



RESEARCH COMMUNICATION

Phylogenetic study of mangrove associate grass *Myriostachya wightiana* (Nees ex Steud.) Hook. f. using *rbcl* gene sequence

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ABSTRACT

Myriostachya is a monotypic genus in the family Poaceae, with the only known species *Myriostachya wightiana* (Nees ex Steud.) Hook.f. It is a mangrove associate grass primarily distributed along the muddy streams and channels in intertidal mangrove swamps of India, Bangladesh, Sri Lanka, Myanmar, Thailand and Sumatra. Molecular identification and evolutionary studies of *M. wightiana* is unreported till now. Therefore, in this study, the phylogenetic analysis of *M. wightiana* was established with related family members by using chloroplast *rbcl* gene-based systematics. The molecular phylogeny was accomplished by DNA extraction, PCR amplification and sequencing of the *rbcl* gene and phylogenetic analysis. The genomic DNA was extracted using the CTAB method and the *rbcl* gene amplification is by using the F-5'ATGTCACCACAAACAGAACTAAAGC3' and R-5'CTTCGGCACAAAATAAGAAACGATCTC3' primers. Phylogenetic analysis of *M. wightiana* was performed by multiple sequence alignment with UPGMA, and the Maximum-parsimony phylogenetic tree was constructed using MEGAX. *Myriostachya wightiana* *rbcl* gene sequence shows the highest similarity to *Paspalum* species, and in the phylogenetic tree *M. wightiana* has a close branch with *Paspalum vaginatum*. The evolutionary divergence from *M. wightiana* is maximum (0.49) to *Sorghum propinquum* and minimum (0.01) to *Oryza officinalis* and *Oryza punctata*. This study concluded that *M. wightiana* has a strong morphological and phylogenetic relationship with salt-tolerant *Paspalum* sp.

Introduction

Myriostachya is monotypic genus in the Poaceae family, with *Myriostachya wightiana* (Nees ex Steud.) Hook. f. being the only species (1). The species is tropical with, its native range from the Indian Subcontinent to West Malesia. It is widely located in the intertidal mangrove swamps of India, Bangladesh, Sri Lanka, Myanmar, Thailand and Sumatra (2). In India, it is mainly distributed on the southeast coast of the Bay of Bengal. *M. wightiana* is a large, densely clumped perennial grass growing up to 3 meters. It frequently occurs along with *Acanthus ilicifolius* L., *Nypa fruticans* Wurmb. and *Porteresia coarctata* (Roxb.) Tateoka. The species grows well in saline water habitat rather than fresh water due to structural adaptations like the thick epidermis, sclerenchymatous vascular system, salt secretion glands, conspicuous metaxylem and broad phloem region in stem and leaf, dense cortex and lignified root exodermis (3).

The taxonomic position of *M. wightiana* is recorded based on the morphological features. However, the morphology-based systematics and

evolutionary studies of Poaceae are not reliable due to the higher number of taxa, simplicity of floral architecture and vegetative structure, dynamic and mosaic evolution (4). Because of these challenges, DNA based molecular systematics are being used to generate promising results. According to the available literature, the evolutionary studies of *M. wightiana* have not been carried out yet. Thus, this study aims to build phylogenetic relationships of *M. wightiana* by DNA dependent molecular systematics. DNA barcoding is a relatively quick and accurate method for identification of any plant or animal species (5). DNA barcoding can be used in molecular systematics to identify the new species in conjugation with conventional taxonomic approaches (6). The fundamental principle of DNA sequence utilisation in phylogenetic analysis is the occurrence of nucleotide changes over time. Therefore, the estimation and reconstruction of evolutionary relationships between different organisms are possible (7).

In plants, the chloroplast genome is quite helpful for evolutionary and phylogenetic experiments, especially above the species level because of its relatively abundant DNA material, single-copy genes,

conservative nucleotide substitution rate. Therefore, most phylogenetic reconstruction carried out so far in plant systematics is based on molecular evidence from the cp DNA genes. The plastid-encoded *rbcL* gene is the most widely used gene to establish molecular phylogeny (8). The *rbcL* gene is a single-copy gene with approximately 1430 base pairs length, and it is free from longitudinal mutations, thereby showing a fair conservative rate of evolution (9). Chloroplast *rbcL* gene is responsible for encoding the largest subunit of ribulose 1,5 carboxylase/oxygenase bisphosphate (RUBISCO/RUBPCase) which consists of all the active sites of the enzyme. Environmental factors could be the selective pressure to increase the CO₂ utilisation efficiency of RUBISCO through adaptive evolution of the *rbcL* gene (10). This study aims to establish *M. wightiana* phylogenetic relationship with related family members using the chloroplast *rbcL* gene.

Materials and Methods

Sample collection

Healthy and young *M. wightiana* (Fig. 1) plant material were collected from the mangroves (Fig. 2) of Bhavanapadu (Long: 18° 33' 52" to 18° 32' 11" N; Lat: 84° 21' 26" E to 84° 18' 22" E) which is located in the northeast of Andhra Pradesh, adjoining the Bay of Bengal, India. The collected plant samples were aseptically transferred to the zip bags and transported to the laboratory. The plant species was authenticated by Dr. S. Hara Sreeramulu, Taxonomist, Department of Botany, Dr. V. S. Krishna College, Visakhapatnam and the voucher specimen (No. 00564/AP) was deposited at the Herbarium, Department of Botany, Andhra University.

DNA extraction

Total DNA content was extracted from the leaves of *M. wightiana* was carried out by the CTAB method (11). Surface sterilised healthy and young leaves were grounded using liquid nitrogen and 500 µg of



Fig. 1. *Myriostachya wightiana* (Nees ex Steud.) Hook. f.

grounded leaf powder was mixed with 750 µl of CTAB, 20 µl of mercaptoethanol in a 2 ml Eppendorf tube. The mixture was vortexed intermittently and then incubated in a water bath at 65 °C for 45 min. After incubation, the mixture was allowed to cool until it reaches room temperature. Subsequently, 750 µl of chloroform/isoamyl alcohol (24:1) was added to the mixture, and the tube was centrifuged at 12000 rpm (Eppendorf, 5810R) for 10 min and the upper aqueous phase was separated into a new tube. The chloroform/isoamyl alcohol extraction was repeated twice to the aqueous phase and centrifuged for 5 min. To the resultant supernatant, 0.1 ml of 3M sodium acetate (pH 4.6), 2 vol. of 95% ethanol was added and incubated at -20 °C for 1 hr to precipitate DNA. Then the precipitate was centrifuged at 12000 rpm for 10 min. The DNA pellet was washed twice with 750 µl of 70 % ethanol, then centrifuged for 10 minutes at 10000 rpm. The resulting DNA pellet was rewashed twice with 96 % ethanol and dried in a desiccator for 15 min. The DNA pellet was stored in a -20 °C freezer. The quantity of isolated DNA was estimated by diphenylamine test (12) and the quality was estimated using 1% agarose gel electrophoresis (Bio-Rad, Mini-sub cell GT electrophoresis system). The gels were visualised in a UV transilluminator (Invitrogen, LB0100) and the image was captured by using a Gel documentation system.

PCR amplification and sequencing of *rbcL* gene

Myriostachya wightiana chloroplast *rbcL* gene amplification was conducted using primers, *rbcL* F-5'ATGTCACCACAAACAGAACTAAAGC3' (13) and *rbcL* R-5' CTTCGGCACAAAATAAGAAACGATCTC 3' (14). A 20 µl of PCR reaction mixture comprising 1 unit of pfu DNA polymerase, 2 µl of 10X PCR buffer with 20 mM MgSO₄, 2 µl of 2 mM dNTPs, 2 µl of 10 M each primer, and 25 ng of template DNA. The PCR amplification (Bio-Rad, PTC-200) was conducted for 35 cycles. The thermocycler temperature configuration was started with the initial denaturation for 5 minutes at 94 °C, then continued to 35 cycles with denaturation for 30 seconds at 94 °C, annealing for 40 seconds at 50 °C, extension for 1 minute at 72 °C, and the final cycle extension time at 72 °C was raised to 7 minutes. The amplified *rbcL* PCR products were observed using 1.4% agarose gel electrophoresis with ethidium bromide staining, a column-based DNA purification kit purified the single band PCR amplicons. The purified PCR product was subjected to the Sanger's di-deoxy sequencing in both forward and reversed directions using an ABI prism 3700 DNA analyser. The resultant chloroplast *rbcL* sequence of *M. wightiana* was submitted to the NCBI database (accession number KY293284).

Sequence alignment and Phylogenetic analysis

The evolutionary analysis of *M. wightiana* was conducted by constructing a phylogenetic tree with some major species of Poaceae. *Myriostachya wightiana* *rbcL* gene sequence was subjected to the BLASTn in the NCBI server to identify the homologous sequence or species. From the NCBI database, 33 species were selected based on their degree of homology with the target *rbcL* gene sequence. The selected 33



Fig. 2. Habitat of *Myriostachya wightiana* (Nees ex Steud.) Hook. f.

species' *rbcL* gene sequences were derived from the nucleotide NCBI database. In the selected thirty-three species for phylogenetic analysis, thirty species belong to the Poaceae family, and the remaining three species were used as an outgroup that consists of marine algae *Saccharina latissimi* (L.) C.E.Lane, C.Mayes, Druehl and G.W.Saunders., *Undaria pinnatifida* (Harv.) Suringar and *Porphyra haitanensis* T.J.Chang and B.F.Zheng, 1960. which belongs to Laminariaceae, Alariaceae and Bangiaceae respectively. The marine algae were used as an out group to known the evolutionary progress from the marine algae to marine grasses and to terrestrial grasses. Multiple sequence alignment was conducted with an advanced cluster method UPGMA using Muscle embedded in MEGAX to search homology of *rbcL* gene sequences between the *M. wightiana* and the selected 33 species. To analyse the variations among the sequences, a distance matrix was determined, and based on the differences expressed in the distance matrix; a maximum-parsimony tree was constructed using MEGAX. The evaluation of phylogenetic tree topologies was done by the bootstrap method with 1000 replicates for all nodes (15).

Results and Discussion

DNA extraction and quantification

The result was given as Mean \pm Standard Deviation obtained from three independent experiments. As part of DNA barcoding, the complete genome of *M. wightiana* was successfully extracted. The isolated DNA appeared as a prominent band on 1% agarose gel (Fig. 3A) and the isolated total DNA content was

measured as $663 \pm 54 \mu\text{g/gm}$. DNA barcoding is one of best way to classify new species and also to collect a database of the reference sequences (16).

PCR amplification and sequence analysis

The PCR amplified *M. wightiana* chloroplast *rbcL* gene was successfully run by the 1.5% agarose gel electrophoresis. In the agarose gel (Fig. 3B), the first well shows marker DNA and the remaining three wells indicate amplified *rbcL* gene product. The thick and single bands in the agarose gels indicate that the *rbcL* gene amplification was done successfully, and the size of the amplified *M. wightiana* *rbcL* gene was approximately $\pm 650\text{bp}$. The yield and consistency of DNA bands in the agarose gels determines the universality of selected primers and their discriminating strength. According to one report, universal primers identify the regions in the *rbcL* gene of Angiosperms and display a high degree of universality in terrestrial plants (14). The direct nucleotide sequencing of PCR amplified products is now emerging as an important field of evolutionary studies and systematics (17). An ideal DNA barcode can be recovered and gives maximum discrimination among species with a single pair of primers suitable for bidirectional sequencing with a minor sequence modification. The chloroplast *rbcL* gene contains the least number of variable regions and it is referred to as a best-characterised gene that is easily retrievable with common PCR primers. It was proposed that, out of the nuclear and plastid genomes, plastid genomes are used for phylogenetic research in plants since they are thought to have similar ancestors (18). Our research used the chloroplast *rbcL* gene sequence, considering that the *rbcL* gene is quickly amplified and sequenced in many terrestrial plants and impacts in phylogenetic studies by placing the species in a correct genus and plant family.

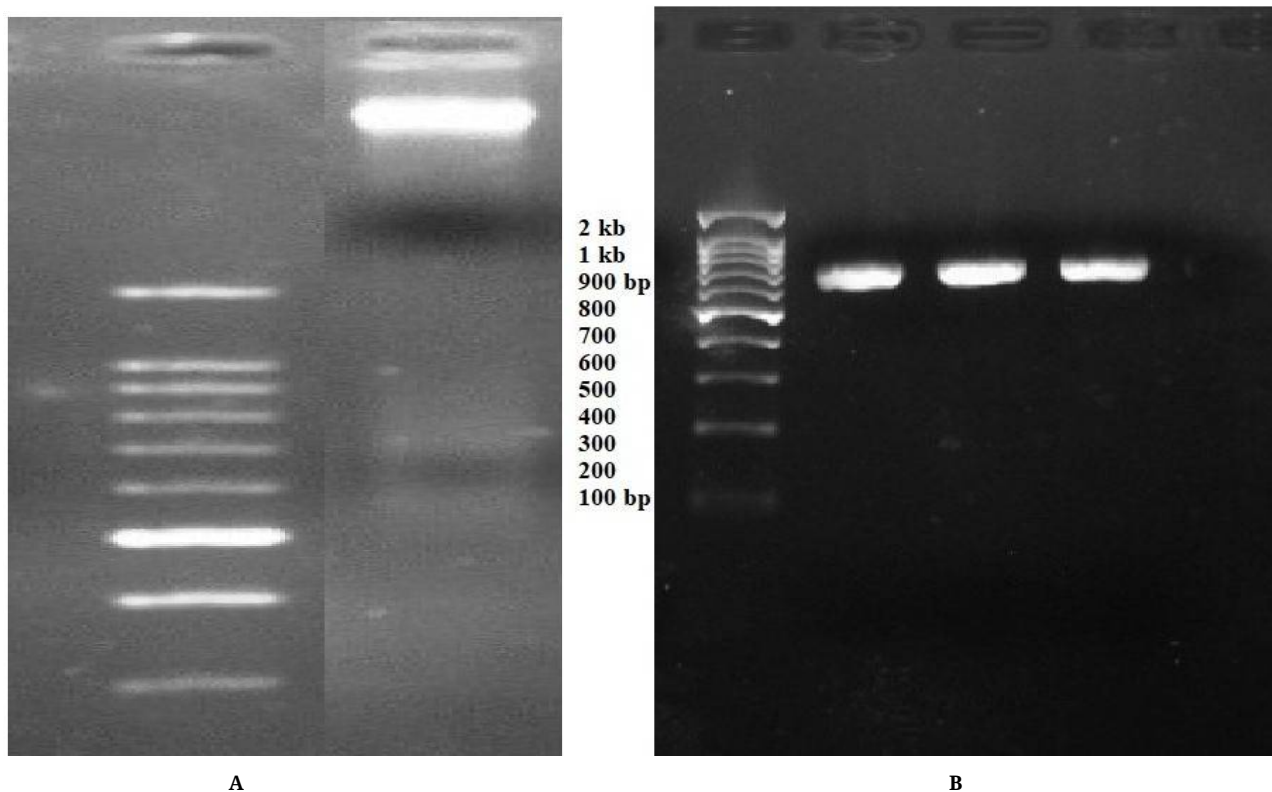


Fig. 3. (A) Photograph showing genomic DNA band in 1% agarose gel. **(B)** Photograph showing the PCR amplified rbcL gene band in 1.5% agarose gel.

M. wightiana chloroplast rbcL gene nucleotide composition was computed by Seqstate V.1.21 server (19). The amplified conserved region of the rbcL gene (GenBank Accession No. KY293284) has 604 nucleotides and was estimated to have a molecular weight of 373 KDa. The nucleotide composition of the rbcL gene consists of 164 bp Adenine (A), 173 bp Thymine (T), 140 bp Guanine (G), 127 bp Cytosine (C), and the percentage of GC was measured as 44.2. The Extinction coefficient of the *M. wightiana* rbcL gene is estimated to be $9805831 \text{ Mol}^{-1} \text{ cm}^{-1}$. Kusumi and Tachida (20) reported that the GC content in the plants differs from 28-42%. It has been reported that the GC content of the rbcL gene in wild *Solanum* sp. is 43.9% (21). The current findings showed that the rbcL genes in *M. wightiana* have comparatively low GC content.

Multiple sequence alignment and phylogenetic tree analysis

The BLAST search results showed similar rbcL gene sequences of other species with their percentage of identity against the *M. wightiana* rbcL gene sequence. The BLASTn results revealed that the *M. wightiana* rbcL gene sequence showed the highest similarity to *Paspalum* sp., with an identity of 99% and an E-value of 0.0. From the hits, a total of 33 rbcL gene sequences from different species were selected to construct evolutionary relationships through multiple sequence alignment using the UPGMA program in MEGAX. The multiple sequence alignments indicated a variable number of deletions and insertions in the chloroplast rbcL. The phylogenetic parameters such as variance, parsimony sites, and overall mean distance were calculated as 0.52, 0.32 and 0.1 respectively in the

rbcL alignments of family Poaceae. The evolutionary divergence of *M. wightiana* with its other species of Poaceae was determined. A satisfactory result was established by using the rbcL gene as a marker to evaluate the phylogenetic relationship among the grass species. The evolutionary divergence among sequences at the generic level varies from 0.01-0.49. *M. wightiana* showed the highest evolutionary divergence with the species of *Sorghum propinquum* (Kunth) Hitchc. (0.49), *Undaria pinnatifida* (Harv.) Suringar (0.33) and *Porphyra haitanensis* T.J.Chang and B.F.Zheng. Whereas *M. wightiana* shows the least evolutionary divergence with the species of *Oryza officinalis* Wall., (0.01), *O. punctata* Kotschy ex Steud. (0.01), *O. glaberrima* Steud. (0.02) and *O. nivara* S.D.Sharma and Shastry (0.02). Multiple sequence alignments of the rbcL gene sequences revealed that the rbcL gene is highly conserved throughout the Poaceae family.

In the phylogenetic tree (Fig. 4), there were five main clades. The first main clade consists of two subclades. The first subclade of main clade-I composed *Paspalum* sp. and the second subclade composed of *Zea* and *Sorghum* species. The second and third main clades consist of *Triticum* and *Oryza* species. The fourth main clade composed only *Sorghum propinquum*. The fifth main clade consists of marine macroalgae *Porphyra haitanensis*, *Saccharina latissimi* and *Undaria pinnata*. The phylogenetic tree indicated that the *M. wightiana* found a close branch with *Paspalum vaginatum* Sw. (GenBank Accession No. LN907995). In the phylogenetic tree, the clades are organised mainly with several species in the same genus according to their similarities. *M. wightiana* is known to be the nearest species to seashore grasses in phylogenetic analysis.

Conclusion

An ideal taxonomic recognition of species is essential for the proper management of any organism. Species-level evolutionary information of a plant can be provided by the chloroplast *rbcL* gene based phylogenetic analysis. The present study of *rbcL* gene sequence and phylogenetic analysis explored the evolutionary divergence and relatedness of the *M.*

wightiana. The *rbcL* gene sequence and multiple sequence alignment revealed that the *M. wightiana* showed 99% homology to *Paspalum* sp. The evolutionary divergence from *S. propinquum* to *M. wightiana* was estimated as 0.49, which was found maximum and the minimum was for *O. officinalis* and *O. punctata* that are 0.01. From the molecular phylogeny by the *rbcL* gene, it was concluded that the

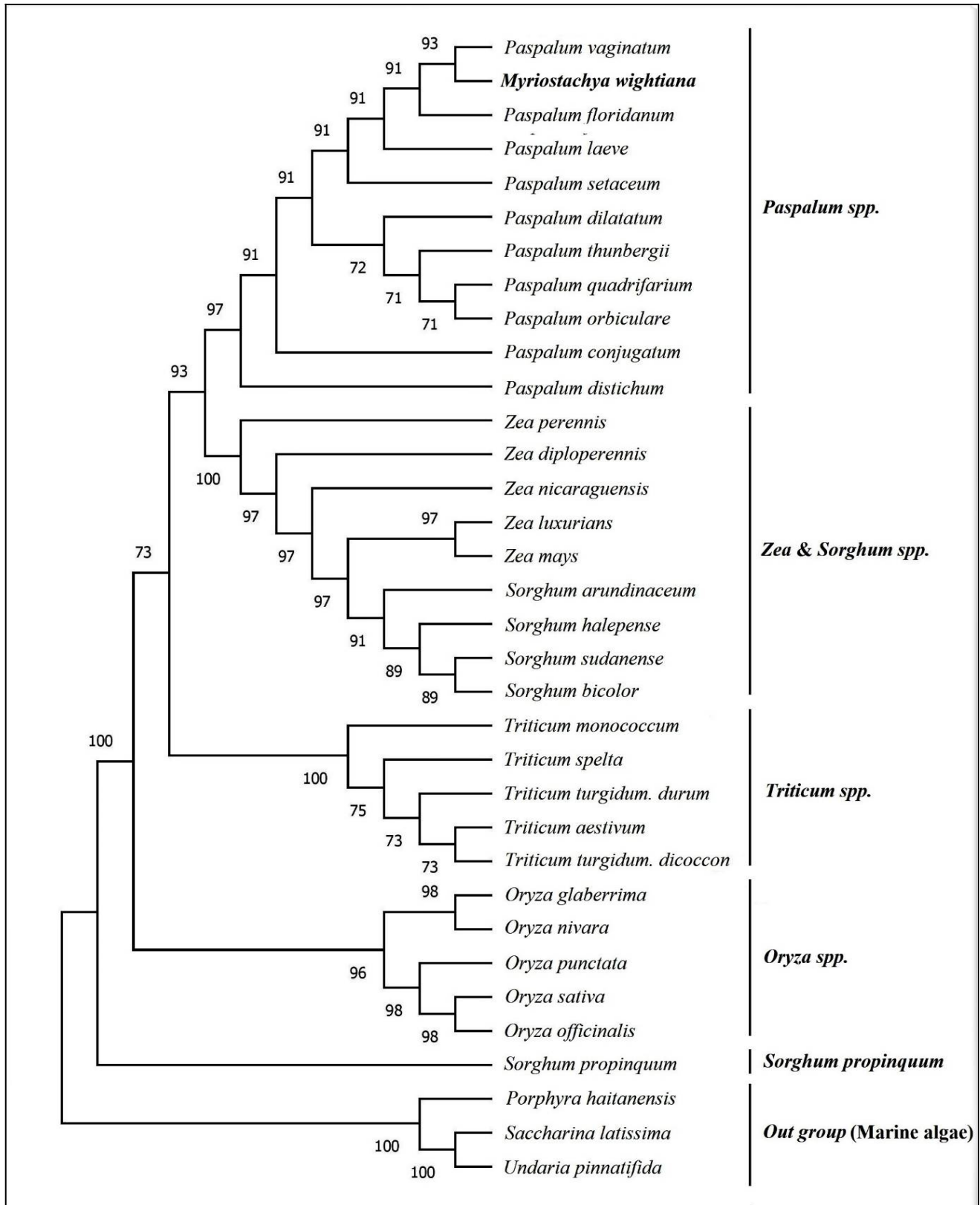


Fig. 4. Maximum Parsimony tree of *Myriostachya wightiana* and other species based on the *rbcL* gene. Bootstrap values are indicated on the branches.

M. wightiana has a strong relationship to salt-tolerant grasses like *Paspalum* sp.

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Authors' contributions

MKK carried out isolation of *rbcl* gene, sequencing, multiple sequence alignment, construction of the phylogenetic tree and drafting of the manuscript. BVS participated in the design and planning of research work, helped to write the article and corrected the manuscript. The final manuscript was read and approved by all the authors.

Conflict of interests

The authors declare that there are no conflicts of interest.

Supplementary file

Table 1. Estimates of evolutionary divergence among the selected species.

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