



REVIEW ARTICLE

CO₂ sequestration: microalgae genome analysis and its application of effective green source technology

R. Ranjith Kumar^{*}, S. Monika Jency, G. Bhavatarini & M. Jeslyn Priscilla Devapriya

Department of Botany, Madras Christian College (Autonomous), Affiliated to The University of Madras, Chennai - 600 059, Tamil Nadu, India.

*Email: ranjithkumar@mcc.edu.in

ARTICLE HISTORY

Received: 07 July 2021 Accepted: 16 January 2022 Available online Version 1.0 (Early Access): 14 March 2022

Check for updates

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/ journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, etc. See https:// horizonepublishing.com/journals/index.php/ PST/indexing_abstracting

Copyright: © The Author(s). This is an openaccess article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (https://creativecommons.org/licenses/ by/4.0/)

CITE THIS ARTICLE

Kumar R R, Jency S M, Bhavatarini G, Devapriya M J P. CO_2 sequestration: microalgae genome analysis and its application of effective green source technology. Plant Science Today (Early Access). https://doi.org/10.14719/ pst.1365

Abstract

Microalgae genome technology for CO₂ sequestration is an appropriate vehicle for articulating the importance of the current need and solution for reduction of CO₂ at the atmospheric level. In comparison with C4 plants, microalgae have greater capability to fix atmospheric CO₂. The rate of CO₂ fixation differs in different strains of microalgae. The photosynthetic enzyme RuBisCO is widely responsible for photosynthetic carbon assimilation in all plants including phototrophic algae. The gene rbcL encodes this enzyme. The catalytic activity of carbonic anhydrase achieves the CO₂ generation in the RuBisCO. CAH3 gene is essential for generating CO₂ concentration for RuBisCO by dehydration of accumulated inorganic carbon. There are also few other microalgae genes which involves for carbon assimilation. Genomic resource databases and several other nucleotide databases are being used for sequencing the microalgal genomes. Even though, recent advances in genomic studies are providing thrust to enhance the research on microalgal species, they are expensive and resources available for microalgal genomic studies are limited. This review article attempts first as a combined revise on microalgae CO₂ sequestration in the field of basic science, applied aspects, and the role of specific gene(s) in the algal system is well defined which could be a supportive involvement of carbon dioxide reduction as "Green-Gene Technology". This Green biotechnology could be used for Global warming reduction as well as creating wealth from the waste through valuable by-products from the selected microalgae strains in future.

Keywords

carbon assimilation, carbonic anhydrase, genomic resources, microalgal genomes, RuBisCO

Abbreviations

IPCC- Intergovernmental Panel on Climate Change, MOFs- metal organic frameworks, CS-H - Calciumsilicate-Hydrate, Pg C - petagram carbon, AllnGaP II - aluminum indium gallium phosphide, CCM – CO2 concentrating mechanism, GHG-Greenhouse gas, ROS - reactive oxygen species, NGS - Next Generation Sequencing, CRISPR-Cas9 - Clustered Regularly Interspaced Short Palindromic Repeats-Crispr associated protein 9, RuBisCO -EPYC1-RuBisCO-Essential Pyrenoid Component 1, SBPase -Sedoheptulose-1,7-bisphosphatase, FBPase -Fructose 1,6-bisphosphatase, GGT- Green Genome Technology. TALEN-Transcription Activator-Like Effector Nucleases.

Introduction

Climatic change is one of the long term processes which causes changes in weather patterns. It will determine the local, global and regional climate of an earth's surface. Climate change is influenced by the utilization of fossil fuels and greenhouse emissions through deforestation, agriculture and other protuberant causes (1). Climate change occurs naturally due to fluctuation in solar irradiance, discrepancy in orbital parameters of earth and volcanic activities. Generally, some amount of solar energy that enters the earth reflects to the space. Atmospheric gases trap some amount of outgoing solar energy, due to which the temperature gets warmer. If natural heat grabbing/grasping/deceiving properties are unavailable, then the earth's average surface temperature would be less than 33°C. Heat energy is deceived by some amount of gases called greenhouse gases (GHG). In recent decades, especially after the industrial revolution, a sharp increase in atmospheric greenhouse gases is evident mainly due to human intervention. A sharp level of increase in the number of greenhouse gases in the atmosphere results in the increase of earth temperature and alteration of energy which is referred to as global warming (2). The main sources of global warming are anthropogenic activities and the emission of greenhouse gases. A conflict arose when there was a sharp increase of atmospheric greenhouse gases due to several activities of humankind at an alarming rate for the past two centuries. Based on human enhanced global warming effect, according to 2004, about 8 billion tons of carbon dioxide were pumped and the effect of thermal radiation was obstructed due to increased level of GHG. Over the last century, the planet has experienced a high amount of increase in surface temperature. From the years 1906 to 2006, the average temperature of earth's surface was about 0.6 to 0.9 °C, but in the in past 5 years the level of earth temperature was doubled. During the 20th century a rise of about 0.17 m was evident in sea levels. These changes in temperature changes primarily due to the presence of greenhouse gases emitted from various sources. An enormous amount of methane is produced in landfills, animal ordure and agricultural decomposition. Various nitrogen- based fertilizers extricate nitrous oxide into the atmosphere including urea, diammonium phosphate. Once they are discharged, these GHG remain in the atmosphere for decades. According to IPCC 35% increase in CO₂ and 148% increase in methane levels are evident since 1750. Among those GHG, CO₂ gas is at the topmost level; a high amount of CO₂ is released from burning fossil fuels (3). Thus, there is a need to reduce atmospheric carbon dioxide. Researchers, scientist and government throughout the world have been trying to find effective CO₂ sequestration methods by various innovative ways. Currently, this issue is in limelight, which urges researchers to find out effective technologies to capture the major greenhouse gas carbon dioxide. Amid those innovative techniques, microalgae green genome technology plays crucial role in targeting gene for CO₂ sequestration by the activity of microalgae.

The Current scenario in CO2 sequestration - worldwide

Metallic organic frameworks

Metallic organic frameworks are one of the auspicious methods in CO_2 mitigation i.e. Carbon Capture and Storage (CCS). For this method, materials having high adsorptions and storage capacity are needed to be used. The adsorbent materials utilised to capture CO_2 from flue gas should possess chemical stability, easy productivity with minimal energy utilization- and be economically feasible. Metal Organic Frameworks (MOF's), highly crystalline porous materials constructed by metal ions and organic ligands, proved to be a meritorious adsorbent material for carbon capture. However, eminent advancement in MOF materials for CO_2 capture has been emerged in the past, assessed accordingly, but new inventions are constantly exposed as the field widens quickly (4).

Industrial carbon capture

In industries, carbon capturing techniques are divided into four major techniques, which includes; pre-combustion, post-combustion, oxy-fuel combustion and electrochemical separation. For reducing CO2 emissions several new power generation concepts have been developed recently. The new concepts also adopt the techniques such as precombustion, post-combustion, chemical looping combustion and oxy-fuel combustion (5-7). (I) In pre-combustion capturing technology, a new gasification technique can be used in order to capture the CO₂ prior to burning to generate the power and also to produce combustible gas. The rate of CO₂ concentration is high in the pre-combustion process, which creates a greater driving force and leads to the separation of CO₂. Though it is a well-described enabling technology, it requires equipments like water gas shift and gasification reactors for capturing process though it is a well described enabling technology. (II) The post-combustion is one of the methods which has high potential to retrofit them to the coal power plants (8-13). In the post-combustion process, the CO₂ is captured when the fossil fuel is completely burnt. For capturing NOx (Nitrogen oxides) and SOx (Sulfur oxides), the CO₂ capturing post-combustion technology can be retrofitted to the coal-burning power plants. Absorption, adsorption and membranes are the most important post-combustion CO_2 capture technologies. There are also there other technology necessary for the post-combustion process, it includes absorption, adsorption and membrane filters (III). Due to loss of CO₂ during the process of absorption and solvent regeneration, the post-combustion method requires high energy to complete the process. Oxy-fuel combustion is the process of burning fossil fuel in oxygen-rich gas; it results in the emission of steam and CO₂ which leads to an easier process of capturing CO₂. In this process, a high concentration of CO₂ in flue gas occurs, and due to the absence of nitrogen, the emission of NOx is eliminated. Due to NOx elimination, the combustors are smaller, which collects only a small amount of gas. Therefore pure oxygen combustion is complicated than air combustion. The process requires large cryogenic air separation units because it avoids high-temperature combustion and 60% of flue gas is recycled back to the combustor. Advanced oxygen

separation membranes can be used to lower the energy long term storage of industrial effluents containing carbon, requirements (14).

CO₂ uptake mechanism in cement-based materials

In cement-based materials the carbonation reaction occurs chemically between CO₂ and cement hydrates, which is an The term abiotic sequestration refers to the process in essential reaction for CO₂ uptake by a cement-based material. The process of carbonation is considered as a weathering degradation of the cementitious composite, because of of sequestration is performed by using only engineering the possibility of the decomposition of CS-H (calcium sili- techniques, physical and chemical reactions. In comparison cate-hydrate) phases in the cementitious composites (15). with biotic sequestration, abiotic sequestration has a great-Owing to these factors, the process of carbonation was con- er sink capacity which has received considerable attention sidered as a negative influence affecting the durability of in geological and oceanic structures (20) (21). Prompt studconcrete (16). However, studies carried out over the years ies are being carried out in developmental and experirevealed that, the process of carbonation does not always mental technologies for capturing, transporting and injecthave a negative influence as it may also contribute to en- $ing CO_2$ (22).

which in-turn aids in reducing the emission of carbon into the atmosphere in the form of $CO_2(1)$ in Fig. 1.

Abiotic sequestration

which sequestration is carried out without the intervention of living organisms such as plants and microbes. This type

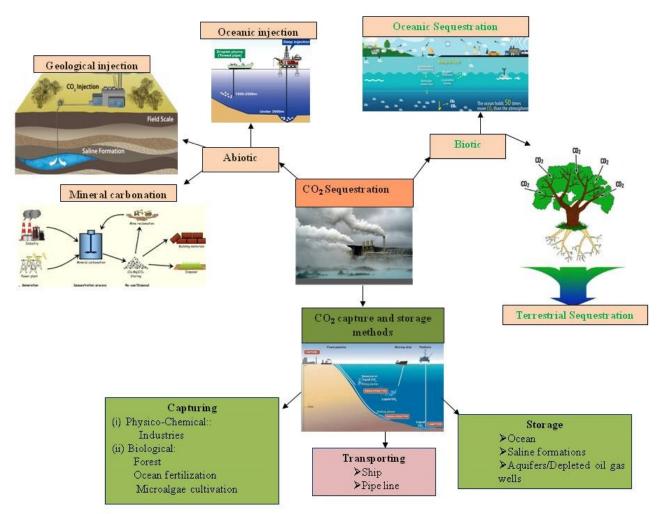


Fig. 1. General types of CO₂ sequestration and its capture methods.

hancing the durability as well as the mechanical properties of the material. As a result, several researchers have suggested that the source material could be potentially used for CO₂uptake (16-19). Even though an adequate level of understanding has been obtained in the carbonation mechanism of cement-based materials, the distinctiveness and differences in the intrinsic carbonation mechanism should be studied in detail and summarized in comparison to other CO₂ uptake methods (17).

Types of carbon sequestration

Oceanic injection

In the late 1970s, the initial proposal of injecting CO_2 into the oceans was started; after that, substantial progress of injecting CO₂ into the ocean was made. This technique requires the injection of CO2 at great depths into the ocean to stabilize and minimize the gassing and it has been widely practised by engineers for about three decades. Techniques used to inject liquefied CO₂ obtained from industrial sources are as follows; liquefied CO₂: (i) should be lighter than water and has tends to rise approximately to 1000 m depth, forming a droplet plume and hence is injected below The process of carbon sequestration involves capturing and 1000 m from a manifold lying on the ocean floor, (ii) it

mixture of CO_2 and seawater which helps it to sink deeper veloping a clear effective technology. into the ocean, (iii) discharged into the ocean using large pipes dragged behind ships, and (iv) pumped onto the bottom of the ocean floor into depression causing the formation of a CO₂ lake. The injection of liquefied CO₂ at approximately 3000 m depth into the ocean helps to achieve stable oceanic sequestration (23). The estimated value of carbon sequestration by oceanic sink capacity is 5000-10000 Pg C (peta-gram carbon), which helps to decrease the net CO_2 emission released into the atmosphere (21).

Geological injection

This technique involves the injection of industrial CO₂ into the geological layers, which includes places such as saline aquifers, old oil wells- to increase yield, stable rock strata and coal seams (24-26). This entire process starts from the capture, liquefaction, transportation and injection of CO₂ into the deep geological layers. There are many concerns raised regarding the geological sequestration which includes; (i) leakage of CO_2 , (ii) safety concerns regarding the storage of vast quantities of CO₂, and (iii) the cost of the whole process. Saline aquifers are located in the underground strata below the freshwater reservoirs. They are made up of very porous sediments filled with brackish (saline) water (27, 28). Despite the concerns, people have argued that the risks of leakage is low by the presence of an impermeable layer in between the saline aquifers and the freshwater reservoirs. Hydrodynamic sequestration takes place after the CO₂ is pumped into the saline aquifer which reacts with dissolved salts and forms carbonate. The CO₂ which is injected into the sea displaces the liquid brine because of its lower viscosity and density. Therefore the CO₂ is injected in supercritical state. А multiphasemulticomponent environment is created when the CO₂ is injected in situ as it dissolves in the aqueous phase and forms a gas-like phase. An economical strategy to enhance oil recovery (EOR) is injecting CO₂ into the oil or gas reservoirs where the CO₂ displaces the gas and oil. This process of CO₂ enhanced recovery has been found to be advantageous to increasing the oil production and gas fields (29). Despite the drawbacks and concerns, this technique of CO₂ sequestration is being carried out in the United States, within the state of Texas, to inject 20 million Mega gram (Mg) of CO_2 yr⁻¹ at a price of \$10 to \$15 Mg⁻¹ per year (30).

Scrubbing and mineral carbonation

Scrubbing is a common method used for capturing carbon. This method involves the chemical absorption of carbon dioxde using a carbonate solvent or an amine (21). Further, the captured CO₂ is purified by passing it through an absorption column containing solvents like amine, nickel and other elements such as ceramic-based compounds, lithium silicate, and K₂CO₃. By passing the captured CO₂through a absorption column having solvents like nickel, amine and elements such as ceramic-based compounds like lithium silicate and K_2CO_3 , the CO_2 is filtered further (31). The CO_2 gas is transformed into mineral carbonates such as MgCO₃, CaCO₃ and other minerals, which are thermodynamically and geologically stable. However, as the mineral carbonates formed are stable rocks, the CO₂ sequestered for-

should be injected at a depth of 500-1000 m with a denser ever within them need to be concentrated more on for de-

Biotic sequestration

Arbitration of higher plants and microorganisms in removing CO₂ from the atmosphere is known as biotic sequestration. This process varies from managing process, but it helps in the reduction of offset emissions. Biotic resources like water and energy are used in managing the terrestrial C pool process. Few other types of biotic sequestration are explained below.

Oceanic sequestration

Carbon sequestration in the ocean is achieved using biological processes, mainly photosynthesis. Phytoplankton is one such mechanism (32), which fixes approximately 45 Pg C yr⁻¹ (Peta gram carbon stock per year) (33). From the phytoplankton, several organic materials get deposited on the ocean floor and get sequestrated (34). Among those organic factors, 'Fe' acts as a limiting factor, therefore various studies are insisted to find out the significance of the 'Fe' factor on biotic sequestration of CO_2 in the ocean (33, 35, 37).

Terrestrial sequestration

Terrestrial carbon sequestration involves the transfer of atmospheric CO₂ into biotic and pedologic Carbon pools. In the atmosphere 8.5 Pg C yr⁻¹ is emitted but anthropogenically emitted 3.5 Pg C is only retained in the atmosphere which owe to terrestrial carbon sinks and the sequestered CO₂ plays a crucial role in the global C cycle. In terrestrial sequestration, the sequestration process takes place in terrestrial ecosystem where the C sink occurrs through photosynthesis and the CO₂ is stored in living and dead organic matter. Terrestrial carbon sequestration can also be called as win-win or no-regrets strategy (38) due to various benefits like improving the quality of soil and water, increasing the yield of the crop and restoring the degraded ecosystem. This type of sequestration provides numerous benefits without affecting the global climate (21).

CO₂ capture and storage methods

Physicochemical carbon capture and sequestrations strategies are collectively assorted as carbon capture and storage (CCS) methodologies. It operates by three major steps which includes CO₂ capture, CO₂ transportation, CO₂ storage. CO₂ is captured from huge sources like power plants and cement manufacturing plants. Consecutive methods are adopted for the capture and separation of CO₂ from other exhaust components such as chemical absorption, physical adsorption, membrane separation, cryogenic distillation (39-41). The highly concentrated CO₂ collected is then constricted and transited through pipelines or shipped to repositories (42, 43). Finally the acquired CO_2 is gathered into reservoirs, such as geological storage, oceanic storage wherein the CO₂ is directly inoculated deep into the ocean, saline formations, aquifers and depleted oil or gas wells (30). The economically high operation, conveyance and environmental threat of long term CO₂ leakage and other uncertainties (44, 45) are some of the major drawbacks.

Physicochemical CCS methods are victorious in trapping CO2 from point sources generating high concentrations of carbon di oxide. But the sources from diffused, non- microalgae to photo acclimate to different intensities of iron and other nutrients promoting increased carbon diox- Phosphide) having 613 mm of wavelength, halogen, fluocultivation (44, 48, 50-52).

The Critical role of microalgae in regulating atmospheric carbon dioxide sequestration

The oldest and significant group of organisms on earth is microalgae. Microalgae is photoautotrophic primitive plant that shows rapid growth and ranges few microns in size. Many algal species have oil substances in it and their weight ranges between 20-50% of dry biomass. 1kg dry algal biomass make use of 1.3 kg of CO₂(53, 54) Approximately half of the atmospheric oxygen on the earth is produced by microalgae, while consuming vast amounts of the greenhouse gas CO₂. Since algae produces huge amount of energy and biomass, it is used as a source of food, feed, fuel, stabilizing agent and waste water treatment. Algae generate a large amount of biomass and energy. Due to its rapid growth ability it has the efficiency to fix CO₂ which is ten folds better than terrestrial plants. The potential of photoautotrophic algal cultures play a major role in diminishing the release of atmospheric CO₂ which help in alleviating the trend towards global warming. The selection of optimal microalgae is crucial to recognize workable CO₂ biological fixations systems. Specific strategies required for CO₂ sequestration influence the selection of optimal microalgae. The production of biomass of microalgae have higher potential to reduce the carbon dioxide emissions and increase the level of world energy supply. Microalgae has greater potential to tolerate high temperature and CO₂, therefore nowadays carbon sequestration is done using microalgae with maximum efficiency. Microalgae can assimilate CO₂ within various ranges of concentration by selecting competent species. The use of microalgae for CO₂ sequestration also yields several byproducts and shows versatile performance (55, 56).

CO₂ tolerating microalgal species based on parameter

Light and Light sources

Biological processes such as microalgal growth, photosynthesis, carbon dioxide fixation etc., are highly dependent on the intensity of light (57, 58). In Calvin-Benson cycle, the enzyme RuBisCo (1,5 Bisphosphate Carboxylase Oxygenase) fixes the CO₂ by using the ATP (Adenosine triphosphate) and NADPH (Nicotinamide adenine dinucleotide phosphate) which are produced in the light reactions during photosynthesis. High light intensity produces high reactive oxygen species which can cause abnormal physiological reactions due to oxidative stress, while in low light intensity CO₂ fixation and biomass concentration are low (59). For the dispensation of different dissolved inorganic spe-These high reactive oxygen species cause the decline of CO₂ cies (CO₂, HCO₃-,CO₃²⁻), the pH is regulated on the chemical during photosynthesis because it affects the essential pro- level. It was reported that pH is an extensive factor that

point emissions and low concentrations of CO₂ cannot be light helps them to carry out the usual metabolic reactions captured (46, 47). Besides physical and chemical CCS, there and physiological processes (60). Different species have is a biological route that capture CO_2 through natural sinks. different photo acclimation periods (61, 62). One of the Biological capture methods includes; i) forestation; affor- main factors for the cultivation of microalgae is light. It can estation, reforestation, and the farming of crops and live- be both natural and artificial. Light sources such as lightstock (38, 48). (ii) ocean fertilization; fertilizing oceans with emitting diodes, AllGaP II (Alluminium Indicum Gallium ide uptake by the phytoplanktons (49) and (iii) microalgae rescent, incandescent etc. can also be used, out of which usage of AllnGaP II and Light-emitting diodes are efficient and Cost-effective (63). Recent studies have found new sources of illumination and controlling the intensity of light which includes; (i) illumination with selected wavelengths of light, (ii) use of dye compounds which helps to lower the energy of photons, and (iii) light filters such as nanoparticles and fluorescent paints (64). Research on the cultivation of microalgae have been carried out using artificial light under controlled conditions as well as large scale outdoor cultivation using solar light, both depending on the location, climate, season and circadian cycle (65). To The exposure to light can be both continuous or with interrupted photoperiods. A successful cultivation system is a system that produces high biomass along with high cell concentrations which causes different gradients of light in the medium and the cells are exposed to different light intensities which in turn helps to minimize the cost of production (66, 67). To achieve a decent amount of CO₂ fixation, the cells should be exposed to a certain period of dark-light cycles with a ratio of 18:6 or 12:12 cyclic patterns (64). The algal growth and its ability to fix CO₂ is affected by either the light intensity or through the light-dark cycle (58).

Temperature

In the cells, the metabolic processes and the availability of physicochemical CO_2 is influenced by temperature (68). Carbon dioxide solubility is indirectly proportional to temperature; it is identified that when the temperature is (>20 ^oC) lower is the CO₂ solubility. In spite of that, when temperature increases the RuBisCO's affinity for carbon dioxide decreases (69). Nevertheless, the effect of temperature on the metabolic reaction rate is strain-dependent (70) experimentally reviewed that the effect of temperature using different species, by indicating adaptations of different species to high temperature and biochemical effects. Generally, under the temperature between 15-30 °C with optimal values at 20-25°C the microalgae grows.

Mechanisms of carbon fixation and metabolic activity are affected below 16°C. On the other hand, the higher the temperature lower is the carbon dioxide solubility (71) and many species will die when the temperature is above 35 °C (72), and it also decreases the growth rate by increasing both respiration and photorespiration. Light and temperature are considered as the greatest important factors which affects the biomass productivity and CO₂ fixation.

pН

teins which are needed for electron transfer. The ability of determines algal growth by affecting the various enzyme

condition is dissolved inorganic carbon species, CO₂ dissolution and CO₂, nitrate uptake cause fluctuation in pH. However, CO₂ input concentration influence these changes. Usage of HCO₃ causes the zin metalloenzymes (carbonic anhydrase) to convert into CO₂ which results in liberation of OH (hydroxyl ions) and increase in pH. It shows that limitations in CO₂ effects the performance of the system, but supplying CO₂ will control the optimum level of pH in the culture. Reports are on maintaining an optimum level of pH by supplying CO₂rich gases to the culture (80). This method was used widely in order to meet the carbon demands for micro algae cultures which shows a high rate of CO₂ fixation and huge production of biomass.

Dissolved O₂ concentration

For measuring photosynthesis activity in microalgae, dissolved O₂ are used; the resulting value is greater than the values obtained under the equilibrium condition of air. Dis-

activity (73, 74). Moreover, the value of pH has a powerful against with CO₂ help rubisco for fixing CO₂. The concentraimpact on the growth of cells and the level of optimum or tions of O_2 and CO_2 determines the process of photosynthetolerating level are species-dependent. The optimum pH is sis (carboxylation) and photorespiration (oxygenation). in the range between 6-8.3 which is neutral to slightly alka- Low ratio level of O_2 /CO₂ diminish the photosynthetic rate line (75-79). Growth takes place, there will be equilibrium in micro algae. Thus, CO_2 fixation favours photorespiration CO_2 release (58).

CO₂ concentration

CO₂ fixation includes solubilization (from gas to liquid phase) and mass transfer, ionic equilibrium condition (CO₂, HCO_3 -, CO_3^2 -) and carbon ingestion by the microalgal cells. The strain has a direct influence on tolerance and optimal CO₂ concentration (82) and most microalgae have the potential to grow well at 2% CO₂, but levels above 5% CO₂ (83) may cease their cell growth (84, 85). This effect may result in acidification of the stroma in the chloroplast (82) and abolition in important enzymes involved in Calvin-Benson cycle. It was reported that rate of CO₂ fixation is high in different micro algae like Nannochloropsis oculata, Botryococcus braunii, Scenedesmus obliques, Chorella vulgaris and Synechococcus sp. by the effect of different level of CO_2 (86). However, it was found that the maximum biomass production obtained in painting the CO₂ tolerant species solved O_2 are mostly consumed by heterotrophic microor- like Scenedesmus sp. (80%), Euglena gracilis (45%) and

Table 1. CO₂ tolerating microalgal major contributing families containing species under optimum condition

Major CO₂ tolerating Family	Microalgae	Tempera- ture (ºC)	lrradiance (µmol m-2 s-1)	рН	CO ₂ tolerance (%)	Conical flask / Photobioreactor	Growt h rate (day- 1)	Biomass productivity (g L–1 day–1)	CO2 biofixation (mg L-1 day-1)	Mixing	References
	Chlorella sp	27	100	7	40	Conical flask, 8 days continuous aeration, air + CO ₂ ; horizontal bub- ble column, 11 days continuous aeration	0.38	0.09	7.2		(90, 91)
	Scenedesmus almeriensis	35	200	7-8	10	Tubular photobioreactor outdoor condition flue gas (pure CO_2) on demand operated continuously	0.34	0.42	790		(58)
Chlorophyce e	a Scenedesmus obtusiusculus	35	300	7-8	10	Bubble column, 14 days, continu- ous aeration, air + CO ₂	0.34	0.52	970ª	Aeration	(58, 93)
	<i>Dunaliella</i> sp.	25	100	8	5	Horizontal bubble column, 11 days	0.25	0.12	10.4		(91)
	Dunaliella salina (DCCBC2)	27	80	8	10	continuous aeration aeration N_2 + CO_2	0.42	0.13	8.2		(91)
	Haematococcus pliviallis	25-28	90	7	34	Bubble column, aeration, air $+ CO_2$	1.29				(94-96)
Bacillari- ophyceae	Phaeodactylum triscornutum	20.4	10	7-8	15	Erylenmeyer flask, aeration Roux bottles 11 days, continous aeration air + CO ₂		0.15	280ª		(97)
Cyanophycea	Spirullina a platensis	30	330	9-10	10	Bubble column, 25 days, continu- ous aeration	0.65	0.15	0.15 280° (S	(98-101)	
e	Spirullina maxi- ma	35		9-10		Container of glass, 15 days, me- chanical agitation	0.6	0.15	280ª		(60, 99)
<u>Euglenaceae</u>	Euglena gracilis	27-31	100	7.8	45	Photobioreactor, 28 days flue gas 11% CO ₂ continous operation	0.31	0.29	74	Fermentor	(102, 103)

Source adopted and modified (65).

^aCalculated from the biomass productivity, according to the equation: CO_2 fixation rate (P_{co2}) = 1.88 × biomass productivity (g L^{-1} day⁻¹), which is derived from the molecular formula of microalgal biomass, $CO_{0.48}$ H_{1.83} N_{0.11} P_{0.01}

growth of micro algae. Dissolved oxygen may exceed 250 % ranges between 5%-20% (87). Maximum growth at 10% CO₂ of saturation during daytime (58, 81). Excessive concentra- was seen in Chlorella sp. KR-1 and tolerated up to 70% CO2 tion of oxygen causes oxidative stress in ROS and content (88), Spirulina sp. (MCRC-A0003) maximum growth and 30-

ganisms and low dissolved O_2 point out complication in the *Chlorella* sp. T-1(100%) when the concentration of CO_2

50 % CO₂ reduction (89). The above-mentioned range is rophyll dimer (117), (118). In the light reaction the cells are grow well in 10 to 15% of CO₂ this type of CO₂ range is generally found in flue gas even though flue gas is one of the GHG causes pollution but for micro algae it is really good carbon source (Table 1).

CO₂ tolerance of microalgae

In a CO₂ sequestration system the flue gas is directly utilized from the power plants (104) and the cost of separating CO₂ becomes minimum when flue gas is used. High CO₂ tolerant species is adequate when the power plant flue gas contain high concentration of CO₂. There are a few microalgae such as Cyanidium caldarium (105) and some other species of Cyanidium has the capability to grow in pure CO₂ (106). However the micro algae can grow under concentrations 5% to 45 % of CO₂ better growth was observed in 5% concentration and the micro algae cannot grow under concentration of CO₂ higher than 45% (102). Scenedesmus sp. concentrations of CO₂. CO₂ acts as a carbon source for the reaction or carbon-fixation cycle is the second stage of phoimportant point that should be taken into consideration is ty of the enzyme along with the utilization of ATP molethat must be worked out is the supply of carbon to microalgal mass culture systems (111-113).

CO₂ tolerance mechanism

The mechanistic presumption of the effect of the increased CO₂ concentration on the growth and efficiency of algae was analysed or studied or examined earlier (90, 114, 115). Analysis reveals that Dunaliella tertiolecta cells enclose more starch granules with a well-developed pyrenoid under ordinary air than cells grown under high CO₂ cells. The chloroplast was detected neighbouring to PM and the envelope was denser electronically under ordinary air while that in enriched air, the chloroplast were found in the inner area of the cells and the envelope was electronically lighter. The effect of CO₂ analysed in the chloroplast cells was contradictory when it was observed or studied under plasma membrane (116). This implies that microalgae have the potential to endure high concentration of CO₂ by altering their anatomical structure and reorganisation of certain cellular organelles (104, 114).

Role of CO2 in algae physiology, biochemical pathway and its fixation

CO₂ fixation in microalgae takes place in two phases (i.e.) the light reaction which includes photoactive complexes called photosystem I (PS-I) and photosystem II (PS-II) and the dark reaction also called as the Calvin cycle. The PS-I and PS-II are responsible for transferring the light energy into the electron transport chain through the excited chlo-

significant because some micro algae have the ability to illuminated by the utilization of light energy to form energystorage molecules ATP and NADPH, which are used to capture and reduce CO_2 (117).

> In the PSII complex, the photosynthetic process takes place when the core P680 chlorophyll dimer reaches adequate excitation energy from the sunlight. And the excited electron is transported to the primary electron acceptor molecule. This course of the process is called photoinduced charge separation. The excited electrons are transported through an electron transport chain to the P700 chlorophyll dimer present in the PS-I. The excited electrons from the P700 chlorophyll dimer are further oxidized into ferredoxin and NADPH by the light excited antenna (118-120).

Energy harvested by means of the light reaction can be used for the phosphorylation of ADP to form ATP, which is called photophosphorylation (121). During the electron has the capability to grow in 80 % of CO₂ concentrations its transport chain each pair of the electron produces 1.3 ATP maximum cell growth was observed under 10 % to 20% molecules (121). The Calvin cycle also called as the dark group development of micro algae. Control the pH (107) of tosynthesis which can function both in the presence and the culture is contributed by CO₂ supply (108). Chemical absence of light (117, 121). RuBisCO enzyme present in the analysis of algal biomass shows that for 1 kg production of Calvin cycle carries out both carboxylase activity and oxybiomass about 1.5 to 2.0 kg of CO_2 is required (109). The genase activity (low affinity for CO_2). The carboxylase activithat CO₂ should not be allowed to reach higher and lower cules, converts CO₂ into sugar (Fig. 2). Glycolate -2concentrations (107). The lowest limitation and highest phosphate is formed as an end product through the activity inhibition concentration vary in different species ranging of the enzyme oxygenase. This glycolate-2-phosphate is between 2.3 x 10⁻² M to 2.3 x 10⁻⁴ M (110). According to previ-responsible for the liberation of CO₂ fixed in the carboxylase ous studies, one of the primary difficulties and limitations activity of RuBisCO and its synthesis consumes significant amounts of cellular energy and this end product is not utilized by the cell.

> The microalgae biomass production may reduce approximately 50% due to the oxygenase activity of Ru-BisCo (69). Many studies have been done with light intensity and quality in terms of light supply which are the variable that impact photosynthetic activity and increase the growth kinetics of microalgae. It was reported that a strain Scenedesmus obliquus was studied with range of light/dark frequencies in an exponential condition where the photosynthetic rate is increased and also shows that in the long dark phase, the microalgal cells exploit more light energy than the light phase (122). Nevertheless, the long dark phase need not to be a necessity for attaining a greater rate of photosynthesis, nor do the cells have the ability to acclimatize in both light and dark cycles. The exposure of microalgal cells to a high spectrum of light at longer duration may ultimately damages the protein D1 in PSII. Since the damage of protein D1 causes poor capturing of photons which may lead to a total reduction of photosynthetic activity.

> There was a study on the influence of photoperiod by using Blue Green Algae cultures in BGN medium and refinery effluent for CO₂ sequestration; during this experiment, it was found that there is a gradual decrease in the biomass production in BGN medium at longer dark period (123). Refinery effluent used for growing microalgae achieved a photosynthetic quotient of 0.74, meaning that

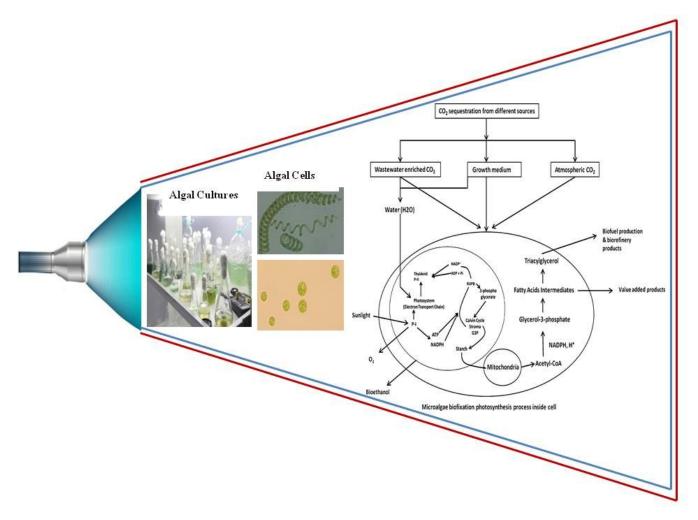


Fig. 2. Microalgal physiological pathway involved in biochemistry of CO₂ fixation Source adapted and modified (55).

1g of CO₂ is utilized to liberate 0.74 g of O₂. The above study such as Calvin cycle, PEP (Phosphoenolpyruvate) carboxconfirm that the intermittent life cycle has an impact on the ylase or synthetic pathways can improve the rate of CO_2 gas exchange pattern of the microalgal system. Sequential fixation (126, 127). Researchers have worked on the enzyme changes in the intensity of light and density of culture pro- RuBisCO via engineering to increase the catalysis rates of motes a high growth rate of microalgae. Low culture densi- carboxylation reaction which in turn enhances the activaty with high exposure of light intensity causes photo inhibi- tion state of the enzyme and reduces the oxygenation reaction of cells where as in high culture density, the spectrum tion. Another method to inhibit the oxygenation reaction is of penetrating light act as a limiting factor. However, the to enhance the regeneration phase of the Calvin cycle and growth and CO₂ mitigation of microalgal cell can be CO₂ enrichment around the enzyme. The results from the achieved more when they are exposed to red and blue above investigations revealed that the media lacking CO₂, spectrum of light (124, 125). Microalgae possess a unique temperature or light cannot support the proper functioning mechanism called as CO₂-concentrating mechanism (CCM), of RuBisCO enzyme for fixing the carbon flux via Calvin cyin which the cytoplasm of the microalgae innately accumu- cle. Chlamydomonas reinhardtii is a single-cell green alga late inorganic carbon in large quantities, compared to that that has a heterotrophic life cycle and is capable of attainon the outside. There is an impulsive decrease in the pH of ing the nuclear genome of an MRL1-deficient strain and the medium when there is a high concentration of CO₂due MRL1 maturation factor at different levels. Due to these to which the growth of many microalgal species cease. The factors, Chlamydomonas reinhardtii is considered as an idevariation in the pH is due to the increase in the CO₂ mass al candidate for RuBisCO engineering. When compared to transfer mechanism from the gas mixture to the medium. the wild type, the deficient strain showed that RuBisCo Fuel gases are said to contain very high concentrations of could maintain phototrophic growth even when it was low- CO_2 and at times more than 30% of CO_2 . Also, the O_2 pro- ered up to 15 %. These discoveries have proposed that for duced during photosynthesis inhibits the microalgal modifying the accumulation of RuBisCO, inducible MRL1 growth. Therefore it is very essential to avoid accumulation promoter can be applied based on the light intensity and of CO_2 by routinely removing it to ensure the continuous CO_2 concentration of the culture (126-129) (Table 2). growth of microalgae (41).

Life cycle assessment of CO₂ sequestration microalgae

Studies have reported that the cycles and pathways Life cycle analysis (LCA) is one of the systematic ecological

Metabolic pathway or biological process Description		Candidate CO ₂ Sensitive genes				
(Step by Step process)		Complemented network (present work)	(128)			
Transport, chloroplast	ţ	DAT1, NAR1.2				
Porphyrin and chlorophyll metabolism	ţ		GSA			
Carbon fixation	ţ	MDH5	RBCS1			
Glyoxylate metabolism	ţ		GLYK			
Glycolysis, gluconeogenesis, valine, leucine and isoleucine degradation	ţ	PGK1	PGK1			
Glycine, serine and threonine metabolism	Ļ		GCSP, THS			
Pentose phosphate pathway	Ļ	TAL1, RPE1, RPI1	RPE1			
Mitochondrial transport	Ļ	MIT28, PTB12, PTB4, PTB2				
Phenylalanine, tyrosine and tryptophan biosynthesis	Ļ	AST4				
Oxidative phosphorylation	ţ	NDA3, IPY1, IPY3				
Extracellular transport	\rightarrow		PTA3, PTA4			

Source adopted and modified (129).

List of candidate CO2 sensitive genes identified in the metabolic pathway of carbon fixation, occurs in various continuous transport process from chloroplast to extracellular components of microalgae are differentially expressed in a transcriptome dataset previously published comparing cells at high vs low CO₂ concentrations using RNA sequence.

methods used to assess the input and output records of lease of CO₂ from fossil energy out of the CO₂ uptake from This overall process includes the addition of raw materials, cultivation, harvesting, lipid extraction and various product crisis arises during different stages of the life cycle can be sis was done on various algal biomass production systems manufacturing cost and also it projects the innovative opspecific functional unit of the product (130, 131). In microalgal system the energy balance is calculated by evaluating the energy inputs needed for each LCA stage against the total expressed energy inputs related to specific product or idea of interest. Since balancing energy might influence in any phase of life cycle, it is necessary to monitor every life cycle phase to prevent any misfortunes or implications in rest of the life cycle chain.

In theory, the mechanism of capturing GHG would involve capturing CO_2 flue gas from various power stations which is subsequently utilized for promoting growth and biofuel production in microalgae (132). It was known that capturing the fuel gas does not credits the production of permissible carbon; the fact behind this is the fuel originates from algae is burnt and went back to the atmosphere, it was arise by the replacement of fossil fuels. Supplementary carbon credits may be available if the spent microalgal biomass is used for the production of electricity. Therefore, the biomass of algae would replace the coal, gas or other materials used for production of energy (133). Comprehensive life cycle calculation of energy process should be done for making biofuel. This calculation will help quantify the emission of the GHG at each stage of the process which enables the researchers to find out whether the fuel emits less CO₂ or not compared to fossil fuels (131, 132). For comparing the systems on quantitative basis different microalgae having the same functions should be selected as a unit and all the flows of energy within the system is standardized (134). To evaluate the balancing of CO_2 in a system, it is important to find and collect the details about the total re-

production systems in the entire life cycle of microalgae. microalgae cultivation (132). LCA basically covers biomass product production, usage and disposal of waste materials formation. However, the treatment of wastewater is not (130). With the help of this assessment the problem shifting covered in detail. Although, a detailed cost-effective analydetected i.e. utilization of lower energy utilization and high (132, 135). Moreover, combining wastewater treatment and algal biomass production could be a cost-effective technolportunities and environmental performance based on the ogy as well as clean to green energy process at a large scale level, thus reducing pollution.

Application of CO₂ enriched algal biomass

The existing and impending uses of microalgae are diverse and abundant. Various microalgae applications include healthcare, food, feed and industry. Even though the utilization of cyanobacterial species in the field of the food industry was done before hundred years, but new advances had been made in the 20th century (136). Several microalgae species like Spirulina platensis, chlorella sp. and diatom Odontella aurita plays a major role in the market for microalgae as food and food supplements. Additionally, Dunaliella salina is used to produce of beta carotene, Haematococcus pluvialis used for the production of astaxanthin and Aphanizomenon flos-aquae is used as a dietary supplement. The most common species of microalgae such as Spirulina, Chlorella and Scenedesmus could become a vital source of land animal feed. In chicken farming, the effect on the colour of the meat and egg yolk is due to the incorporating of 5 to 10% microalgae in the diet (137). Due to the presence of high-value compounds (HVC) in microalgae helps to produce products related to human health care. Most of the plants and animals doesn't have certain enzymes to produce long-chain polyunsaturated fatty acids (PUFAs), but various marine microalgae produces long-chain PUFA's including Arachidonic (AA), gamma-liolenic (GLA), docosahexaenoic acid (DHA) and Eicosapentaenoic (EPA). Adequate consumption of such fatty acids could have valuable effects on human health. 35–45% DHA is present in the oil from the Stramenopile, Schizochytrium sp. (permitted as a food ingredient). In comparison, 10% alpha-linolenic acid,

the precursor of omega-3 is found in most conventional oils regulatory pathways of algal cells (146). rich in omega-3 (walnut oil, canola oil). The production of these PUFAs will undoubtedly be a main dispute in the coming years. Algal pigments, such as carotenoids, betacarotene, alpha carotene, lutein, lycopene and zeaxanthin are already commercially exploited but are also the subject of intensive research. Moreover to colour salmon 95% synthetic astaxanthin used but in Japan and Canada natural source of astaxanthin extracted from Hematococcus pluvialis was used (138). Phycobiliprotiens produced by various microalgae plays a vital role in clinical and immunological therapy purposes (139). Certain microalgae species produce free radicals which results in the formation of Reactive Oxygen Species (ROS). These ROS play an imperative role in numerous chronic diseases or acute relations. Even though, production of these ROS is also employed in human health therapy. However, numerous microalgae applications have been found out in various fields, but still, genomic applications are under construction.

Genomic studies in microalgae for CO₂ sequestration

The acquirement of important genomic data on microalgae since the 1990s is due to the ascend of next-generation sequencing (NGS) technologies. An increase in NGS technologies paved the way for the availability of microbial genomes, in addition to 14 nuclear genomes, the gene repertoire of many additional species is now accessible through transcriptomics. Among the photosynthetic organisms, Chlamydomonas reinhardtii (Chlorophyta) was chosen as a model for sequencing its entire nuclear genome in 2007 (140). On the species, genetic and post-genomics tools like RNA, microarrays, antibodies and used over the past few decades. These approaches led to the identification of biological processes in response to stress, metabolic pathway and circadian clock system of microalgae species (141), photosynthetic electron transport chains (142), mechanisms of carbon concentration (143) and flagellar assembly. Also, significant research contributions have been provided found in the periplasmic space of the cell which catalyzes by proteomic studies in photosynthesis, molecular biology and evolution (144, 145). Several other sequenced micro ing formula: algae have been selected based on their phylogenetic distribution, ecological role or nature of harmfulness. These algal species are sequenced to provide extensive information on the evolution of species, aid in identifying the metabolic pathways and other processes involved in various phase of its life cycle. Data containing these genomic sequence information plays crucial role in investigating post -genomic studies, which including proteomic and transcriptomic analyses. In some cases sequencing the full genome of an algal species is excluded due to its larger genome size. Gene catalogues for such species are built by transcriptome sequencing. Transcriptomic data are utilized for exploring phylogenomic and functional post- genomic studies. Constructing these transcriptomic and genomic data took several months or years. These technological development have paved way for chief fundamental research in global ecology, functional biology and evolution of organisms. These revolution in genomic data will speed up well commercialization of algal-based bioactive compounds through the understanding of the fundamental and

Targeting microalgal genes and their function accounting CO₂ capturing

RuBisCO enzyme plays a vital role in CO₂ assimilation of algae (147). The gene rbcL is responsible for the production of this enzyme. This enzyme is found in specific location; carboxysomes in cyanobacteria and in other algae it will be located in pyrenoids. However there are few studies that shows the hypothesis of localization of RuBisCO for attainment of environmental CO_2 (148). RuBisCO is abundant in carboxysomes (149) than pyrenoids. But in microalgae Ru-BisCO is the major protein component of pyrenoids (148). At the site of RuBisCO enzyme the machinery of cells elevates the CO₂ concentration by increasing the cellular inorganic carbon (Ci) which facilitate the carboxylase activity of RubisCO. Several Ci transporters found in the plasma membranes and chloroplasts are considered as main proteins on the carbon uptake process (12). By anchoring around the pyrenoid tubules the RuBisCO-EPYC1 (Essential Pyrenoid Component 1) enhance the CO₂ fixation (150). Enzyme Fructose 1, 6- bisphosphate aldolase (FBA) involves in the Calvin cycle which leads to CO₂ sequestration. Very recently the over expression of cyanobacterial FBA by engineering the calvin cycle was found to enhance the photosynthetic capacity of C. vulgaris (151). On the other hand, the over expression of Chlamydomonas SBPase (Sedoheptulose-1,7bisphosphatase) was found to improve the photosynthetic activity in Dunaliella bardawil (152). In microalgae and Photosynthetic microbes the reaction catalyzed by the FBPase (Fructose 1, 6-bisphosphatase) can be targeted to improve the photosynthetic efficiency and accumulation of biomass. CIA5 factor, transporter of Ci and carbonic anhydrases (CA) are considered as the targets of manipulation inorder to increase the photosynthetic performance and eventually biomass yield (153-156). CA is another enzyme that is also involved in CO₂ fixation of algae. It is a zinc metalloprotein the interconversion of CO₂ and HCO₃- based on the follow-

$CO_2+H_2O\leftrightarrow H_2CO_3\leftrightarrow H^++HCO_3^-$

Carbonic anhydrases will enable the way to accumulate HCO3- within the cell. Various microalgae growing under limiting CO_2 conditions produced high amount of CA(147). In C. reinhardtii and Dunaliella salina the enzyme periplasmic CA's encoding genes are identified (157). CA1 is one of the periplasmic CA which are recognized as prominent low CO₂ inducible proteins. These proteins are identified in C. *reinhardtii*. With the help of periplasmic CA microalgae cells use external HCO₃- to do the prices of photosynthesis. For photosynthesis, the usage of external Ci is decreased due to the presence of external CA (148, 158, 159). The periplasmic CA probably increases the efficiency with which the cells can take in external Ci. This includes both the supply of CO₂ for diffusion across the plasma membrane and the supply of HCO₃⁻ for the plasma membrane's HCO₃⁻ transport system.

The CAH genes play an important role in cellular carbon uptake (152), (148) and are the targeting genes for CO_2

CO2 influences the expression of CAHs genes. The main role of these CAHs is catalyzation and inter conversion of CO₂ maintain the accumulated manganese of PSII (163, 164). and carbonic acid (H2CO₃) which increases the level of carbon uptake at the site of chloroplast for photosynthesis. A family of enzymes known as the carbonic anhydrases (CAs) have a class of \propto -CAs in them which are responsible for the diffusion of CO₂ across the plasma membrane of the cell. Proteins found to be responsible for the variations in the concentration of CO₂ are CAH1, CAH3, CAH4, CAH5 and CA-H6. It has been reported that at low CO₂ levels, a protein known as low-CO₂ inducible protein-A (LCIA) is induced and highly expressed which encodes a nitrate transporter that increases the transport of bicarbonate (HCO₃) in the stroma (CIA) (152).

Sensitive gene

Genes that are responsible for the changes involved in carbon fluxes in the system are known as sensitive genes (129, 160). Some examples of the sensitive nodes responsible for the simulations in the metabolic network are H-protein (GCSH), low-CO₂ inducible protein-A (LCIA), phosphoglucomutase (GPM2), E1 component, genes coding for glycerate kinase (GLYK), dual-function alcohol dehydrogenase/ acetaldehyde dehydrogenase (ADH1), carbonic anhydrase-5 (CAH5), NAD-dependent malate dehydrogenase (MDH3), glycine cleavage system and alpha subunit (PDC3).

Selected genes are cultivated in varying concentrations of CO₂ to study their carbon uptake properties. Experimental studies have revealed that, the carbon concentrating mechanism (CCM) related genes when cultivated in low CO₂ condition are over expressed and the cells change their metabolism which enhances the production of enzymes to uptake carbon.

Earlier comparative transcriptomics studies had revealed that when Chlamydomonas cells were subjected to a wide range of CO_2 concentrations namely very low (0.02%), low (0.05%) and high concentration (5%) (152) the results cc125 vs. acia5 when exposed from low vs. high concentration of CO₂ showed the presence of at least 345 genes that were differentially expressed. When the wild type cells were exposed from very low vs. high concentration of CO₂, they showed the presence of 696 differentially expressed genes (13, 161).

Level of CO₂ optimization in microalgae by specific gene

Microalgal cells exposed to high levels of CO₂ showed increased capability toward biomass production in previous experimental results and also the expressions of the gene transcripts for CAHs such as CAH1, CAH4, CAH5 and LCI1 were increased in low CO₂ concentrations. Microalgae have various advanced modes of CCMs to alleviate RuBisCO and arrest atmospheric CO₂ in a broad range. But one common expression in these mechanisms is the crucial act of the numerous uniquely confined intra and extracellular carbonic anhydrases (162). The specific gene CAH3 codes for the protein/metalloenzyme 'Carbonic Anhydrase' whose function is the reversible hydration of CO₂. CAH3 has two major physiological functions. One is that, it is a perfect entity for

sequestration. In microalgae the environmental level of the CA that formulates the acquired bicarbonate into CO₂ for further fixation. And the other being is the ability to

Genetically modified microalgal strains for CO₂ sequestration

In general, aquatic microorganisms often limits the availability of Ci and CO₂ using CCM's that allow them to optimize carbon acquisition. With the increasing knowledge in mutational approaches and genetic engineering, consolidated image of functional components and molecular details regarding CCM regulation is developing (165). Genetic and metabolomic engineering aids in the beneficial modification of microalgal strains. CRISPR-Cas9 (Clustered Regularly Interspaced Short Palindromic Repeats-Crispr associated protein 9), TALEN (Transcription Activator-like Effector Nucleases) and CRISPR- Cas9 (Clustered Interspaces Short Palindromic Repeats - Cripe associated protein 9) are some genome editing tools used currently for gene alterations. Synthetic biology brings in the concept of 'Biobricks' to generate artificial regulatory pathways which can alter the metabolism and inturn administer beneficial cellular characteristics. Ribosome binding sites, promotors, terminators, etc are some of the interchangeable units that serve as Biobricks.

The Calvin cycle of *Chlorella vulgaris* was genetically engineered (151) which enhanced its photosynthetic capacity by -1.2-fold (166). This strain mutated by NTG (N-methyl-N'-nitro-nitrosoguanidine) mutagenesis thrived efficiently in 10% CO₂ and is approached for CO₂ sequestration. Some cyanobacteria members were genetically modified to produce and secrete CAs into the medium which was efficiently transformed the CO₂ into HCO₃ and which in turn was taken up by the microalgae and further fixed into biomass through the process of photosynthesis.

Genomic Resources of microalgae and CRISPR/Cas9 tools

Until 2008, only three microalgal species were sequenced which includes Chlamydomonas reinhardtii, Thalassiosira showed that the mutant strain cc2702 and wild type strain pseudonana and Phaeodactylum tricornutum (167). But over the past years, the great revolution of next-generation sequencing technology has created a prompt rise in the availability of complete genomes of various algal species and drafts. Currently about 60 algal accessions are sequenced completely and it's completed genome profiles are available in phytozone and "The Greenhouse".

> There are about three databases available for algal genomics. The first database is pico-PLAZA (https:// bioinformatics.psb.ugent.be/pico-plaza/), which consists of genomic informations and instinctual tools of functional genomics of the 16 algal species (168). The second database is Algae Path (http://algaepath.itps.ncku.edu.tw); it gives information about the expression of gene predicting metabolic pathway of Chlamydomonas reinhardtii and Neodesmus sp. UTE X 2219-4 (169). The third one is ALCOdb (http://alcodb.jp), which provides details about coexpression gene data of two algal species (Cyanidioachyzon merolae and Chlamydomonas reinhardtii) (170). Additionally, the completed genome sequence draft are also available in phytozome (https://phytozome.jgi.doe.gov) and JGI Ge

nome portal (https://genome.jgi.doe.gov) (Supplementary Table 1).

Challenges in genomic studies

Bringing out new technologies and products into the mar- Progress in microalgal genome analysis ket for commercialization faces many restrictions. Genetic engineering is one of the most used technology to augment CO₂ assimilation in microalgae. As genetically engineered organisms such as transgenic or recombinant algae are required to undergo regulatory compliances in the form of laws and policies in many parts of the world, this technology faces several challenges. Due to the challenges faced, even though extended research is carried out to genetically modify and produce an improved and enhanced performing microalgae, their commercialization is restricted. Therefore, advances in the technology can only occur when the investigation/research and policy complement each other.

- Research centres must be well equipped and possess expert human resources who are well trained with hands-on experience in genomic analysis/ sequencing and complete the tasks efficiently.
- Defining the full genome sequences and Complementary DNA Sequences (cDNA) of an organism is the foremost step of the scientific community to get whole sequence information about particular organisms, which can be established further by training the agencies and other communities through workshops or tutorials.
- Lack of prior knowledge in the field of microalgal genomics, proteomics and its biochemicalphysiological process increases problems during genomic studies (183).
- The path toward algal genomics has yet not touched its horizons, required for pan-genomic studies.
- Phylogenetic and BLAST analysis are the significant tools used effectively for genomic studies, but they only explain the presence, absence and variability among known genetic loci. Multiple species are tion (190). extensively sequenced to get an understanding of Future prospects the basics of algal species (184).
- studied to improve of photosynthetic CO₂ utilization in the field of genetic engineering. Even though, it resulted in limited success (185, 186).
- Evaluating and correlating useful genomic inputs is very necessary to scrutinize for predicting the function of proteins. There should be awareness among the researchers about the quality of published genome assemblies because most of the genome assemblies are incomplete and inaccurate (187).
- There has been a rapid increase in the completed genomes and the available number of draft of algal species due to the revolution in the past decade on the "next-generation sequencing" technologies.

Several attempts made to define the genomic sequence of microalgal species of disparate group (160, 167).

Substantial advances in the evolution of new genetic manipulation tools helps to manipulate the central carbon metabolism in the microalgae. Most of such advances could be used in defining the industrially relevant organisms (188). Auspicious developmental advances in the metabolic engineering tools not only increases the endogenous carbon storage compounds production like starch and TAGs, it also helps in the direct production of hydrocarbons which can be used in fuel production. The usage of these metabolic engineering tools in microalgae will enhance the production of sources of renewable fuel (188).

C. reinhardtii and P. tricornutum are considered as reference organisms because it is used to characterize the specific adaptations of algae at the molecular and genetic level (187). Microalgal species such as Fragilariopsis cylindrus, Pseudo-nitzschia, Thalassiosira rotula, Botryococcus braunii, Chlorella vulgaris, Dunaliella salina, Micromonas pusilla, Galdieria sulphuraria, Porphyra purpurea, Volvox carteri and Aureococcus anophageferrens are some of the genome sequencing projects in progress (189). Only three microalgal species Phaeodactylum tricornutum, Chlamydomonas reinhardtii and Thalassiosira pseudonana were sequenced till 2008 (167). Due to the technological revolution in genomics, copious number of completed genomes of various algal species were drafted and stored. Review are on the efforts of sequencing the genome of miscellaneous group of microalgal species (160, 167). Cas9 is one of the developing and systematic nuclease guided genome editing tool which helps in enhancement of algal genomics. Nowadays CRISPR/Cas9 is extensively cast-off by researchers because it is easily adaptable and cost-effective (182). The CRISPR/Cas9-based genome-editing method was done in oleaginous microalga Nannochloropsis oceanica, using the enzyme nitrate reductase (NR; g7988) to increase the scalable amount of oil production and carbon sequestra-

Expanding knowledge on the diversity of algae that can be In RuBisCO, the rate of catalytic activity has been used as a competent and economical alternative for wastewater treatment, CO₂ fixation, lipid synthesis towards economic biofuel production etc., is essential (191). This can be achieved by combining genomic studies along with technologies such as CRISPR-Cas systems and genome engineering, which will result in exploration and knowledge gaining of a broader range of organisms (187). Pivotal need should be taken as an expectant alternative to current mitigation strategies is an integrated CO₂ bio-fixation, biofuel production and as a value addition for algal biomass.

Conclusion

This review covers the general method of CO₂ sequestration and found that the CO₂ sequestration using Green Genome Technology (GGT) is a very acute method than others be-

cause specific target gene(s) present in microalgae shows the reduction of CO₂ which could enhance the efficacy of the method in a short period. Moreover after CO₂ sequestra- 6. tion, microalgal biomass production can be used to extract various high-value added products and socio-econometric. This will help to improve the economic feasibility of algal by -products from CO₂ enriched biomass and protect climate change. Consciousness about applications of algal biomass could make a common man into an entrepreneur. Currently, microalgae Genome studies have become very popular for CO₂ sequestration, and attempts are under way to Lab science to Land science technology because microalgae genome resources is a quantum jump to development in the efficacy of CO₂ sequestration. This research review article could bring their scope and hope to make the CO_2 se- 9. questration in a combact green genome technology which could be more effective on time and every time emphazising the 4Es that is Education, Environment, Economy and Empowerment of our world legacies.

Acknowledgements

The authors are grateful to the Principal and Head, Department of Botany, Madras Christian College (Autonomous) for their support and providing the necessary infrastructure.

Authors contributions

All authors have read and approved the final manuscript. As a corresponding author, I certify that the submission is an original work and is not under review at any other publication, nor has it previously been submitted to any other journal.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflicts of interests to declare.

Ethical issues: None.

Supplementary data

Table 1. CRISPR/Cas9 tools.

References

- Saklani N, Khurana A. Global Warming: Effect on Living Organisms, Causes and its Solutions. Int J Eng Manag Res. 2019;09 18. (05):24–26.
- 2. Sivaramanan S. Global warming and Climate change. Int J Glob Sci Res. 2019;6(2).
- 3. Umair Shahzad R. Global Warming Causes, Effects and Solution'S Trials. JES J Eng Sci. 2012;40(4):1233–54.
- Liu Y, Zhou ZUW, Hong-Cai. Recent advances in carbon dioxide capture with metal-organic frameworks. Greenh Gases Sci Technol. 2012;2(5):352–68.
- Habib MA, Badr HM, Ahmed SF, Ben Mansour R, Mezghani K, Imashuku S *et al.* A review of recent developments in carbon capture utilizing oxy fuel combustion in conventional and ion transport membrane systems. Int J energy Res [Internet].

2011;35(9):741–64. Available from: https:// onlinelibrary.wiley.com/doi/epdf/10.1002/er.1798

- Meis NNAH, Bitter JH, Jong KP de. Support and size effects of activated hydrotalcites for precombustion CO₂ capture. Ind Eng Chem Res [Internet]. 2010;49:1229-35. Available from: https:// pubs.acs.org/doi/10.1021/ie901114d
- Rubin ES, Mantripragada H, Marks A, Versteeg P, Kitchin J. The outlook for improved carbon capture technology. Prog Energy Combust Sci [Internet]. 2012;38(5):630–71. Available from: http:// dx.doi.org/10.1016/j.pecs.2012.03.003
- Thiruvenkatachari R, Su S, An H, Yu X. Post combustion CO₂ capture by carbon fibre monolithic adsorbents. Prog Energy Combust Sci [Internet]. 2009;35(5):438–55. Available from: https:// www.cheric.org/research/tech/periodicals/view.php? seq=1334808
- Liang Y, Harrison DP, Gupta RP, Green DA, McMichael WJ. Carbon Dioxide Capture Using Dry Sodium-Based Sorbents. Energy Fuels [Internet]. 2004;18(2):569–75. Available from: https:// pubs.acs.org/doi/10.1021/ef030158f
- Abanades JC, Alonso M, Rodríguez N. Biomass combustion with in situ CO₂ capture with CaO. I. process description and economics. Ind Eng Chem Res. 2011;50(11):6972–81.
- Qi G, Wang Y, Estevez L, Duan X, Anako N, Park A-HA et al. High efficiency nanocomposite sorbents for CO₂ capture based on amine-functionalized mesoporous capsules. Energy Environ Sci, [Internet]. 2011;4:444–52. Available from: https://pubs.rsc.org/ en/content/articlelanding/2011/ee/c0ee00213e#!divAbstract
- Wang Y, Duanmu D, Spalding MH. Carbon dioxide concentrating mechanism in *Chlamydomonas reinhardtii*: Inorganic carbon transport and CO₂ recapture. Photosynth Res. 2011;109(1–3):115 –22.
- 13. Jones CW. CO_2 capture from dilute gases as a component of modern global carbon management. Annu Rev Chem Biomol Eng. 2011;2:31–52.
- 14. Kenarsari SD, Yang D, Jiang G, Zhang S, Wang J, Russell AG *et al.* Review of recent advances in carbon dioxide separation and capture. RSC Adv. 2013;3(45):22739–73.
- Kobayashi K, Uno Y. Influence of alkali on carbonation of concrete. Part 1 – Preliminary tests with mortar specimens. Cem Concr Res [Internet]. 1989;19(5):821–26. Available from: https:// www.sciencedirect.com/science/article/abs/ pii/0008884689900537
- Jang JG, Kim GM, Kim HJ, Lee HK. Review on recent advances in CO₂ utilization and sequestration technologies in cement-based materials. Constr Build Mater [Internet]. 2016;127(30):762–73. Available from: https://www.sciencedirect.com/science/article/ abs/pii/S0950061816316300
- Jang J, HK, KK, KL. Resistance of coal bottom ash mortar against the coupled deterioration of carbonation and chloride penetration. Mater Des [Internet]. 2016;93:160–67. Available from: http:// dx.doi.org/10.1016/j.matdes.2015.12.074
- Fernández Bertos M, Simons SJR, Hills CD, Carey PJ. A review of accelerated carbonation technology in the treatment of cementbased materials and sequestration of CO2. J Hazard Mater. 2004;112(3):193–205.
- Lange LC, Hills CD, Poole AB. The effect of accelerated carbonation on the properties of cement-solidified waste forms. Waste Manag [Internet]. 1996;16(8):757–63. Available from: https://www.sciencedirect.com/science/article/abs/pii/S0956053X97000226
- Freund P, Ormerod WG. Progress towards storage of carbon dioxide. Energy Convers Manag. 1997;38:198–205.
- 21. Lal R. Carbon sequestration. Philos Trans R Soc B Biol Sci. 2008;363(1492):815–30.

14 KUMAR ET AL

- 22. Kerr RA. Bush Backs Spending for a "Global Problem." Science. 41. Pires JCM, Alvim-Ferraz MCM, Martins FG, Simões M. Carbon diox-2001 Jun;1982-3. Available from: http:// ubc.summon.serialssolutions.com/2.0.0/link/0/
- 23. O'Connor WK, Dahlin DC, Nilsen DN, Rush GE, Walters RP, Turner PC. Carbon dioxide sequestration by direct mineral carbonation: Results from recent studies and current status. In: 1st Annual 42. DOE Carbon Sequestration Conference, Washington, DC. Washington, D.C; 2001. p. 11.
- 24. Tsang CF, Benson SM, Kogelski B, Smith RE. Scientific considerations related to regulation development for CO₂ sequestration in brine formations. Environ Geol. 2002;42:275-81.
- 25. Klara SM, Srivastava RD, McIlvried HG. Integrated collaborative technology development program for CO2 sequestration in geologic formations - United States Department of Energy R&D. En- 44. ergy Convers Manag. 2003;44(17):2699-712.
- 26. Baines SJ, Worden RH. Geological storage of carbon dioxide. Geol Soc Spec Publ. 2004;233(June):1-6.
- 27. Kintisch E. Report Backs More Projects to Sequester CO₂ From Coal. Science [Internet]. 2007;315(5818):1481. Available from: https://science.sciencemag.org/content/315/5818/1481.1/tab-eletters
- 28. Schrag DP. Preparing to capture carbon. Science (80-) [Internet]. 2007;9(315(5813)):812-23. Available from: https:// pubmed.ncbi.nlm.nih.gov/17289991/
- 29. Klusman RW. Evaluation of leakage potential CO_2 EOR/ sequestration project. Energy Convers Manag. 2003;33:1921-40.
- 30. Lackner KS. A guide to CO₂ sequestration. Science (80-). 2003;300 (5626):1677-78.
- 31. Park A-HA, Fan L-S. CO₂ mineral sequestration: Physically activat- 47. ed dissolution of serpentine and pH swing process. Chem Eng Sci. 2004;59(22-23):5241-47.
- 32. Rivkin RB, Legendre L. Biogenic carbon cycling in the upper ocean: Effects of microbial respiration. Science (80). 2001;291 48. (5512):2398-400.
- 33. Falkowski P, Scholes RJ, Boyle E, Canadell J, Canfield D, Elser J, et al. The global carbon cycle: A test of our knowledge of earth as a system. Science (80-). 2000;290(5490):291-96.
- 34. Raven JA, Falkowski PG. Oceanic sinks for atmospheric CO2. Plant, Cell Environ. 1999;22(6):741-55.
- 35. Falkowski PG. Letters to nature. Nature. 1997;387:243-44.
- 36. Martin JH, Fitzwater SE. Iron deficiency limits phytoplankton growth in the north-east Pacific subarctic. Nature [Internet]. 50. 1988;331(6154):341-43. Available from: https:// app.dimensions.ai/details/publication/pub.1002094778
- 37. Boyd P, Law C, Wong C, Al. E. The decline and fate of an ironinduced subarctic phytoplankton bloom. Nature [Internet]. 2004;428:549–53. Available from: https://www.nature.com/ 51. articles/nature02437#citeas
- 38. Lal R, Follett RF, Kimble JM. Achieving soil carbon sequestration in the united states: A challenge to the policy makers. Soil Sci [Internet]. 2003;168(12):827–45. Available from: https:// 52. journals.lww.com/soilsci/Abstract/2003/12000/ achieving_soil_carbon_sequestration_in_the_united.1.aspx
- 39. Figueroaa DJ, TimothyFouta, SeanPlasynskia, Howard- 53. McIlvriedb, Rameshwar. Advances in CO2 capture technology-The U.S. Department of Energy's Carbon Sequestration Program. Greenh Gas Control [Internet]. 2008;2(1):9–20. Available from: https://www.sciencedirect.com/science/article/abs/pii/ S1750583607000941
- 40. Pires JCM, Martins FG, Alvim-Ferraz MCM, Simões M. Recent developments on carbon capture and storage: An overview. Chem Eng Res Des [Internet]. 2011;89(9):1446-60. Available from: http://dx.doi.org/10.1016/j.cherd.2011.01.028

- ide capture from flue gases using microalgae: Engineering aspects and biorefinery concept. Renew Sustain Energy Rev [Internet]. 2012;16(5):3043-53. Available from: http:// dx.doi.org/10.1016/j.rser.2012.02.055
- Svenssona R, Odenbergera M, Johnssona F, Strömberg L. Transportation systems for CO2--application to carbon capture and storage. Energy Convers Manag [Internet]. 2004;45(15-16):2343-53. Available from: https://www.sciencedirect.com/science/ article/abs/pii/S0196890403003662
- 43 McCoy ST, Rubin ES. An engineering-economic model of pipeline transport of CO2 with application to carbon capture and storage. Int J Greenh Gas Control. 2008;2(2):219-29.
- Lam MK, Lee KT, Mohamed AR. Current status and challenges on microalgae-based carbon capture. Int J Greenh Gas Control 2012;10:456-69. from: https:// [Internet]. Available www.sciencedirect.com/science/article/abs/pii/ S1750583612001673
- De Silva GPD, Ranjith PG, Perera MSA. Geochemical aspects of 45. CO2 sequestration in deep saline aquifers: A review. Fuel [Internet]. 2015;155:128-43. Available from: http:// dx.doi.org/10.1016/j.fuel.2015.03.045
- 46. Nouha K, John RP, Yan S, Tyagi RD. Carbon capture and sequestration: Biological technologies. In: Surampalli RY, Zhang TC, Tyagi RD, Ravi Naidu BR, Gurjar CS, Ojha P et al., editors. Carbon Capture and Storage - Physical, Chemical and Biological Methods [Internet]. Reston, VA 20191-4400: American Society of Civil Engineers; 2015. Available from: https://app.knovel.com/web/toc.v/ cid:kpCCSPCBM8/viewerType:toc/
- Farrelly DJ, Everard CD, Fagan CC, McDonnell KP. Carbon sequestration and the role of biological carbon mitigation: A review. Renew Sustain Energy Rev [Internet]. 2013;21:712-27. Available from: http://dx.doi.org/10.1016/j.rser.2012.12.038
- Cheah WY, Ling TC, Juan JC, Lee DJ, Chang JS, Show PL. Biorefineries of carbon dioxide: From carbon capture and storage (CCS) to bioenergies production. Bioresour Technol [Internet]. 2016;215:346-56. Available from: http://dx.doi.org/10.1016/ j.biortech.2016.04.019
- Williamson P, Wallace DWR, Law CS, Boyd PW, Collos Y, Croot P, 49. et al. Ocean fertilization for geoengineering: A review of effectiveness, environmental impacts and emerging governance. Process Saf Environ Prot [Internet]. 2012;90(6):475-88. Available from: http://dx.doi.org/10.1016/j.psep.2012.10.007
- Yadav G, Sen R. Microalgal green refinery concept for biosequestration of carbon-dioxide vis-à-vis wastewater remediation and bioenergy production: Recent technological advances in climate research. J CO2 Util [Internet]. 2017;17:188-206. Available from: http://dx.doi.org/10.1016/j.jcou.2016.12.006
- Zhou W, Wang J, Chen P, Ji C, Kang Q, Lu B et al. Bio-mitigation of carbon dioxide using microalgal systems: Advances and perspectives. Renew Sustain Energy Rev [Internet]. 2017;76(March):1163-75. Available from: http://dx.doi.org/10.1016/j.rser.2017.03.065
- Singh J, Dhar DW. Overview of carbon capture technology: Microalgal biorefinery concept and state-of-the-art. Front Mar Sci. 2019:6(FEB):1-9.
- Slade R, Bauen A. Micro-algae cultivation for biofuels: Cost, energy balance, environmental impacts and future prospects. Biomass and Bioenergy [Internet]. 2013;53(0):29-38. Available from: http://dx.doi.org/10.1016/j.biombioe.2012.12.019
- Valdovinos-García EM, Barajas-Fernández J, de los Ángeles Olán-54. Acosta M, Petriz-Prieto MA, Guzmán-López A, Bravo-Sánchez MG. Techno-Economic Study of CO2 Capture of a Thermoelectric Plant Using Microalgae (Chlorella vulgaris) for Production of Feedstock for Bioenergy. Energies. 2020;13(2):1-19.
- 55. Bhola V, Swalaha F, Ranjith Kumar R, Singh M, Bux F. Overview of

the potential of microalgae for CO2 sequestration. Int J Environ Sci Technol. 2014;11(7).

- 56. Gaikwad RW, Gudadhe MD, Bhagat SL. Carbon dioxide capture, tolerance and sequestration using microalgae- A review. Int J Pharm Chem Biol Sci [Internet]. 2016;6(3):345-49. Available from: http://www.ijpcbs.com/files/13-07-2016/13.pdf
- 57. Fernández I, Acién FG, Fernández JM, Guzmán JL, Magán JJ, 70. Berenguel M. Bioresource Technology Dynamic model of microalgal production in tubular photobioreactors. 2012;126:172–81.
- 58. Costache TA, Gabriel Acien Fernandez F, Morales MM, Fernández-71. Sevilla JM, Stamatin I, Molina E. Comprehensive model of microalgae photosynthesis rate as a function of culture conditions in photobioreactors. Appl Microbiol Biotechnol. 2013;97(17):7627-37.
- 59. Seo SH, Ha JS, Yoo C, Srivastava A, Ahn CY, Cho DH et al. Light intensity as major factor to maximize biomass and lipid productivity of Ettlia sp. in CO2-controlled photoautotrophic chemostat [Internet]. Vol. 244, Bioresource Technology. Elsevier Ltd; 2017. 621-28. http://dx.doi.org/10.1016/ Available from: j.biortech.2017.08.020
- Vonshak A, Giuseppe T. Environmental Stress Physiology. In: 60. Richmond A editor. Handbook of Microalgal Culture: Biotechnology and Applied Phycology. Iowa 50014-8300, USA: Blackwell 74. Publishing Ltd; 2004. p. 57-82.
- 61. Falkowski P., Chen. Y.B. Photoacclimation of Light Harvesting Systems in Eukaryotic Algae. In: B.R. G, W.W. P editors. Light-Harvesting Antennas in Photosynthesis Advances in Photosynthesis and Respiration [Internet]. Springer, Dordrecht; 2003. p. 423-47. Available from: https://link.springer.com/ chapter/10.1007/978-94-017-2087-8_15#citeas
- 62. Nikolaou A, Hartmann P, Sciandra A, Chachuat B, Bernard O. Dynamic coupling of photoacclimation and photoinhibition in a model of microalgae growth. J Theor Biol [Internet]. 2016;390:61 -72. Available from: http://dx.doi.org/10.1016/j.jtbi.2015.11.004
- 63. Kommareddy A, Anderson G. Study of Light as a parameter in the growth of algae in a Photo-Bioreactor (PBR). In: The Society for engineering in agricultural, food and biological systems [Internet]. Las Vegas, NV; 2003. p. 2–23. Available from: https:// www.researchgate.net/

publica-

. tion/280720969_Study_of_Light_as_a_parameter_in_the_growt h_of_algae_in_a_Photo-Bioreactor_PBR#:~:text=A photo-bio Reactor is, parameter in a photosynthetic process. & text=Light 79. with wavelengths between 600, the most effici

- 64. Luveshan R, Rawat I, Bux F. Light enhancement strategies improve microalgal biomass productivity. Renew Sustain Energy 80. Rev [Internet]. 2017;80:765-73. Available from: https:// www.sciencedirect.com/science/article/abs/pii/ S1364032117308389
- 65. Morales M, Sánchez L, Revah S. The impact of environmental factors on carbon dioxide fixation by microalgae. FEMS Microbiol Lett. 2018;365(3):1-11.
- Molina Grima E, Belarbi EH, Acién Fernández FG, Robles Medina 66. A, Chisti Y. Recovery of microalgal biomass and metabolites: Process options and economics. Biotechnol Adv. 2003;20(7-8):491-515.
- 67. Béchet Q, Shilton A, Guieysse B. Modeling the effects of light and temperature on algae growth: State of the art and critical assessment for productivity prediction during outdoor cultivation. Biotechnol Adv [Internet]. 2013;31(8):1648-63. Available from: 83. Cheng L, Zhang L, Chen H, Gao C. Carbon dioxide removal from http://dx.doi.org/10.1016/j.biotechadv.2013.08.014
- 68. Ördög V, Stirk WA, Bálint P, Aremu AO, Okem A, Lovász C et al. Effect of temperature and nitrogen concentration on lipid productivity and fatty acid composition in three Chlorella strains. Algal Res [Internet]. 2016;16:141–49. Available from: http://

dx.doi.org/10.1016/j.algal.2016.03.001

- 69. Kumar A, Ergas S, Yuan X, Sahu A, Zhang Q, Dewulf J et al. Enhanced CO2 fixation and biofuel production via microalgae: Recent developments and future directions. Trends Biotechnol [Internet]. 2010;28(7):371-80. Available from: http:// dx.doi.org/10.1016/j.tibtech.2010.04.004
- Ras M, Steyer JP, Bernard O. Temperature effect on microalgae: A crucial factor for outdoor production. Rev Environ Sci Biotechnol. 2013;12(2):153-64.
- Zhao B, Su Y. Process effect of microalgal-carbon dioxide fixation and biomass production: A review. Renew Sustain Energy Rev. 2014:31:121-32.
- 72. Briassoulis D, Panagakis P, Chionidis M, Tzenos D, Lalos A, Tsinos C et al. An experimental helical-tubular photobioreactor for continuous production of Nannochloropsis sp. Bioresour Technol [Internet]. 2010;101(17):6768-77. Available from: http:// dx.doi.org/10.1016/j.biortech.2010.03.103
- 73. Morais MG De, Alberto J, Costa V. Carbon dioxide fixation by Chlorella kessleri, C. vulgaris, Scenedesmus photobioreactors Carbon dioxide fixation by Chlorella kessleri, C. vulgaris, Scenedesmus obliguus and Spirulina sp. cultivated in flasks and vertical tubular photobioreactors. 2007;(October 2014).
- Zhang H, Zeng R, Chen D, Liu J. A pivotal role of vacuolar H+-ATPase in regulation of lipid production in Phaeodactylum tricornutum. Sci Rep [Internet]. 2016;6(April):1-17. Available from: http://dx.doi.org/10.1038/srep31319
- 75. Moss B. The influence of environmental factors on the distribution of freshwater algae: An experimental study: I. Introduction and the influence of calcium concentration. J Ecol [Internet]. 1972;60(3):917–32. Available from: https://www.jstor.org/ stable/2258575?seq=1
- 76. Azov Y. Effect of pH on inorganic carbon uptake in algal cultures. Appl Env Microbiol [Internet]. 1982;43(6):1300-06. Available from: https://pubmed.ncbi.nlm.nih.gov/16346029/
- Olaizola M. Commercial development of microalgal biotechnology: From the test tube to the marketplace. Biomol Eng. 2003;20(4 -6):459-66.
- 78. Tiwari A, Pandey JP, Pathak N. Standardization of pH and Light Intensity for the Biomass Production of Spirulina platensis. J Algal Biomass Utln [Internet]. 2010;2010(2):93-102. Available from: https://www.researchgate.net/publication/299597097
- Ying K, Gilmour DJ, Zimmerman WB. Effects of CO₂ and pH on growth of the microalga Dunaliella salina. J Microb Biochem Technol. 2014;6(3):167–73.
- Duarte-Santos T, Mendoza-Martín JL, Acién Fernández FG, Molina E, Vieira-Costa JA, Heaven S. Optimization of carbon dioxide supply in raceway reactors: Influence of carbon dioxide molar fraction and gas flow rate. Bioresour Technol [Internet]. 2016;212:72-81. Available from: http://dx.doi.org/10.1016/ j.biortech.2016.04.023
- 81. Bilanovic D, Holland M, Starosvetsky J, Armon R. Co-cultivation of microalgae and nitrifiers for higher biomass production and better carbon capture. Bioresour Technol [Internet]. 2016;220:282-88. Available from: http://dx.doi.org/10.1016/ j.biortech.2016.08.083
- Solovchenko A, Khozin-Goldberg I. High-CO2 tolerance in micro-82. algae: Possible mechanisms and implications for biotechnology and bioremediation. Biotechnol Lett. 2013;35(11):1745-52.
- air by microalgae cultured in a membrane-photobioreactor. Sep Purif Technol. 2006;50(3):324-29.
- Yun Y, Lee SB, Park JM, Lee C, Yang J. Carbon Dioxide Fixation by 84. Algal Cultivation Using Wastewater Nutrients. J Chem Technol Biotechnol [Internet]. 1997;69(4):451-55. Available from: https://

15

onlinelibrary.wiley.com/doi/abs/10.1002/%28SICI%291097-4660%28199708%2969%3A4%3C451%3A%3AAID-JCTB733% 3E3.0.CO%3B2-M

- 85. Chiu SY, Kao CY, Chen CH, Kuan TC, Ong SC, Lin CS. Reduction of CO2 by a high-density culture of Chlorella sp. in a semicontinuous photobioreactor. Bioresour Technol. 2008;99(9):3389-96.
- 86. Singh SP, Singh P. Effect of CO₂ concentration on algal growth: A review. Renew Sustain Energy Rev. 2014;38:172-79.
- 87. Singh UB, Ahluwalia AS. Microalgae: a promising tool for carbon sequestration. Mitig Adapt Strateg Glob Chang [Internet]. 2013;18:73–95. Available from: https://link.springer.com/ article/10.1007/s11027-012-9393-3#citeas
- 88. Sung KD, Lee JS, Shin CS, Park SC. Isolation of a new highly CO2 tolerant fresh water microalga Chlorella sp. KR-1. Renew Energy. 1999;16(1-4):1019-22.
- 89. Sivakumar M, Ranjith Kumar R, Shashirekha V, Seshadri S. Influence of carbon-dioxide on the growth of Spirulina sp. (MCRC-A0003) isolated from Muttukadu backwaters, South India. World J Microbiol Biotechnol. 2014;30(10).
- 90. Hanagata N, Takeuchi T, Fukuju Y, Barnes DJ, Karube I. Tolerance of microalgae to high CO2 and high temperature. Phytochemistry. 1992;31(10):3345-48.
- 91. Kim W, Park JM, Gim GH, Jeong SH, Kang CM, Kim DJ et al. Optimization of culture conditions and comparison of biomass productivity of three green algae. Bioprocess Biosyst Eng. 2012;35(1-2):19-27.
- 92. Cabello J, Toledo-Cervantes A, Sánchez L, Revah S, Morales M. Effect of the temperature, pH and irradiance on the photosynthetic activity by Scenedesmus obtusiusculus under nitrogen replete and deplete conditions. Bioresour Technol [Internet]. 2015;181:128-35. Available from: http://dx.doi.org/10.1016/ j.biortech.2015.01.034
- 93. Toledo-Cervantes A, Morales M, Novelo E, Revah S. Carbon dioxide fixation and lipid storage by Scenedesmus obtusiusculus. Bioresour Technol [Internet]. 2013;130:652–58. Available from: http://dx.doi.org/10.1016/j.biortech.2012.12.081
- 94. Fan L, Vonshak A, Boussiba S. Effect of temperature and irradiance on growth of Haematococcus pluvialis (Chlorophyceae). J Phycol. 1994;30:829-33.
- 95. Sarada R, Tripathi U, Ravishankar GA. Influence of stress on astaxanthin production in *Haematococcus pluvialis* grown under different culture conditions. Process Biochem. 2002;37(6):623-27.
- 96. Huntley ME, Redalje DG. CO2 mitigation and renewable oil from photosynthetic microbes: A new appraisal. Vol. 12, Mitigation and Adaptation Strategies for Global Change. 2007; 573-608.
- 97. Bitaubé Pérez E, Caro Pina I, Pérez Rodríguez L. Kinetic model for growth of Phaeodactylum tricornutum in intensive culture photobioreactor. Biochem Eng J. 2008;40(3):520-25.
- 98. Kebede E, Ahlgren G. Optimum growth conditions and light utilization efficiency of Spirulina platensis (= Arthrospira fusiformis) (Cyanophyta) from Lake Chitu, Ethiopia. Hydrobiologia. 1996;332 113. Tapie P, Bernard A. Microalgae production: Technical and eco-(2):99-109.
- 99. De Oliveira MACL, Monteiro MPC, Robbs PG, Leite SGF. Growth and chemical composition of Spirulina maxima and Spirulina platensis biomass at different temperatures. Aquac Int. 1999;7 (4):261-75.
- 100. Colla LM, Oliveira Reinehr C, Reichert C, Costa JAV. Production of biomass and nutraceutical compounds by Spirulina platensis under different temperature and nitrogen regimes. Bioresour Technol. 2007;98(7):1489–93.
- 101. Kumar A, Yuan X, Sahu AK, Ergas SJ, Van Langenhove H, Dewulf J. A hollow fiber membrane photo-bioreactor for CO₂ sequestration 116. Tsuzuki M, Gantar M, Aizawa K, Miyachi S. Ultrastructure of Dufrom combustion gas coupled with wastewater treatment: A

process engineering approach. J Chem Technol Biotechnol. 2010;85(3):387-94.

- 102. Nakano Y, Miyatake K, Okuno H, Hamazaki K, Takenaka S, Honami N et al. Growth of photosynthetic algae euglena in high CO₂ conditions and its photosynthetic characteristics. Acta Hortic [Internet]. 1996;49-54. Available from: https://www.ishs.org/ishs -article/440_9#:~:text=Acta Horticulturae 440- Growth of photosynthetic algae euglena in high conditions and its photosynthetic characteristics & text=Photoautotrophic growth of Euglena gracilis, than 45 %25 CO2 concentration
- 103. Kitaya Y, Azuma H, Kiyota M. Effects of temperature, CO₂ /O₂ concentrations and light intensity on cellular multiplication of microalgae, Euglena gracilis. Adv Sp Res. 2005;35(9 SPEC. ISS.):1584 -88
- 104. Benemann JR. Utilization of carbon dioxide from fossil fuelburning power plants with biological systems. Energy Convers Manag [Internet]. 1993;34(9-11):999-1004. Available from: https://www.sciencedirect.com/science/article/abs/ pii/019689049390047E#:~:text=Biological processes for CO2, gases in a practical process.
- 105. Nobutaka H, Toshifumi Takeuchia, Fukuju Y, J.Barnesa D, ISao Karubea. Tolerance of microalgae to high CO₂ and high temperature. Phytochemistry [Internet]. 1992;10(31):3345-48. Available https://www.sciencedirect.com/science/article/abs/ from: pii/003194229283682O#:~:text=Scenedesmus was better able to,able to tolerate elevated temperatures
- 106. Graham LKE, Wilcox LW. The origin of alternation of generations in land plants: A focus on matrotrophy and hexose transport. Philos Trans R Soc B Biol Sci. 2000;355(1398):757-67.
- 107. Anguselvi V, Masto RE, Mukherjee A, Singh PK. CO₂ Capture for Industries by Algae. In: Wong YK editor. Algae [Internet]. London: IntechOpen; 2019. Available from: https://www.intechopen.com/ books/algae/co-sub-2-sub-capture-for-industries-by-algae
- 108. Brown LM. Uptake of carbon dioxide from flue gas by microalgae. Energy Convers Manag. 1996;37(6-8):1363-67.
- 109. Sobczuk TM, Camacho FG, Rubio FC, Fernández FGA, Grima EM. Carbon dioxide uptake efficiency by outdoor microalgal cultures in tubular airlift photobioreactors. Biotechnol Bioeng [Internet]. 2000;67(4):465-75. Available from: https:// onlinelibrary.wiley.com/doi/10.1002/(SICI)1097-0290(20000220) 67:4%3C465::AID-BIT10%3E3.0.CO;2-9
- 110. Lee YK, Hing HK. Supplying CO2 to photosynthetic algal cultures by diffusion through gas-permeable membranes. Appl Microbiol Biotechnol. 1989;31(3):298-301.
- 111. Benemann JR, Tillett DM, Weissman JC. Microalgae biotechnology. Trends Biotechnol. 1987;5(2):47-53.
- 112. Oswald JA. Large-Scale Algal Culture Systems (Engineering Aspects). In: Borowitzka LJ, Borowitzka MA, editors. Microalgal biotechnology [Internet]. Cambridge: Cambridge University Press: 1988. p. 357-95. Available from: https:// books.google.co.in/books/about/ Micro_algal_Biotechnology.html?id=FpprQgAACAAJ
- nomic evaluations. Biotechnol Bioeng. 1988;32(7):873-85.
- 114. Papazi A, Makridis P, Divanach P, Kotzabasis K. Bioenergetic changes in the microalgal photosynthetic apparatus by extremely high CO₂ concentrations induce an intense biomass production. Physiol Plant [Internet]. 2008;132(3):338–49. Available from: https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1399-3054.2007.01015.x
- 115. Hessen DO, Anderson TR. Excess carbon in aquatic organisms and ecosystems: Physiological, ecological and evolutionary implications. Limnol Oceanogr. 2008;53(4):1685-96.
- naliella tertiolecta Cells Grown under Low and High CO₂ Concen-

from: https://academic.oup.com/pcp/articleabstract/27/4/737/1850783?redirectedFrom=fulltext

- 117. Calvin M. Forty years of photosynthesis and related activities. Photosynth Res [Internet]. 1989;21(1):3-16. Available from: 132. Khoo HH, Sharratt PN, Das P, Balasubramanian RK, Naraharisetti https://link.springer.com/article/10.1007/BF00047170
- 118. Ho SH, Chen CY, Lee DJ, Chang JS. Perspectives on microalgal CO2-emission mitigation systems - A review. Biotechnol Adv 2011;29(2):189-98. [Internet]. Available from: http:// dx.doi.org/10.1016/j.biotechadv.2010.11.001
- 119. Červený J, Šetlík I, Trtílek M, Nedbal L. Photobioreactor for cultivation and real-time, in-situ measurement of O2 and CO2 exchange rates, growth dynamics and of chlorophyll fluorescence 2009;9(3):247-53.
- 120. Cohen Y, Nisnevitch M, Anker Y. Innovative large-scale photobioreactor for coal propelled power plant effluents treatment. Algal Res [Internet]. 2020;52(September):102101. Available from: 135. Alabi AO, Tampier M BE. Microalgae technologies and processes https://doi.org/10.1016/j.algal.2020.102101
- 121. Yang C, Hua Q, Shimizu K. Energetics and carbon metabolism during growth of microalgal cells under photoautotrophic, mixotrophic and cyclic light-autotrophic/dark-heterotrophic conditions. Biochem Eng J. 2000;6(2):87-102.
- 122. Grobbelaar JU, Nedbal L, Tichý V. Influence of high frequency light/dark fluctuations on photosynthetic characteristics of microalgae photoacclimated to different light intensities and implications for mass algal cultivation. J Appl Phycol. 1996;8(4-5):335 -43.
- 123. Jacob-Lopes E, Gimenes Scoparo CH, Queiroz MI, Franco TT. Biotransformations of carbon dioxide in photobioreactors. Energy Convers Manag [Internet]. 2010;51(5):894–900. Available from: http://dx.doi.org/10.1016/j.enconman.2009.11.027
- 124. You T, Barnett SM. Effect of light quality on production of extracellular polysaccharides and growth rate of Porphyridium cruentum. Biochem Eng J. 2004;19(3):251-58.
- 125. Ravelonandro PH, Ratianarivo DH, Joannis-Cassan C, Isambert A, Raherimandimby M. Influence of light quality and intensity in the cultivation of Spiruling platensis from Toliara (Madagascar) in a closed system. J Chem Technol Biotechnol [Internet]. 2008;83 (6):842-48. Available from: https://onlinelibrary.wiley.com/doi/ abs/10.1002/jctb.1878
- 126. Rosgaard L, de Porcellinis AJ, Jacobsen JH, Frigaard NU, Sakuragi Y. Bioengineering of carbon fixation, biofuels and biochemicals in cyanobacteria and plants. J Biotechnol [Internet]. 2012;162(1):134-47. Available from: http://dx.doi.org/10.1016/ j.jbiotec.2012.05.006
- 127. Gimpel JA, Specht EA, Georgianna DR, Mayfield SP. Advances in microalgae engineering and synthetic biology applications for biofuel production. Curr Opin Chem Biol [Internet]. 2013;17 (3):489-95. Available from: http://dx.doi.org/10.1016/ j.cbpa.2013.03.038
- 128. Chang RL, Ghamsari L, Manichaikul A, Hom EFY, Balaji S, Fu W et al. Metabolic network reconstruction of Chlamydomonas offers insight into light-driven algal metabolism. Mol Syst Biol. 2011;7 (518).
- naMartins CM, Caldana C, AndrésGonzález Barrios F. Analysis of sensitive CO₂ pathways and genes related to carbon uptake and accumulation in *Chlamydomonas reinhardtii* through genomic scale modeling and experimental validation. Front Plant Sci. 2016;7(FEB2016):1-12.
- 130. Tsoutsos T, Kouloumpis V, Zafiris T, Foteinis S. Life cycle assessment for biodiesel production under Greek climate conditions. J Clean Prod [Internet]. 2010;18(4):328–35. Available from: http:// 146. Cadoret JP, Garnier M, Saint-Jean B. Microalgae, Functional Gedx.doi.org/10.1016/j.jclepro.2009.11.002

- trations. Plant Cell Phys [Internet]. 1986;27(4):737–39. Available 131. Gnansounou E, Dauriat A, Villegas J, Panichelli L. Life cycle assessment of biofuels: Energy and greenhouse gas balances. Bioresour Technol [Internet]. 2009;100(21):4919-30. Available from: http://dx.doi.org/10.1016/j.biortech.2009.05.067
 - PK, Shaik S. Life cycle energy and CO₂ analysis of microalgae-tobiodiesel: Preliminary results and comparisons. Bioresour Technol [Internet]. 2011;102(10):5800-07. Available from: http:// dx.doi.org/10.1016/j.biortech.2011.02.055
 - 133. Campbell PK, Beer T, Batten D. Life cycle assessment of biodiesel production from microalgae in ponds. Bioresour Technol [Internet]. 2011;102(1):50-56. Available from: http:// dx.doi.org/10.1016/j.biortech.2010.06.048
- emission of photoautotrophic microorganisms. Eng Life Sci. 134. Kadam KL. Microalgae production from power plant flue gas: Environmental implications on a life cycle basis [Internet]. Contract. 2001. Available from: www.nrel.gov/docs/ fy01osti/29417.pdf
 - for biofuels/ bioenergy production in British Columbia. [Internet]. Canada; 2009. Available from: https:// www.etipbioenergy.eu/databases/reports/69-microalgaetechnologies-processes-for-biofuels-bioenergy-production-inbritish-columbia-current-technology-barriers-to-implementation
 - 136. Habib MAB, Parvin M, Huntington TC, Hasan MR. A review on culture, production and use of Spirulina as food for humans and feeds for domestic animals and fish [Internet]. FAO Fisheries and Aquaculture Circular. No. 1034. Rome; 2008. Available from: http://www.fao.org/3/i0424e/i0424e00.htm
 - 137. Becker EW. Microalgae for human and animal nutrition. In: Richmond A editor. Handbook of Microalgal Culture: Biotechnology and Applied Phycology [Internet]. State Avenue, Ames, Iowa, USA: a Blackwell Publishing Company. 2013;312-51. Available from: http://dl.icdst.org/pdfs/ files/8a0e128e46252f218192dc8d9cfa090e.pdf
 - 138. Todd Lorenz R, Cysewsk GR. Commercial potential for Haematococcus microalgae as a natural source of astaxanthin. Trends Biotechnol [Internet]. 2000;18(4):160-67. Available from: https:// www.sciencedirect.com/science/article/abs/pii/ S0167779900014335
 - 139. Sekar S, Chandramohan M. Phycobiliproteins as a commodity: Trends in applied research, patents and commercialization. J Appl Phycol. 2008;20(2):113-36.
 - 140. Merchant SS, Prochnik SE, Vallon O, Harris EH, Sanderfoot SA, Spalding MH et al. The Chlamydomonas Genome Reveals the Evolution of Key. Natl institutes Heal. 2007;318(5848):245-50.
 - 141. Matsuo T, Ishiura M. Chlamydomonas reinhardtii as a new model system for studying the molecular basis of the circadian clock. FEBS Lett. 2011;585(10):1495-502.
 - 142. Hermsmeier D, Schulz R, Senger H. Formation of light-harvesting complexes of photosystem II in Scenedesmus - II. Different patterns of light-regulation of Lhc-gene expression in green and greening cells. Planta. 1994;193(3):406-12.
 - 143. Yamano T, Fukuzawa H. Carbon-concentrating mechanism in a green alga, Chlamydomonas reinhardtii, revealed by transcriptome analyses. J Basic Microbiol. 2009;49(1):42-51.
- 129. FlaviaWinck V, DavidPáez Melo O, DiegoRiaño-Pachón M, Mari- 144. Rolland N, Atteia A, Decottignies P, Garin J, Hippler M, Kreimer G et al. Chlamydomonas proteomics. Curr Opin Microbiol. 2009;12 (3):285-91.
 - 145. Mühlhaus T, Weiss J, Hemme D, Sommer F, Schroda M. Quantitative shotgun proteomics using a uniform 15N-labeled standard to monitor proteome dynamics in time course experiments reveals new insights into the heat stress response of Chlamydomonas reinhardtii. Mol Cell Proteomics. 2011;10(9):1-27.
 - nomics and Biotechnology. Vol. 64, Advances in Botanical Re-

search. 2012. 285-341.

- 147. Raven JA. Inorganic Carbon Acquisition by Marine Autotrophs. www.sciencedirect.com/science/article/pii/S0065229608602815
- 148. Moroney JV, Somanchi A. How do algae concentrate CO₂ to increase the efficiency of photosynthetic carbon fixation? Plant Physiol. 1999;119(1):9-16.
- 149. Price GD, Coleman JR, Badger MR. Association of carbonic anhydrase activity with carboxysomes isolated from the cyanobacterium Synechococcus PCC7942. Plant Physiol. 1992;100(2):784-93.
- 150. Meyer MT, Itakura AK, Patena W, Wang L, He S, Emrich-Mills T et al. Assembly of the algal CO₂-fixing organelle, the pyrenoid, is guided by a Rubisco-binding motif. bioRxiv. 2020;(November):1-11.
- 151. Yang B, Liu J, Ma X, Guo B, Liu B, Wu T et al. Genetic engineering of the Calvin cycle toward enhanced photosynthetic CO₂ fixation in microalgae. Biotechnol Biofuels. 2017;10(1):1-13.
- 152. Fang L, Lin HX, Low CS, Wu MH, Chow Y, Lee YK. Expression of the Chlamydomonas reinhardtii Sedoheptulose-1,7-bisphosphatase in Dunaliella bardawil leads to enhanced photosynthesis and increased glycerol production. Plant Biotechnol J. 2012;10 (9):1129-35.
- 153. Moroney J V., Ma Y, Frey WD, Fusilier KA, Pham TT, Simms TA et al. The carbonic anhydrase isoforms of Chlamydomonas reinhardtii: Intracellular location, expression and physiological roles. Photosynth Res. 2011;109(1-3):133-49.
- 154. Yamano T, Sato E, Iguchi H, Fukuda Y, Fukuzawa H. Characterization of cooperative bicarbonate uptake into chloroplast stroma in the green alga Chlamydomonas reinhardtii. Proc Natl Acad Sci U S A. 2015;112(23):7315-20.
- 155. Gee CW, Niyogi KK. The carbonic anhydrase CAH1 is an essential component of the carbon-concentrating mechanism in Nannochloropsis oceanica. Proc Natl Acad Sci U S A. 2017;114(17):4537-42.
- 156. Khandavalli LVNS, Lodha T, Abdullah M, Guruprasad L, Chintalapati S, Chintalapati VR. Insights into the carbonic anhydrases and autotrophic carbon dioxide fixation pathways of high CO2 tolerant Rhodovulum viride JA756. Microbiol Res [Internet]. 2018;215 (July):130–40. Available from: https://doi.org/10.1016/ j.micres.2018.07.006
- 157. Fujiwara S, Fukuzawa H, Tachiki A. Structure and differential expression of two genes encoding carbonic anhydrase in Chlamydomonas reinhardtii. Proc Nati Acad Sci USA [Internet]. 1990;87(December):9779-83. Available from: https:// pubmed.ncbi.nlm.nih.gov/2124702/
- 158. Moroney J V., Husic HD, Tolbert NE. Effect of Carbonic Anhydrase reinhardtii. Plant Physiol. 1985;79(1):177-83.
- 159. Winck FV, Arvidsson S, Riaño-Pachón DM, Hempe S, Koseska A, Nikoloski Z, et al. Genome-wide identification of regulatory elegreen alga Chlamydomonas reinhardtii under carbon deprivation. PLoS One. 2013;8(11):1-16.
- 160. Kumar G, Shekh A, Jakhu S, Sharma Y, Kapoor R, Sharma TR. Bioengineering of Microalgae: Recent Advances, Perspectives and Regulatory Challenges for Industrial Application. Front Bioeng Biotechnol. 2020;8(September).
- 161. Winck FV, Melo DOP, Riaño-pachón DM, Martins MCM. Analysis of Sensitive CO 2 Pathways and Genes Related to Carbon Uptake and Accumulation in Chlamydomonas reinhardtii through Genomic Scale Modeling and Experimental Validation. 2016;7 (February):1-12.
- 162. Jungnick N, Ma Y, Mukherjee B, Cronan JC, Speed DJ, Laborde SM et al. The carbon concentrating mechanism in Chlamydomo-

nas reinhardtii: Finding the missing pieces. Photosynth Res. 2014;121(2-3):159-73.

- Adv Bot Res [Internet]. 1997;27:85–209. Available from: https:// 163. Park Y II, Karlsson J, Rojdestvenski I, Pronina N, Klimov V, Öquist G, et al. Role of a novel photosystem II-associated carbonic anhydrase in photosynthetic carbon assimilation in Chlamydomonas reinhardtii. FEBS Lett. 1999;444(1):102-05.
 - 164. Villarejo A, Shutova T, Moskvin O, Forssén M, Klimov V V., Samuelsson G. A photosystem II-associated carbonic anhydrase regulates the efficiency of photosynthetic oxygen evolution. EMBO J. 2002;21(8):1930-38.
 - 165. Spalding MH. Microalgal carbon-dioxide-concentrating mechanisms: Chlamydomonas inorganic carbon transporters. J Exp Bot. 2008;59(7):1463-73.
 - 166. Kuo CM, Lin TH, Yang YC, Zhang WX, Lai JT, Wu HT, et al. Ability of an alkali-tolerant mutant strain of the microalga Chlorella sp. AT1 to capture carbon dioxide for increasing carbon dioxide utilization efficiency. Bioresour Technol [Internet]. 2017;244:243-51. Available http://dx.doi.org/10.1016/ from: j.biortech.2017.07.096
 - 167. Fu W, Nelson DR, Mystikou A, Daakour S, Salehi-Ashtiani K. Advances in microalgal research and engineering development. Curr Opin Biotechnol [Internet]. 2019;59:157-64. Available from: https://doi.org/10.1016/j.copbio.2019.05.013
 - 168. Vandepoele K, Van Bel M, Richard G, Van Landeghem S, Verhelst B, Moreau H, et al. pico-PLAZA, a genome database of microbial photosynthetic eukaryotes. Environ Microbiol. 2013;15(8):2147-53.
 - 169. Zheng H, Chien C, Hsu BJ, Liu T, Chen CN, Chang W. AlgaePath: comprehensive analysis of metabolic pathways using transcript abundance data from next-generation sequencing in green algae. BMC Genomics [Internet]. 2014;15(196):1–12. Available from: https://bmcgenomics.biomedcentral.com/articles/10.1186/1471-2164-15-196
 - 170. Aoki Y, Okamura Y, Ohta H, Kinoshita K, Obayashi T. ALCOdb: Gene coexpression database for microalgae. Plant Cell Physiol. 2016;57(1):e3.
 - 171. Bae S, Park J, Kim JS. Cas-OFFinder: A fast and versatile algorithm that searches for potential off-target sites of Cas9 RNAguided endonucleases. Bioinformatics. 2014;30(10):1473-75.
 - 172. Lei Y, Lu L, Liu HY, Li S, Xing F, Chen LL. CRISPR-P: A web tool for synthetic single-guide RNA design of CRISPR-system in plants. Mol Plant. 2014;7(9):1494-96.
 - 173. Liu H, Ding Y, Zhou Y, Jin W, Xie K, Chen LL. CRISPR-P 2.0: An Improved CRISPR-Cas9 Tool for Genome Editing in Plants. Mol Plant [Internet]. 2017;10(3):530-2. Available from: http:// dx.doi.org/10.1016/j.molp.2017.01.003
- Inhibitors on Inorganic Carbon Accumulation by Chlamydomonas 174. Liu T, Pan S, Li Y, She Q. Type III CRISPR-Cas System: Introduction And Its Application for Genetic Manipulations. Curr Issues Mol Biol [Internet]. 2017;26:1-14. Available from: https:// europepmc.org/article/med/28879852
- ments and reconstruction of gene regulatory networks of the 175. Park J, Bae S, Kim JS. Cas-Designer: A web-based tool for choice of CRISPR-Cas9 target sites. Bioinformatics. 2015;31(24):4014-6.
 - 176. Stemmer M, Thumberger T, Del Sol Keyer M, Wittbrodt J, Mateo JL. CCTop: An intuitive, flexible and reliable CRISPR/Cas9 target prediction tool. PLoS One. 2015;10(4):1-11.
 - 177. Labun K, Montague TG, Gagnon JA, Thyme SB, Valen E. Chopchop v2: a web tool for the next generation of CRISPR genome engineering. Nucleic Acids Res. 2016;44(W1):W272-6.
 - 178. Montague TG, Cruz JM, Gagnon JA, Church GM, Valen E. Chopchop: A CRISPR/Cas9 and TALEN web tool for genome editing. Nucleic Acids Res. 2014;42(W1):1-7.
 - 179. Doench JG, Fusi N, Sullender M, Hegde M, Vaimberg EW, Donovan KF, et al. Optimized sgRNA design to maximize activity and minimize off-target effects of CRISPR-Cas9. Nat Biotechnol [Internet].

nbt.3437

- 180. Park J, Lim K, Kim JS, Bae S. Cas-analyzer: An online tool for assessing genome editing results using NGS data. Bioinformatics. 2017;33(2):286-8.
- 181. Kim D, Bae S, Park J, Kim E, Kim S, Yu HR, et al. Digenome-seq: Genome-wide profiling of CRISPR-Cas9 off-target effects in human cells. Nat Methods [Internet]. 2015;12(3):237-43. Available from: http://dx.doi.org/10.1038/nmeth.3284
- 182. Tanwar A, Sharma S, Kumar S. Targeted genome editing in algae using CRISPR/Cas9. Indian J Plant Physiol [Internet]. 2018;23 (4):653-69. Available from: https://doi.org/10.1007/s40502-018-0423-3
- 2005;137(2):410-27.
- 184. Tettelin H, Masignani V, Cieslewicz MJ, Donati C, Medini D, Ward NL, et al. Erratum: Genome analysis of multiple pathogenic iso- 191. Fulke A, Chakrabarti T, Kannan K. CO2 Sequestration by Microallates of Streptococcus agalactiae: Implications for the microbial "pan-genome" (Proceedings of the National Academy of Sciences of the United States of America (September 27, 2005) 102, 39 (13950-13955)). Proc Natl Acad Sci U S A. 2005;102(45):16530.
- 185. Du YC, Peddi SR, Spreitzer RJ. Assessment of structural and functional divergence far from the large subunit active site of ribulose -1,5-bisphosphate carboxylase/oxygenase. J Biol Chem. 2003;278 (49):49401-05.

- 2016;34(2):184–91. Available from: http://dx.doi.org/10.1038/ 186. Spreitzer RJ, Peddi SR, Satagopan S. Phylogenetic engineering at an interface between large and small subunits imparts landplant kinetic properties to algal Rubisco. Proc Natl Acad Sci U S A. 2005;102(47):17225-30.
 - 187. Blaby-Haas CE, Merchant SS. Comparative and functional algal genomics. Annu Rev Plant Biol. 2019;70(August):605-38.
 - 188. Radakovits R, Jinkerson RE, Darzins A, Posewitz MC. Genetic engineering of algae for enhanced biofuel production. Eukaryot Cell. 2010;9(4):486-501.
 - 189. Liolios K, Mavromatis K, Tavernarakis N, Kyrpides NC. The genomes on line database (GOLD) in 2007: Status of genomic and metagenomic projects and their associated metadata. Nucleic Acids Res. 2008;36(SUPPL. 1):475-79.
- 183. Grossman AR. Paths toward algal genomics. Plant Physiol. 190. Wang Q, Lu Y, Xin Y, Wei L, Huang S, Xu J. Genome editing of model oleaginous microalgae Nannochloropsis spp. by CRISPR/Cas9. Plant J. 2016;88(6):1071-81.
 - gae: Advances and Perspectives. In: Vasconcelos JR, Gerken H, Liu J, Sun Z, editors. Recent Advances in Microalgal Biotechnology [Internet]. OMICS International; 2016. Available from: https:// www.esciencecentral.org/ebooks/ebookdetail/recent-advancesin-microalgal-biotechnology