



RESEARCH ARTICLE

# Effects of gamma radiation on physio-morphological traits of finger millet (*Eleusine coracana* (L.) Gaertn.)

Latha Sellapillaibanumathi<sup>1</sup>, Arulbalachandran Dhanarajan<sup>1\*</sup>, Aamir Raina<sup>2,3</sup> & Aswini Ganesan<sup>1</sup>

<sup>1</sup>Division of Crop Breeding and Molecular Breeding Laboratory, Department of Botany, Periyar University, Salem, Tamil Nadu, India

<sup>2</sup>Mutation Breeding Laboratory, Department of Botany, Aligarh Muslim University, Aligarh, Uttar Pradesh, India

<sup>3</sup>Botany Section, Women's College, Aligarh Muslim University, Aligarh, Uttar Pradesh, India

\*Email: arul78bot@gmail.com



## ARTICLE HISTORY

Received: 22 February 2021

Accepted: 11 July 2021

Available online

Version 1.0 (Early Access): 30 November 2021

Version 2.0 : 01 January 2022



## Additional information

**Peer review:** Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

**Reprints & permissions information** is available at [https://horizonepublishing.com/journals/index.php/PST/open\\_access\\_policy](https://horizonepublishing.com/journals/index.php/PST/open_access_policy)

**Publisher's Note:** Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Indexing:** Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, etc. See [https://horizonepublishing.com/journals/index.php/PST/indexing\\_abstracting](https://horizonepublishing.com/journals/index.php/PST/indexing_abstracting)

**Copyright:** © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>)

## CITE THIS ARTICLE

Sellapillaibanumathi L, Dhanarajan A, Raina A, Ganesan A. Effects of gamma radiation on physio-morphological traits of finger millet (*Eleusine coracana* (L.) Gaertn.). Plant Science Today. 2022;9(1):89–95. <https://doi.org/10.14719/pst.1142>

## Abstract

The present study was carried out to analyse the effects of gamma radiations on physio-morphological traits of seven days old M<sub>1</sub> seedlings of *Eleusine coracana* (L.) Gaertn. (finger millet). The finger millet seeds were irradiated with different doses of gamma rays viz., 100 Gy, 200 Gy, 300 Gy, 400 Gy, 500 Gy, 600 Gy, 700 Gy, 800 Gy, 900 Gy and 1000 Gy. Lower and higher doses of gamma rays induced a substantial increase and decrease in the mean values of physiological and morphological traits respectively. The results revealed a progressive decrease in chlorophyll fluorescence with an increasing dose of gamma rays. Among all the mutagen doses employed, 600 Gy gamma-irradiated seeds showed maximum mean values of physiological and morphological traits in finger millet. Hence, 600 Gy gamma rays may be employed in other related crop species to improve the agro-economic traits.

## Keywords

Finger millet, gamma irradiation, photosynthetic pigments, physio-morphological traits

## Introduction

*Eleusine coracana* (L.) Gaertn., commonly known as finger millet is one of the major nutritious food crops with high quantities of mineral nutrients such as calcium, magnesium, phosphorus and iron (1). The finger millet possesses fibre rich starch that enhances digestion and reduces intestinal disorders (2, 3). Millets are least affected by continuous climate change due to their ability to thrive at high temperatures and in low moisture soil (4, 5). Genetic variability is one of the primary prerequisites for crop improvement programmes. Among the breeding strategies, mutation breeding is a coherent tool to increase genetic variability and has been successful in the development of thousands of new varieties in hundreds of crop species (6). Different mutagens have been used by the workers from time to time to accomplish the desired results, however, radiation-induced mutagenesis is mostly used for the improvement of different agro-economic traits (7-10). Among the radiations, gamma rays are widely preferred physical mutagens that affect plant growth and development by altering the biochemical, physiological and morphological traits (11-13). Moreover, the highest number of mutants have been developed via the use of gamma rays due to their ability to induce single and double-stranded DNA breaks (14, 15). Gamma rays interact with plant tissues in two ways: direct and indirect interaction. In direct interaction, gamma rays deposit energy packets on DNA and cause ionisation and excitation of electrons. Depending upon the amount of energy

deposition, gamma rays induce single and double-stranded breaks in the DNA that later on result in variations in different agro-economic traits (16, 17). In indirect interaction, gamma rays interact with water molecules and lead to the formation of highly reactive free radicals in a process called radiolysis of water. These free radicals then interact with the DNA and lead to several types of mutations such as substitution, base alterations, base deletions, chromosomal abnormalities and DNA breaks (18-20). The photosynthetic apparatus in plants are reported to be most sensitive part to mutagen induced alterations (21). Gamma rays affect photo pigments in positive or negative ways, depending on the dose and duration of exposure. Therefore, the visualization of fluorescence emitted by photosynthetic pigments in response to mutagen doses is imperative to find out steady-state fluorescence peaks under light illumination (22- 24). The fluorescence spectra in turn depend on chlorophyll contents and the absorbance of the leaves (25). In the present study, we investigated the effects of gamma rays on physio-morphological traits and analysed chlorophyll fluorescence of gamma rays treated M<sub>1</sub> finger millet seedlings.

## Materials and Methods

### Biological Material

The seeds of finger millet cultivar Paiyur-2 (parent- VL 145 x Selection 10) was procured from Tamil Nadu Agriculture Research Centre, Paiyur, Krishnagiri, Tamil Nadu, India. The experiments were performed at the Department of Botany, School of Life Sciences, Periyar University, Salem, Tamil Nadu, India.

### Gamma radiation treatment

The chosen finger millet seeds were cleaned and packed in paper bags and treated with different doses of gamma rays viz., 100 Gy, 200 Gy, 300 Gy, 400 Gy, 500 Gy, 600 Gy, 700 Gy, 800 Gy, 900 Gy and 1000 Gy at Indira Gandhi Centre for Atomic Research, Kalpakkam, Tamil Nadu, India using cobalt 60 as a source of gamma rays. The average measurement rate of uncertainty range of 1-3 Gy/Sec was maintained for this investigation. The calibration of the gamma chamber was accomplished with the Fricke dosimetry system for absorbed doses in line with the guidelines of the International Atomic Energy Agency.

### Growth conditions

The control and treated seeds (n = 3) were kept moistened in germinating paper inside glass petri dishes (9 × 3 cm). Seedlings were grown individually for all doses in a greenhouse environment. Seven days old seedlings were used to determine the chlorophyll fluorescence (CF) intensity and content of photosynthetic pigments viz., chlorophyll-*a*, chlorophyll-*b* and carotenoids. Seedlings were raised in a nursery bed for three weeks followed by subsequent transplantation in the field beds and sown in randomized block design with three replications. All recommended agricultural practices (irrigation, fertilizers and weeding) were followed throughout the field study.

### Chlorophyll extraction

Young leaves (100 mg) collected from seven days old seedlings were ground in 1 ml of 80 % acetone with pestle and mortar. The leaf extract was then spun at 2500 revolutions per minute (rpm) for 10 min at 20 °C in a centrifuge (Remi R 24). The homogenate was re-extracted with 80 % acetone until the green colour disappeared in the residue and the extract was pooled and made up to 2 ml with 80 % acetone. Then 2 ml of extract was transferred into a cuvette and the absorbance was read at 663, 645 and 480 nm in a UV spectrometer (Elico), against 80 % acetone as blank. Chlorophyll *a*, Chlorophyll *b* and carotenoid contents were estimated as per the following formula (26, 27).

$$\text{Chlorophyll } a \text{ } (\mu\text{g/ml}) = (12.7 \times A_{663}) - (2.69 \times A_{645})$$

$$\text{Chlorophyll } b \text{ } (\mu\text{g/ml}) = (22.9 \times A_{645}) - (4.68 \times A_{663})$$

$$\text{Carotenoids } (\mu\text{g/ml}) = A_{480} + (0.114 \times A_{663}) - (0.638 \times A_{645})$$

Where, A<sub>645</sub> = absorbance at 645 nm; A<sub>663</sub> = absorbance at 663 nm; A<sub>480</sub> = absorbance at 480 nm.

### Chlorophyll fluorescence

Chlorophyll fluorescence was determined in the leaves of both control and gamma-irradiated seedlings using a fluorescence spectrophotometer (Jasco, FP-8200 and Japan). The data interval (0.5 nm), scan rate (200 nm/min) and wavelengths Ex / Em (excitation/emission) were kept in a range from 400 to 700 nm. The wavelengths used for emission/excitation could reveal the variations after exposure to different doses of gamma rays.

### Morphological and yield parameters

Regular field evaluations were carried out to collect the data on morphological and yield traits such as plant height (cm), leaf length (cm), number of leaves per plant, number of tillers per plant, days to 50% flowering, panicle length (cm), number of panicle per plant and 1000 seed weight.

### Statistical analysis

The gamma irradiated samples were analyzed with three replications in petri dish studies on 7-days old M<sub>1</sub> seedlings. One-way Analysis of Variance (ANOVA) and Pearson's correlation was performed in SPSS ver. 21.0 that allowed us to visualize the significance of data

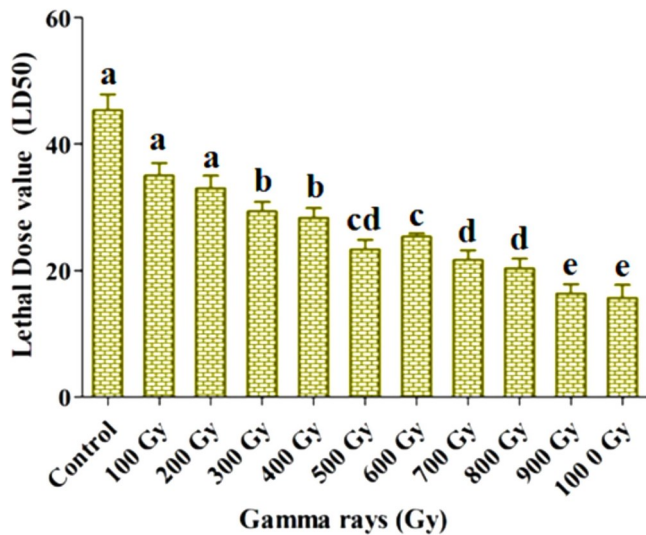
## Results

### Lethal Dose (LD50)

Based on 50% mortality, the radio sensitivity of seedlings to different doses of gamma rays was evaluated. The apparent variation of 50% reduction in germination was observed at 600 Gy gamma rays treatment (25.33%) as compared to control (45.30%). The seedling survival depicted a negative correlation with increasing doses of radiation given in Fig. 1.

### Effects of gamma rays on photosynthetic pigments

Our results revealed a progressive decrease in the contents of the photosynthetic pigments viz., chlorophyll *a* and chlorophyll *b* with increasing doses of gamma rays. The maximum decrease in photosynthetic pigments was



**Fig. 1.** Evaluation of LD<sub>50</sub> value in seven day old M<sub>1</sub> seedlings of finger millet treated with different doses of gamma rays. Mean within columns followed by the same letter is not different at the 1% level of significance based on Duncan's Multiple Range Test.

recorded in 1000 Gy gamma rays treatment (chlorophyll *a*: 0.52 chlorophyll *b*: 0.16 mg g<sup>-1</sup>FW) as compared to control (chlorophyll *a*: 1.54 chlorophyll *b*: 0.20 mg g<sup>-1</sup>FW). The results also revealed a dose-dependent significant increase in carotenoid contents (Car: F= 270.65, P< 0.01). The maximum increase in carotenoids contents was observed in 1000 Gy gamma rays treatment (3.88 mg g<sup>-1</sup>FW) as compared to control (1.31 mg g<sup>-1</sup>FW) shown in Table 1.

**Table 1.** Effects of different doses of gamma rays on chlorophyll *a*, chlorophyll *b* and carotenoid content (mg g<sup>-1</sup>FW) in 7 days old M<sub>1</sub> finger millet seedlings. The data is presented as mean ± SE (standard error) (n=3). Mean within columns followed by the same letter is not different at the 1% level of significance based on Duncan's Multiple Range Test.

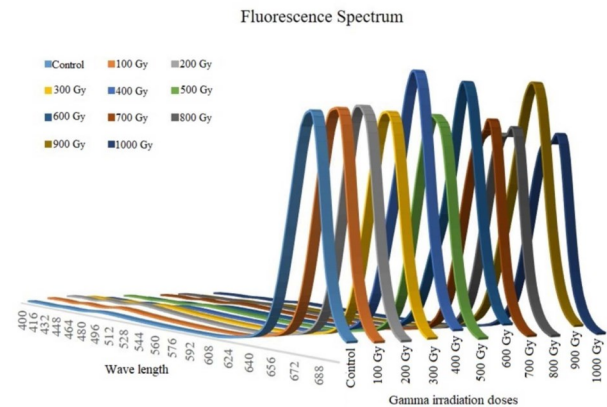
Gamma rays	Chl <i>a</i>	Chl <i>b</i>	Carotenoids
Control	1.54±0.01c	0.20±0.00de	0.98±0.00e
100 Gy	2.67±0.28a	0.29±0.00cde	1.06±0.12e
200 Gy	2.75±0.09a	0.30±0.00cde	1.30±0.04d
300 Gy	2.65±0.14a	0.33±0.04cd	1.38±0.01d
400 Gy	2.85±0.17a	0.36±0.04c	1.46±0.02cd
500 Gy	2.74±0.18a	0.61±0.15a	1.64±0.03c
600 Gy	3.53±0.16a	0.86±0.07a	2.12±0.00b
700 Gy	2.10±0.00b	0.63±0.03a	2.62±0.04a
800 Gy	1.69±0.23c	0.49±0.01bc	2.96±0.11a
900 Gy	1.11±0.00d	0.26±0.08cde	3.48±0.14a
1000 Gy	0.52±0.05e	0.16±0.01e	3.88±0.07a

The statistical analysis revealed a significant enhancement of the pigments at optimum doses (300 Gy, 400 Gy, 500 Gy, 600 Gy) compared to control (Chl *a*: F= 50.94, P<0.01) (Chl *b*: F= 18.71, P<0.01).

### Spectrofluorometric studies

The spectra of chlorophyll fluorescence were recorded within excitation wavelength varied from 400 to 700 nm. The peak value of 7529.67 obtained in the 400 Gy leaf extract sample revealed a maximum excitation spectra at 677 nm compared with a peak value of 6352.97 in control. The

results revealed a progressive decrease in fluorescence intensity with the increasing dose of gamma rays depicted in Fig. 2.



**Fig. 2.** Fluorescence spectrum of seven days old M<sub>1</sub> finger millet seedlings treated with different doses of gamma rays.

### Quantitative traits

In this study, mean performances of different quantitative traits were recorded in all doses in M<sub>1</sub> generation. The optimum doses (300 Gy, 400 Gy, 500 Gy and 600 Gy) increased the mean performance of quantitative traits. The maximum increase in plant height (130.89±4.75 cm) was recorded in 600 Gy gamma rays treatment as compared to control (90.84±0.47 cm). Plant height decreased significantly with increasing dose of gamma rays (F= 453.53, P<0.01) and a maximum reduction was noted in 1000 Gy gamma rays treatment (37.93±1.90 cm) shown in Table 2. The maximum

**Table 2.** Effects of different doses of gamma rays on morphological traits of finger millet in M<sub>1</sub> generation. The data is presented as mean ± SE (standard error) (n=3) Mean within columns followed by the same letter is not different at the 1% level of significance based on Duncan's Multiple Range Test

Gamma rays	Plant height	No. of leaves / plant	Leaf length	Tillers/plant
Control	90.84±0.47e	13.20±0.34gh	14.66±0.52h	3.40±0.17h
100 Gy	93.57±0.41de	14.80±0.49fg	25.71±1.43f	4.40±0.57f
200 Gy	95.48±0.13cd	16.70±0.84f	33.78±1.12e	6.10±0.24e
300 Gy	96.09±0.39bc	83.9±6.54e	64.91±0.81d	6.80±0.37d
400 Gy	98.55±0.16b	45.6±1.03d	56.91±0.94c	7.80±0.26c
500 Gy	100.90±0.34a	42.1±0.65c	43.13±0.88b	9.40±0.50b
600 Gy	130.89±4.75a	25.5±0.77a	37.25±0.32a	11.80±0.46a
700 Gy	102.72 ±10.93a	21.2±2.81b	37.25±0.32d	5.90±0.77e
800 Gy	90.18±1.42e	18.3±2.30e	27.20±1.35f	3.70±0.54g
900 Gy	54.60±3.69f	12.20±0.60g	18.07±1.07g	1.90±0.55i
1000 Gy	37.93±1.90g	6.10±1.00h	13.77±0.42h	1.00±0.35j

increase in the number of leaves per plant was recorded in 300 Gy gamma rays treatment (83.9±6.54). The results showed a decreasing trend in the number of leaves per plant with the increasing dose of gamma rays. The highest decrease in the number of leaves per plant was recorded in 1000 Gy gamma rays treatment (6.1±1.00) as compared to control (13.20±0.34) given in Table 2. The maximum leaf

length was recorded in 300 Gy gamma rays treatment ( $64.91 \pm 0.81$ ). The mean performance of leaf length per plant exhibited gradual reduction as the gamma rays dose increases. A higher reduction in plant leaf length was recorded in 1000 Gy gamma rays treatment ( $13.77 \pm 0.17$  cm) as compared to control ( $14.66 \pm 0.52$ ). The number of tillers per plant showed more reduction in 1000 Gy gamma rays treatment ( $1.0 \pm 0.35$ ) as compared to control ( $3.40 \pm 0.17$ ). Increased mean tillers per plant were recorded in 600 Gy gamma rays treatment ( $11.80 \pm 0.46$ ) shown in Table 2.

The maximum reduction of panicle number was recorded in 1000 Gy gamma rays treatment ( $0.5 \pm 0.17$ ) as compared to control ( $2.0 \pm 0.27$ ). The minimum day to 50% flowering was recorded in 600 Gy gamma rays treatment ( $77.4 \pm 0.54$ ) as compared to control ( $81.2 \pm 0.14$ ) (Table 3). The maximum increase in days to 50% flowering was recorded in 1000 Gy gamma rays treatment ( $89.4 \pm 0.17$ ). Our results revealed maximum panicle length in 600 Gy gamma rays treatment ( $66.02 \pm 1.17$ ) as compared to control ( $21.84 \pm 1.27$ ) depicted in Table 3.

**Table 3.** Effect of different doses of gamma rays on yield related traits of finger millet in  $M_1$  generation. The data is presented as mean  $\pm$  SE (standard error) ( $n=3$ ). Mean within columns followed by the same letter is not different at the 1% level of significance based on Duncan's Multiple Range Test.

Gamma rays	Days to 50% flowering	Panicle length	No. of panicles/ plant	1000 seed weight
Control	$81.20 \pm 0.14$ ef	$21.84 \pm 1.27$ gh	$2.0 \pm 0.27$ f	$2.47 \pm 0.01$ g
100 Gy	$81.00 \pm 0.22$ ef	$22.73 \pm 0.17$ fg	$2.0 \pm 0.27$ f	$2.67 \pm 0.02$ f
200 Gy	$81.50 \pm 0.82$ de	$24.40 \pm 0.16$ e	$2.7 \pm 0.22$ e	$2.76 \pm 0.01$ e
300 Gy	$79.90 \pm 0.48$ g	$27.10 \pm 0.70$ d	$3.2 \pm 0.34$ d	$2.87 \pm 0.04$ d
400 Gy	$80.60 \pm 0.70$ fg	$36.07 \pm 1.13$ c	$3.6 \pm 0.17$ c	$2.95 \pm 0.03$ c
500 Gy	$81.50 \pm 0.23$ def	$49.90 \pm 1.43$ b	$4.7 \pm 0.22$ b	$3.18 \pm 0.02$ b
600 Gy	$77.40 \pm 0.57$ h	$66.02 \pm 1.17$ a	$8.8 \pm 0.14$ a	$3.40 \pm 0.03$ a
700 Gy	$82.00 \pm 1.11$ d	$23.27 \pm 0.46$ ef	$2.0 \pm 0.27$ f	$2.69 \pm 0.08$ ef
800 Gy	$85.50 \pm 0.17$ c	$21.01 \pm 0.19$ h	$1.5 \pm 0.17$ g	$2.37 \pm 0.03$ h
900 Gy	$86.50 \pm 0.32$ b	$13.61 \pm 0.77$ i	$0.7 \pm 0.22$ h	$2.17 \pm 0.03$ i
1000 Gy	$89.40 \pm 0.17$ a	$7.86 \pm 1.21$ j	$0.5 \pm 0.17$ h	$1.72 \pm 0.08$ j

However, it decreased in seedlings raised from seeds treated with 700 Gy to 1000 Gy gamma rays with a maximum reduction in 1000 Gy gamma rays treatment ( $7.86 \pm 1.21$ ). The results revealed a maximum reduction in

seed weight in 1000 Gy gamma rays treatment ( $1.72 \pm 0.08$  g) as compared to control ( $2.47 \pm 0.01$  g). The maximum increase in 1000 seed weight was recorded in 600 Gy gamma rays treatment ( $3.4 \pm 0.03$  g). Pearson's correlation coefficients for phenotypic and yield-related traits of finger millet in different doses of gamma irradiation are given in Table 4. The correlation between the morphological and yield traits was significant and positive. (Table 4)

## Discussion

### Lethal Dose

The induction of mutations in plant breeding is a well-established tool to accomplish the objectives of crop improvement programmes (28). Prior to induction of mutations, the evaluation of  $LD_{50}$ , a mutagen dose that induces 50% reduction in seed germination is imperative. It allows the breeders to assess the mutagenic efficiency in relation to the mutagen induced biological damage (29). In the present study, the  $LD_{50}$  was determined in seven days old finger millet  $M_1$  seedlings. The maximum damage induced was recorded at higher gamma rays treatments. This may be attributed to the deleterious effects of higher doses of gamma rays on seed meristematic tissues that led to chromosomal aberrations, growth regulator disturbances, DNA damage and lethality. The reduced germination may also be due to inhibitory effects of gamma rays on vital functions of cells leading to cell and embryo fatality (30). A reliable indicator for the optimization of radiation dose requires the evaluation of  $LD_{50}$  and the effects of gamma rays on physio-morphological traits (31). The previous results of  $LD_{50}$  in the range of 599 - 731 Gy was reported earlier in pearl millet (32). A paper germination test to optimize  $LD_{50}$  depends upon germination percentage and reduction in root and shoot length. As compared to control, a reduction in root and shoot length has been previously reported in higher doses of gamma rays treatments (33). In the present investigation, higher doses of gamma rays affected seedling growth parameters and similar findings were also reported (34). The destructed cell components in seeds treated with higher doses of gamma rays lead to chromosomal damage and reduced seedling growth. Therefore, in induced mutagenesis, the determination of  $LD_{50}$  is important for the overall success of the breeding programme.

**Table 4.** Pearson's correlation coefficients for morphological and yield-related traits of  $M_1$  finger millet seedlings

	Plant height	No. of leaves/ plant	Leaf length	Tillers/ plant	Days to 50% flowering	Panicle length	No. of panicles/ plant	1000 seed weight
Plant height	1							
No. of leaves/ plant	.744**	1						
Leaf length	.703**	.817**	1					
Tillers/ plant	.725**	.728**	.907**	1				
Days to 50% flowering	-.739**	-.497**	-.641**	-.663**	1			
Panicle length	.666**	.812**	.922**	.876**	-.670**	1		
No. of panicles/ plant	.620**	.764**	.864**	.831**	-.672**	.924**	1	
1000 seed weight	.787**	.696**	.877**	.859**	-.798**	.857**	.810**	1

### Chlorophyll content

In the present investigation, the chlorophyll content gradually decreased with the increase in doses of gamma rays. This may be due to the enhanced production of free radicals at higher doses. In contrast, carotenoid contents increased linearly with the increase in gamma rays doses. The results were in agreement with the previous findings (35, 36) that also reported augmented carotenoid contents in mutant lines. The significant stable and steady increase in carotenoid contents indicate its potent role in photo protection. The enhanced carotenoid contents in higher doses may be attributed to their vital role in free radical scavenging activity during stress induced by radiations (37).

### Chlorophyll fluorescence

Chlorophyll fluorescence is extensively utilized to evaluate the relative influence of environmental stress on photosynthetic properties (38-44). From photosystem II the maximum amount of excited energy is converted into chemical energy and then utilized in carbon fixation. Even though the excess energy is converted into heat and a very little fraction is lost as fluorescence (45). Evaluation of CF provides insight into photosystem functioning. CF values observed in the present study show maximum peak values at 600 Gy gamma rays treatment.

### Quantitative traits

The results revealed a progressive decrease in the mean values of yield and yield attributing traits with the increase in gamma-ray doses. The diminution of mean values of yield attributes may be due to mutagen induced physiological and biochemical disturbances, and destruction of auxin and ascorbic acid content that may lead to inhibition in cell division and cell elongation (35, 36). The results were in agreement with the findings of the earlier studies (11, 46, 47). Our results revealed a progressive decrease in root and shoot length with an increase in gamma rays doses. The results were in agreement with previous findings in rice (48). Our findings showed that plant height correlated significantly in a positive direction with other agronomic traits such as number of leaves per plant, leaf length, number of tillers per plant, days to 50% flowering, panicle length, number of panicles per plant and 1000 seeds weight. These findings correlated with the results in rice that also reported plant height, flowering time and yield are the significant agronomic traits (49). The results also revealed that plant height, number of leaves per plant, leaf length, number of tillers per plant, number of panicles per plant, panicle length and 1000 seed weight decreased as the dose increased. The results were in line with the findings in sesame treated with ethyl methanesulphonate that showed a linear decrease in seedling emergence, seedling survival and delayed 50% flowering and maturity with the increase in gamma rays doses (50). The present study also revealed delayed seed germination and plant growth at higher radiation doses which may be attributed to late flowering in finger millet. Days to 50% flowering showed a significant positive correlation with the number of panicles per plant. The results were in agreement with previous studies that reports a significant positive correlation of pods with the days

to flowering in *Brassica juncea*, *Brassica napus* and *Brassica campestris* (51). Similarly, in linseed, plant height, seed yield, number of branches during primary and secondary stages, capsule number per plant were positively correlated (52). The results of the present study revealed that optimum doses of gamma rays stimulated growth regulators and enzymes associated with growth, yield and 1000 seed weight. This may be attributed to gamma rays induced morphogenetic, cytological, biochemical and physiological changes in cells and tissues (7, 53).

### Conclusion

*Eleusine coracana* (L.) Gaertn. (Finger millet) seeds irradiated at 600 Gy gamma rays dose revealed positive significant effects on physio-morphological and yield traits as compared to higher doses. Similarly, gamma irradiation also increased photosynthetic pigment contents at optimum doses (300 Gy, 400 Gy, 500 Gy and 600 Gy) compared to control. Therefore, it is concluded that the gamma-irradiated finger millet seeds with optimum doses improved the yield attributes of finger millet. In conclusion, lower doses of gamma rays treatment could be employed for the improvement of finger millet and other related crops.

### Acknowledgements

We are highly thankful to Periyar University, Salem, Tamil Nadu, India for providing the necessary laboratory facilities to perform the research work.

### Authors contributions

LS conceptualization, data curation, formal analysis, investigation, methodology, visualization, writing original draft, writing review and editing. AD conceptualization, data curation, formal analysis, investigation, methodology, visualization, writing original draft, writing review and editing. AG analysed and methodology construction. After the original draft was framed AR rewrote the article, analyzed and interpreted the data.

### Compliance with ethical standards

**Conflict of interest:** The authors declare no conflict of interest.

**Ethical issues:** None.

### References

1. Rachie KO. The millets: importance, utilization and outlook. 1975.
2. Annison G, Topping DL. Nutritional role of resistant starch: chemical structure vs physiological function. Annual Review of Nutrition. 1994 Jul;14(1):297-320. <https://doi.org/10.1146/annurev.nu.14.070194.001501>
3. Gee JM, Johnson IT, Lind L. Physiological properties of resistant starch. European Journal of Clinical Nutrition. 1992;46:125-31.
4. Lata C, Gupta S, Prasad M. Foxtail millet: a model crop for genetic

- and genomic studies in bioenergy grasses. *Critical Reviews in Biotechnology*. 2013 Sep 1;33(3):328-43. <https://doi.org/10.3109/07388551.2012.716809>
5. Bergamini N, Padulosi S, Ravi SB, Yenagi N. *Minor millets in India: a neglected crop goes mainstream*. Routledge: London, UK; 2013 Jun 26.
  6. Goud JV, Nayar KM, Rao MG. Radio-sensitivity in ragi (*Eleusine coracana*). *Canadian Journal of Genetics and Cytology*. 1969 Jun 1;11(2):254-65. <https://doi.org/10.1139/g69-032>
  7. Goyal S, Wani MR, Laskar RA, Raina A, Khan S. Assessment on cytotoxic and mutagenic potency of Gamma rays and EMS in *Vigna mungo* L. Hepper. *Biotechnology Vegetal*. 2019 Sep;19(3):193-204.
  8. Goyal S, Wani MR, Laskar RA, Raina A, Khan S. Mutagenic effectiveness and efficiency of individual and combination treatments of gamma rays and ethyl methanesulfonate in black gram [*Vigna mungo* (L.) Hepper]. *Advances Zool Bot*. 2020;8(3):163-68. <https://doi.org/10.13189/azb.2020.080311>
  9. Goyal S, Wani MR, Raina A, Laskar RA, Khan S. Phenotypic diversity in mutagenized population of urdbean (*Vigna mungo* (L.) Hepper). *Heliyon*. 2021 May 1; 7(5):e06356. <https://doi.org/10.1016/j.heliyon.2021.e06356>
  10. Ansari SB, Raina A, Amin R, Jahan R, Malik S, Khan S. Mutation Breeding for Quality Improvement: A Case Study for Oilseed Crops In: Bhat TA (ed.) *Mutagenesis, Cytotoxicity and Crop Improvement: Revolutionizing Food Science, 2021*; pp. 171-221, Cambridge Scholars Publishing, Lady Stephenson Library, Newcastle upon Tyne, NE6 2PA, UK.
  11. Khursheed S, Raina A, Khan S. Physiological response of two cultivars of faba bean using physical and chemical mutagenesis. *International Journal of Advance Research in Science and Engineering*. 2018;7(4):897-905.
  12. Goyal S, Wani MR, Laskar RA, Raina A, Khan S. Performance evaluation of induced mutant lines of black gram (*Vigna mungo* (L.) Hepper). *Acta fytotechn zotechn*. 2020 Jun 18;23(2):70-77. <https://doi.org/10.15414/afz.2020.23.02.70-77>
  13. Raina A, Sahu D, Parmeshwar K, Laskar RA, Rajora N, Soa R, Khan S, Ganai RA. Mechanisms of genome maintenance in plants: playing it safe with breaks and bumps. *Frontiers in Genetics*. 2021;12:861. <https://doi.org/10.3389/fgene.2021.675686>
  14. Raina A, Khan S, Wani MR, Laskar RA, Mushtaq W. Chickpea (*Cicer arietinum* L.) cytogenetics, genetic diversity and breeding. In: *Advances in Plant Breeding Strategies: Legumes 2019*; pp. 53-112. Springer, Cham [https://doi.org/10.1007/978-3-030-23400-3\\_3](https://doi.org/10.1007/978-3-030-23400-3_3)
  15. Raina A, Ansari SB, Khursheed S, Wani MR, Khan S, Bhat TA. Mutagens their types and mechanism of action with an emphasis on sodium azide and gamma radiations. In: *Mutagenesis, Cytotoxicity and Crop Improvement: Revolutionizing Food Science, 2021b* (1-37), Cambridge Scholars Publishing, Lady Stephenson Library, Newcastle upon Tyne, NE6 2PA, UK.
  16. Raina A, Laskar RA, Jahan R, Khursheed S, Amin R, Wani MR, Nisa TN, Khan S. Mutation breeding for crop improvement. In: *Introduction to Challenges and Strategies to Improve Crop Productivity in Changing Environment*. 2018a; 303-17, Enriched Publications. PVT. LTD, New Delhi.
  17. Wani MR, Tomlekova N, Raina A, Laskar RA, Khursheed S, Khan S, Tak MA, Bhat TA. Mutation breeding technique for the improvement of pulse crops with special reference to Faba bean (*Vicia faba*). In: *Mutagenesis, Cytotoxicity and Crop Improvement: Revolutionizing Food Science*. 2021;222-43. Cambridge Scholars Publishing, Lady Stephenson Library, Newcastle upon Tyne, NE6 2PA, UK.
  18. Raina A, Khan S. Increasing rice grain yield under biotic stresses: mutagenesis, transgenics and genomics approaches. In: *Rice Research for Quality Improvement: Genomics and Genetic Engineering 2020* (pp. 149-78). Springer, Singapore. [https://doi.org/10.1007/978-981-15-5337-0\\_8](https://doi.org/10.1007/978-981-15-5337-0_8)
  19. Raina A, Khan S, Sahu PK, Sao R. Increasing Rice Grain Yield Under Abiotic Stresses: Mutagenesis, Genomics and Transgenic Approaches. In: *Rice Research for Quality Improvement: Genomics and Genetic Engineering 2020* (pp. 753-77). Springer, Singapore. [https://doi.org/10.1007/978-981-15-4120-9\\_31](https://doi.org/10.1007/978-981-15-4120-9_31)
  20. Raina A, Laskar RA, Malik S, Wani MR, Khan S, Bhat TA. Plant Mutagenesis: Principle and Application in Crop Improvement. In *Mutagenesis, Cytotoxicity and Crop Improvement: Revolutionizing Food Science, 2021c* (38-65). Cambridge Scholars Publishing Lady Stephenson Library, Newcastle upon Tyne, NE6 2PA, UK.
  21. Kulandaivelu G, Noorudeen AM. Comparative study of the action of ultraviolet C and ultraviolet B radiation on photosynthetic electron transport. *Physiologia Plantarum*. 1983 Jul;58(3):389-94. <https://doi.org/10.1111/j.1399-3054.1983.tb04199.x>
  22. Zaccone C, D'Orazio V, Shotykh W, Miano TM. Chemical and spectroscopic investigation of porewater and aqueous extracts of corresponding peat samples throughout a bog core (Jura Mountains, Switzerland). *Journal of Soils and Sediments*. 2009 Oct 1;9(5):443-56. <https://doi.org/10.1007/s11368-009-0093-x>
  23. Coccozza C, Parente A, Zaccone C, Mininni C, Santamaria P, Miano T. Chemical, physical and spectroscopic characterization of *Posidonia oceanica* (L.) Del. residues and their possible recycle. *Biomass and Bioenergy*. 2011 Feb 1;35(2):799-807. <https://doi.org/10.1016/j.biombioe.2010.10.033>
  24. Maurya R, Prasad SM, Gopal R. LIF technique offers the potential for the detection of cadmium-induced alteration in photosynthetic activities of *Zea mays* L. *Journal of Photochemistry and Photobiology C: Photochemistry Reviews*. 2008 Mar 1;9(1):29-35. <https://doi.org/10.1016/j.jphotochemrev.2008.03.001>
  25. Buschmann C. Variability and application of the chlorophyll fluorescence emission ratio red/far-red of leaves. *Photosynthesis Research*. 2007 May;92(2):261-71. <https://doi.org/10.1007/s11120-007-9187-8>
  26. Arnon DI. Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant Physiology*. 1949 Jan; 24(1):1. <https://doi.org/10.1104/pp.24.1.1>
  27. Kirk JT, Allen RL. Dependence of chloroplast pigment synthesis on protein synthesis: effect of actidione. *Biochemical and Biophysical Research Communications*. 1965 Dec 21;21(6):523-30. [https://doi.org/10.1016/0006-291X\(65\)90516-4](https://doi.org/10.1016/0006-291X(65)90516-4)
  28. Arulbalachandran D, Mullainathan L, Velu S. Genetic variation in quantitative traits of black gram (*Vigna mungo* (L.) Hepper) induced by gamma rays treatment in M3 generation. *Journal of Phytology*. 2009 Oct 2;1(5). <https://doi.org/10.9755/ejfa.v2i12.5163>
  29. Raina A, Khursheed S, Khan S. Optimisation of mutagen doses for gamma rays and sodium azide in cowpea genotypes. *Trends Biosci*. 2018;11(13):2386-89.
  30. Olasupo FO, Ilori CO, Forster BP, Bado S. Mutagenic effects of gamma radiation on eight accessions of Cowpea (*Vigna unguiculata* (L.) Walp.). *American Journal of Plant Sciences*. 2016 Feb 3;7(2):339-51. <https://doi.org/10.4236/ajps.2016.72034>
  31. Ousmane SD, Elegba W, Danso K. Radio-sensibility of pearl millet (*Pennisetum glaucum* (L.) R. Br.) and cowpea (*Vigna unguiculata* (L.) Walp.) seeds germination and seedling growth. *International Journal of Innovation and Applied Studies*. 2013 Dec 1;4(4):665.
  32. Horn LN, Chikelou MB, Soleymanne B, Ipinge SN. Radiosensitivity studies in pearl millet (*Pennisetum glaucum*), Cowpea (*Vigna unguiculata*) and Sorghum (*Sorghum bicolor*) varieties in Namibia. *Agricola*. 2010;15(1):38-40.
  33. Ambavane AR, Sawardekar SV, Sawantdesai SA, Gokhale NB. Studies on mutagenic effectiveness and efficiency of gamma rays and its effect on quantitative traits in finger millet (*Eleusine coracana*)

- cana* (L.) Gaertn.). *Journal of Radiation Research and Applied Sciences*. 2015 Jan 1;8(1):120-25. <https://doi.org/10.1016/j.jrras.2014.12.004>
34. Kiong AL, Lai AG, Hussein S, Harun AR. Physiological responses of *Orthosiphon stamineus* plantlets to gamma irradiation. *American-Eurasian Journal of Sustainable Agriculture*. 2008 May 1;2(2):135-49.
  35. Laskar RA, Laskar AA, Raina A, Khan S, Younus H. Induced mutation analysis with biochemical and molecular characterization of high yielding lentil mutant lines. *International Journal of Biological Macromolecules*. 2018 Apr 1;109:167-79. <https://doi.org/10.1016/j.ijbiomac.2017.12.067>
  36. Raina A, Laskar RA, Tantray YR, Khursheed S, Wani MR, Khan S. Characterization of induced high yielding cowpea mutant lines using physiological, biochemical and molecular markers. *Scientific Reports*. 2020 Feb 28;10(1):1-22. <https://doi.org/10.1038/s41598-020-60601-6>
  37. Fukuzawa K, Inokami Y, Tokumura A, Terao J, Suzuki A. Rate constants for quenching singlet oxygen and activities for inhibiting lipid peroxidation of carotenoids and  $\alpha$ -tocopherol in liposomes. *Lipids*. 1998 Aug;33(8):751-56. <https://doi.org/10.1007/s11745-998-0266-y>
  38. Van Kooten O, Snel JF. The use of chlorophyll fluorescence nomenclature in plant stress physiology. *Photosynthesis Research* 1990 Sep;25(3):147-50. <https://doi.org/10.1007/BF00033156>
  39. Hendry GA, Grime JP (ed.). *Methods in comparative plant ecology: a laboratory manual*. Springer Science & Business Media; 1993 Jan 31. <https://doi.org/10.1007/978-94-011-1494-3>
  40. Samson G, Prášil O, Yaakoubd B. Photochemical and thermal phases of chlorophyll a fluorescence. *Photosynthetica*. 1999 Sep;37(2):163-82. <https://doi.org/10.1023/A:1007095619317>
  41. Maxwell K, Johnson GN. Chlorophyll fluorescence—a practical guide. *Journal of Experimental Botany*. 2000 Apr 1;51(345):659-68. <https://doi.org/10.1093/jexbot/51.345.659>
  42. Roháček K. Chlorophyll fluorescence parameters: the definitions, photosynthetic meaning and mutual relationships. *Photosynthetica*. 2002 Mar;40 (1):13-29. <https://doi.org/10.1023/A:1020125719386>
  43. Baker NR, Rosenqvist E. Applications of chlorophyll fluorescence can improve crop production strategies: an examination of future possibilities. *Journal of Experimental Botany*. 2004 Aug 1;55 (403):1607-21. <https://doi.org/10.1093/jxb/erh196>
  44. Netto AT, Campostrini E, de Oliveira JG, Bressan-Smith RE. Photosynthetic pigments, nitrogen, chlorophyll a fluorescence and SPAD-502 readings in coffee leaves. *Scientia Horticulturae*. 2005 Mar 30;104(2):199-209. <https://doi.org/10.1016/j.scienta.2004.08.013>
  45. Seaton GG, Walker DA. Chlorophyll fluorescence as a measure of photosynthetic carbon assimilation. *Proceedings of the Royal Society of London. Series B: Biological Sciences*. 1990 Oct 22;242 (1303):29-35. <https://doi.org/10.1098/rspb.1990.0099>
  46. Ojiewo CO, Agong SG, Murakami K, Tanaka SG, Hase Y, Masuda M. Male-sterility induced by gamma-ray irradiation of African nightshade (*Solanum nigrum* L. ssp. *villosum*) seed. *The Journal of Horticultural Science and Biotechnology*. 2005 Jan 1;80(6):699-704. <https://doi.org/10.1080/14620316.2005.11512001>
  47. Ojiewo CO, Agong SG, Murakami K, Tanaka A, Hase Y, Masuda M. Biological effects of carbon-ion beam on induction of male-sterility and a novel season-dependent floral homeotic mutant in *Solanum villosum* Miller. *The Journal of Horticultural Science and Biotechnology*. 2006 Jan 1;81(4):559-64. <https://doi.org/10.1080/14620316.2006.11512105>
  48. Benjavad Talebi A, Benjavad Talebi A. Radiosensitivity study for identifying the lethal dose in MR219 (*Oryza sativa* L. spp. *indica* cv. MR219). *International Journal of Agricultural Science, Research and Technology in Extension and Education Systems*. 2012 Jul 1;2(2):63-68.
  49. Xue W, Xing Y, Weng X, Zhao Y, Tang W, Wang L, Zhou H, Yu S, Xu C, Li X, Zhang Q. Natural variation in Ghd7 is an important regulator of heading date and yield potential in rice. *Nature Genetics*. 2008 Jun;40(6):761-67. <https://doi.org/10.1038/ng.143>
  50. Sheeba A, Ibrahim SM, Yogameenakshi P, Babu S. Studies on induced chlorophyll mutation in sesame (*Sesamum indicum* L.). 2004.
  51. Mahto RN, Lal Mahto J. Correlation, regression and path coefficient analysis in rainfed linseed. *Madras Agricultural Journal*. 1997;84:84-86.
  52. Khan S, Farhatullah R, Khalil IH, Khan MY, Ali N. Genetic variability, heritability and correlation for some quality traits in F3: 4 Brassica populations. *Sarhad J Agric*. 2008;24(2):223-31.
  53. Jan S, Parween T, Siddiqi TO. Effect of gamma radiation on morphological, biochemical and physiological aspects of plants and plant products. *Environmental Reviews*. 2012 Mar; 20(1):17-39. <https://doi.org/10.1139/a11-021>

§§§