2019.
- Singh S, Singh AK, Kumar S, Kumar M, Pandey PK, Singh MC. Medicinal properties and uses of orchids: a concise review. *Elixir Appl Bot.* 2012;52:11627–34.
- Singh SK, Agrawala DK, Jalal JS, Dash SS, Mao AA, Singh P. *Orchids of India: a pictorial guide.* Kolkata: Botanical Survey of India; 2019.
- Sharma A, Pathak P. The budding potential of orchids in the cosmeceutical sector: role of orchids in skincare and health. *J Orchid Soc India.* 2020;34:79–85.
- Das S, Baruah S, Goyal AK. *In vitro* studies on quality assessment and ethnobotany of *Acampe rigida* (Buch.-Ham. ex Sm.) PF Hunt encountered in Ultapani Forest Range, Assam. *Plant Sci Today.* 2022;9(sp2):24–29. <https://doi.org/10.14719/pst.1720>
- Xiaohua J, Singchi C, Yibo L. Taxonomic revision of *Dendrobium monifolium* complex (Orchidaceae). *Sci Hortic.* 2009;120(1):143–45. <https://doi.org/10.1016/j.scienta.2008.10.002>
- Moudi M, Go R, Yien CYS, Saleh MN. A review on molecular systematic of the genus *Dendrobium* Sw. *Acta Biol Malaysiana.* 2013;2(2):71–78. <http://doi.org/10.7593/abm/2.2.71>
- Gogoi K, Das R, Yonzon R. Orchids of Assam, North East India: an annotated checklist. *Int J Pharm Life Sci.* 2015;6(1):1–10.
- Basumatary N, Sarma CM. Epiphytic orchid flora of Chirang Reserve Forest. *J Phytol Res.* 2004;17(1):33–37.
- Basumatary S, Baruah S, Singh LJ. Two new additions to the orchid flora of Assam, India. *J Threat Taxa.* 2021;13(11):19665–70.
- Das S, Baruah S, Goyal AK. Reduction in orchid diversity in Ultapani Forest range of Kokrajhar District of Assam. *J Emerg Technol Innov Res.* 2021;8(5):g677–g684.
- Jain SK, Rao RR, editors. *Field and herbarium methods.* New Delhi: Today and Tomorrow Publishers; 1977.
- Aslam I, Iqbal J, Peerzada S, Afridi MS, Ishtiaq S. Microscopic investigations and pharmacognostic techniques for the standardization of *Caralluma edulis* (Edgew.) Benth. ex Hook. f. *Microsc Res Tech.* 2019;82(11):1891–902. <https://doi.org/10.1002/jemt.23357>
- Evans WC. *Trease and Evans' pharmacognosy.* 16th ed. Saunders Elsevier; 2009.
- Khandelwal KR. *Practical pharmacognosy: techniques and experiments.* 25th ed. Nirali Prakashan; 2017.
- Trease GE, Evans WC. *Pharmacognosy.* 11th ed. London: Macmillian Publishers; 1989. p. 10–15.
- Harborne AJ. *Phytochemical methods: a guide to modern techniques of plant analysis.* Dordrecht: Springer Science & Business Media; 1998. p. 49–188.
- Kolawole OM. Studies on the efficacy of *Bridelia ferruginea* Benth. bark extract in reducing the coliform load and BOD of domestic wastewater. *Ethnobot Leaflet.* 2006;10:228–38.
- Edeoga HO, Okwu DE, Mbaebie BO. Phytochemical constituents of some Nigerian medicinal plants. *Afr J Biotechnol.* 2005;4(7):685–88. <https://doi.org/10.5897/AJB2005.000-3127>
- Goyal AK, Middha SK, Sen A. Evaluation of the DPPH radical scavenging activity, total phenols and antioxidant activities in Indian wild *Bambusa vulgaris* “Vittata” methanolic leaf extract. *J Nat Pharm.* 2010;1(1):40–45. <https://doi.org/10.4103/2229-5119.73586>
- Ranjith D. Fluorescence analysis and extractive values of herbal formulations used for wound healing activity in animals. *J Med Plants Stud.* 2018;6(2):189–92.
- Singleton VL, Rossi JA. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *Am J Enol Vitic.* 1965;16(3):144–58.
- Zhishen J, Mengcheng T, Jianming W. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chem.* 1999;64(4):555–59.
- Oyaizu M. Studies on products of browning reaction: antioxidative activities of products of browning reaction prepared from glucosamine. *Jpn J Nutr Diet.* 1986;44(6):307–15.
- Usha T, Middha SK, Shanmugarajan D, Babu D, Goyal AK, Yusufoglu HS, et al. Gas chromatography–mass spectrometry metabolic profiling, molecular simulation and dynamics of diverse phytochemicals of *Punica granatum* L. leaves against estrogen receptor. *Front Biosci (Landmark Ed).* 2021;26(9):423–41. <https://doi.org/10.52586/4957>
- Pruyn ML. *Parenchyma.* eLS. 2001;1–8. <https://doi.org/10.1002/9780470015902.a0002083.pub2>
- Carlquist S. Living cells in wood 3. Overview; functional anatomy of the parenchyma network. *Bot Rev.* 2018;84:242–94. <https://doi.org/10.1007/s12229-018-9198-5>
- Mukhi S, Bose A, Panda P, Rao MM. Pharmacognostic, physicochemical and chromatographic characterization of Samasharkara Churna. *J Ayurveda Integr Med.* 2016;7(2):88–99.

- <https://doi.org/10.1016/j.jaim.2015.11.004>
32. Schneider A. The probable function of calcium oxalate crystals in plants. *Bot Gaz.* 1901;32(2):142–44.
 33. Bhagat PA, Bhuktar AS. Diversity of mineral crystals in various medicinal plants. *Bioinfollet Q J Life Sci.* 2017;14(4b):448–53.
 34. Leon P. The homoiomerics of Anaxagoras. *Class Q.* 1927;21(3–4):133–41.
 35. Karabelas AJ, Kostoglou M, Koutsou CP. Modeling of spiral wound membrane desalination modules and plants—review and research priorities. *Desalination.* 2015;356:165–86. <https://doi.org/10.1016/j.desal.2014.10.002>
 36. Hussain G, Rasul A, Anwar H, Aziz N, Razzaq A, Wei W, et al. Role of plant derived alkaloids and their mechanism in neurodegenerative disorders. *Int J Biol Sci.* 2018;14(3):341–57. <https://doi.org/10.7150/ijbs.23247>
 37. Lee S, Kim DC, Baek HY, Lee KD, Kim YC, Oh H. Antineuroinflammatory effects of tryptanthrin from *Polygonum tinctorium* Lour. in lipopolysaccharide-stimulated BV2 microglial cells. *Arch Pharm Res.* 2018;41(4):419–30. <https://doi.org/10.1007/s12272-018-1020-8>
 38. Kurek J, editor. Introductory chapter: alkaloids—their importance in nature and for human life. In: *Alkaloids—Their Importance in Nature and Human Life.* London: IntechOpen; 2019. <https://doi.org/10.5772/intechopen.85400>
 39. Adamski Z, Blythe LL, Milella L, Bufo SA. Biological activities of alkaloids: from toxicology to pharmacology. *Toxins.* 2020;12(4):210. <https://doi.org/10.3390/toxins12040210>
 40. Mann J. Dietary carbohydrate: relationship to cardiovascular disease and disorders of carbohydrate metabolism. *Eur J Clin Nutr.* 2007;61(1):S100–S111. <https://doi.org/10.1038/sj.ejcn.1602940>
 41. Maureen Z, Beth S, editors. The functions of carbohydrates in the body. In: *An introduction to nutrition.* 2012. p. 165–232.
 42. Kanter M. High-quality carbohydrates and physical performance: expert panel report. *Nutr Today.* 2018;53(1):35–39. <https://doi.org/10.1097/NT.0000000000000238>
 43. Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. *Sci World J.* 2013;2013:162750. <https://doi.org/10.1155/2013/162750>
 44. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. *J Nutr Sci.* 2016;5:e47. <https://doi.org/10.1017/jns.2016.41>
 45. Ruiz-Cruz S, Chaparro-Hernández S, Hernández-Ruiz KL, Cira-Chávez LA, Estrada-Alvarado MI, Gassos Ortega LE, et al. Flavonoids: important biocompounds in food. In: *Flavonoids—from biosynthesis to human health.* InTech; 2017. p. 353–69. <https://doi.org/10.5772/67864>
 46. Soto-Blanco B. Herbal glycosides in healthcare. In: *Herbal biomolecules in healthcare applications.* Academic Press; 2022. p. 239–82. <https://doi.org/10.1016/B978-0-323-85852-6.00021-4>
 47. Rahman MM, Rahman MS, Islam MR, Rahman F, Mithi FM, Alqahtani T, et al. Role of phenolic compounds in human disease: current knowledge and future prospects. *Molecules.* 2021;27(1):233. <https://doi.org/10.3390/molecules27010233>
 48. Shi J, Arunasalam K, Yeung D, Kakuda Y, Mittal G, Jiang Y. Saponins from edible legumes: chemistry, processing and health benefits. *J Med Food.* 2004;7(1):67–78. <https://doi.org/10.1089/109662004322984734>
 49. Pizzi A. Tannins medical/pharmacological and related applications: a critical review. *Sustain Chem Pharm.* 2021;22:100481. <https://doi.org/10.1016/j.scp.2021.100481>
 50. Yang W, Chen X, Li Y, Guo S, Wang Z, Yu X. Advances in pharmacological activities of terpenoids. *Nat Prod Commun.* 2020;15(3):1–13. <https://doi.org/10.1177/1934578X20903555>
 51. Adhya D, Annuario E, Lancaster MA, Price J, Baron-Cohen S, Srivastava DP. Understanding the role of steroids in typical and atypical brain development: advantages of using a “brain in a dish” approach. *J Neuroendocrinol.* 2018;30(2):e12547. <https://doi.org/10.1111/jne.12547>
 52. Awoyinka OA, Balogun IO, Ogunnowo AA. Phytochemical screening and *in vitro* bioactivity of *Cnidioscolus aconitifolius* (Euphorbiaceae). *J Med Plants Res.* 2007;1(3):63–65.
 53. Paudel MR, Chand MB, Pant B, Pant B. Assessment of antioxidant and cytotoxic activities of extracts of *Dendrobium crepidatum*. *Biomolecules.* 2019;9(9):478. <https://doi.org/10.3390/biom9090478>
 54. Chakraborty S, Mitra A, Dey P. Phytochemical profiling and antioxidant properties of *in vitro*-raised *Dendrobium nobile* Lindl. *Ind Crops Prod.* 2020;145:112099. <https://doi.org/10.1016/j.indcrop.2019.112099>
 55. Longchar TB, Deb CR. Comparative analysis of nutraceutical potential phytochemicals and antioxidant activities in different parts of wild and *in vitro* regenerated plantlets of *Dendrobium heterocarpum* Wall. ex Lindl.: a medicinal orchid. *J Pharmacogn Phytochem.* 2021;10(4):331–36. <https://doi.org/10.22271/phyto.2021.v10.i4d.14169>
 56. Rungsang T, Srivilai J, Rakasawapokin P, Mungmai L, Saesue K, Aoonboontum P, et al. Assessment of antioxidant, anti-lipid peroxidation, antiglycation, anti-inflammatory and anti-tyrosinase properties of *Dendrobium sulcatum* Lindl. *Cosmetics.* 2023;10(2):43. <https://doi.org/10.3390/cosmetics10020043>
 57. Sa’ad H, Peppelenbosch MP, Roelofsen H, Vonk RJ, Venema K. Biological effects of propionic acid in humans; metabolism, potential applications and underlying mechanisms. *BBA Mol Cell Biol Lipids.* 2010;1801(11):1175–83. <https://doi.org/10.1016/j.bbalip.2010.07.007>
 58. Kinnunen T, Hannuksela M. Skin reactions to hexylene glycol. *Contact Dermatitis.* 1989;21(3):154–58. <https://doi.org/10.1111/j.1600-0536.1989.tb04728.x>
 59. Kinnunen T, Koskela M. Antibacterial and antifungal properties of propylene glycol, hexylene glycol and 1,3-butylene glycol *in vitro*. *Acta Derm Venereol.* 1991;71(2):148–50.
 60. Ismail GA, Gheda SF, Abo-Shady AM, Abdel-Karim OH. *In vitro* potential activity of some seaweeds as antioxidants and inhibitors of diabetic enzymes. *Food Sci Technol.* 2019;40:681–91. <https://doi.org/10.1590/fst.15619>
 61. Lingfa L, Tirumala A, Ankanagari S. GC-MS profiling of anticancer and antimicrobial phytochemicals in the vegetative leaf, root and stem of *Withania somnifera* (L.) Dunal. *Int J Second Metabolite.* 2024;11(1):63–77. <https://doi.org/10.21448/ijsm.1256932>
 62. Keskin D, Ceyhan N, Uğur A, Dbeys AD. Antimicrobial activity and chemical constitutions of West Anatolian olive (*Olea europaea* L.) leaves. *J Food Agric Environ.* 2012;10(2):99–102.
 63. Musini A, Rao MJP, Giri A. Phytochemical investigations and antibacterial activity of *Salacia oblonga* Wall ethanolic extract. *Ann Phytomedicine.* 2013;2(1):102–07.
 64. Alok Prakash AP, Suneetha V. *Punica granatum* (pomegranate) rind extract as a potent substitute for L-ascorbic acid with respect to the antioxidant activity. *Res J Pharm Biol Chem Sci.* 2014;5(2):597–603.
 65. Ye X, Li H, Anjum K, Zhong X, Miao S, Zheng G, et al. Dual role of indoles derived from intestinal microbiota on human health. *Front Immunol.* 2022;13:903526. <https://doi.org/10.3389/fimmu.2022.903526>
 66. Kim DH, Han SI, Go B, Oh UH, Kim CS, Jung YH, et al. 2-methoxy-4-vinylphenol attenuates migration of human pancreatic cancer cells via blockade of FAK and AKT signaling. *Anticancer Res.* 2019;39(12):6685–91. <https://doi.org/10.21873/anticancer.13883>
 67. Shoeb A, Chowta M, Pallemati G, Rai A, Singh A. Evaluation of antidepressant activity of vanillin in mice. *Indian J Pharmacol.* 2013;45(2):141–44. <https://doi.org/10.4103/0253-7613.108292>
 68. Dhanalakshmi C, Manivasagam T, Nataraj J, Justin Thenmozhi A, Essa MM. Neurosupportive role of vanillin, a natural phenolic compound, on rotenone induced neurotoxicity in SH-SY5Y

- neuroblastoma cells. Evid Based Complement Alternat Med. 2015;2015:626028. <https://doi.org/10.1155/2015/626028>
69. Anand A, Khurana R, Wahal N, Mahajan S, Mehta M, Satija S, et al. Vanillin: a comprehensive review of pharmacological activities. *Plant Arch*. 2019;19(2):1000–04.
 70. Fernandez-Bolanos JG, Lopez O, Fernandez-Bolanos J, Rodriguez-Gutierrez G. Hydroxytyrosol and derivatives: isolation, synthesis and biological properties. *Curr Org Chem*. 2008;12(6):442–63. <https://doi.org/10.2174/138527208784083888>
 71. Vahdati SN, Lashkari A, Navasatli SA, Ardestani SK, Safavi M. Butylated hydroxyl-toluene, 2,4-di-tert-butylphenol and phytol of *Chlorella* sp. protect the PC12 cell line against H₂O₂-induced neurotoxicity. *Biomed Pharmacother*. 2022;145:112415. <https://doi.org/10.1016/j.biopha.2021.112415>
 72. Varsha KK, Devendra L, Shilpa G, Priya S, Pandey A, Nampoothiri KM. 2,4-di-tert-butyl phenol as the antifungal, antioxidant bioactive purified from a newly isolated *Lactococcus* sp. *Int J Food Microbiol*. 2015;211:44–50. <https://doi.org/10.1016/j.ijfoodmicro.2015.06.025>
 73. Kuwahara H, Kanazawa A, Wakamatu D, Morimura S, Kida K, Akaike T, et al. Antioxidative and antimutagenic activities of 4-vinyl-2,6-dimethoxyphenol (canolol) isolated from canola oil. *J Agric Food Chem*. 2004;52(14):4380–87. <https://doi.org/10.1021/jf040045+>
 74. Ganesan T, Subban M, Christopher Leslee DB, Kuppannan SB, Seedeve P. Structural characterization of n-hexadecanoic acid from the leaves of *Ipomoea eriocarpa* and its antioxidant and antibacterial activities. *Biomass Convers Biorefin*. 2024;1–12. <https://doi.org/10.1007/s13399-022-03576-w>
 75. Hui Y, Wang X, Yu Z, Fan X, Cui B, Zhao T, et al. Scoparone as a therapeutic drug in liver diseases: pharmacology, pharmacokinetics and molecular mechanisms of action. *Pharmacol Res*. 2020;160:105170. <https://doi.org/10.1016/j.phrs.2020.105170>
 76. Hao J, Shen X, Lu K, Xu Y, Chen Y, Liu J, et al. Scoparone attenuates cholestatic liver injury through regulating hepatic bile acid metabolism, ameliorating periductal fibrosis and inhibiting inflammatory response. *Pharmacol Res Mod Chin Med*. 2023;8:100292. <https://doi.org/10.1016/j.prmcm.2023.100292>
 77. Choi BR, Kim HK, Park JK. Penile erection induced by scoparone from *Artemisia capillaris* through the nitric oxide–cyclic guanosine monophosphate signaling pathway. *World J Mens Health*. 2017;35(3):196–204. <https://doi.org/10.5534/wjmh.17023>
 78. Al-Marzoqi AH, Hameed IH, Idan SA. Analysis of bioactive chemical components of two medicinal plants (*Coriandrum sativum* and *Melia azedarach*) leaves using gas chromatography-mass spectrometry. *Afr J Biotechnol*. 2015;14(40):2812–30. <https://doi.org/10.5897/AJB2015.14956>
 79. Lerman A, Zeiher AM. Endothelial function: cardiac events. *Circulation*. 2005;111(3):363–68. <https://doi.org/10.1161/01.CIR.0000153339.27064.14>
 80. Sakakibara S, Murakami R, Takahashi M, Fushimi T, Murohara T, Kishi M, et al. Vinegar intake enhances flow-mediated vasodilatation via upregulation of endothelial nitric oxide synthase activity. *Biosci Biotechnol Biochem*. 2010;74(5):1055–61. <https://doi.org/10.1271/bbb.90953>
 81. Samad A, Azlan A, Ismail A. Therapeutic effects of vinegar: a review. *Curr Opin Food Sci*. 2016;8:56–61. <https://doi.org/10.1016/j.cofs.2016.03.001>
 82. Brighenti F, Castellani G, Benini L, Casiraghi MC, Leopardi E, Crovetti R, et al. Effect of neutralized and native vinegar on blood glucose and acetate responses to a mixed meal in healthy subjects. *Eur J Clin Nutr*. 1995;49(4):242–47.
 83. Johnston CS, Steplewska I, Long CA, Harris LN, Ryals RH. Examination of the antiglycemic properties of vinegar in healthy adults. *Ann Nutr Metab*. 2010;56(1):74–79. <https://doi.org/10.1159/000272133>
 84. Liatis S, Grammatikou S, Poulia KA, Perrea D, Makrilakis K, Diakoumopoulou E, et al. Vinegar reduces postprandial hyperglycaemia in patients with type II diabetes when added to a high, but not to a low, glycaemic index meal. *Eur J Clin Nutr*. 2010;64(7):727–32. <https://doi.org/10.1038/ejcn.2010.89>
 85. Khabibrakhmanova AM, Faizova RG, Lodochnikova OA, Zamalieva RR, Latypova LZ, Trizna EY, et al. The novel chiral 2(5H)-furanone sulfones possessing terpene moiety: synthesis and biological activity. *Molecules*. 2023;28(6):2543. <https://doi.org/10.3390/molecules28062543>
 86. Trizna EY, Khakimullina EN, Latypova LZ, Kurbangalieva AR, Sharafutdinov IS, Evtyugin VG, et al. Thio derivatives of 2(5H)-furanone as inhibitors against *Bacillus subtilis* biofilms. *Acta Naturae*. 2015;7(2):102–07.
 87. Sung WS, Jung HJ, Park K, Kim HS, Lee IS, Lee DG. 2,5-dimethyl-4-hydroxy-3(2H)-furanone (DMHF); antimicrobial compound with cell cycle arrest in nosocomial pathogens. *Life Sci*. 2007;80(6):586–91. <https://doi.org/10.1016/j.lfs.2006.10.008>
 88. Aqeel MT, Rahman NU, Khan AU, Khan MT, Ashraf Z, ul Hassan SS, et al. Cardioprotective effect of 2-methoxy phenol derivatives against oxidative stress-induced vascular complications: an integrated *in vitro*, *in silico* and *in vivo* investigation. *Biomed Pharmacother*. 2023;165:115240. <https://doi.org/10.1016/j.biopha.2023.115240>
 89. Bharath B, Perinbam K, Devanesan S, AlSalhi MS, Saravanan M. Evaluation of the anticancer potential of hexadecanoic acid from brown algae *Turbinaria ornata* on HT–29 colon cancer cells. *J Mol Struct*. 2021;1235:130229. <https://doi.org/10.1016/j.jmolstruc.2021.130229>

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonpublishing.com/journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonpublishing.com/journals/index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>)

Publisher information: Plant Science Today is published by HORIZON e-Publishing Group with support from Empirion Publishers Private Limited, Thiruvananthapuram, India.