Pollen viability and incompatibility in indigenous rice bean (Vigna umbellata (Thunb.) Ohwi & Ohashi)

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Abstract
Pollen viability, germination and compatibility are essential in determining the success of pollination and seed setting of high-valued crops. Rice bean (Vigna umbellata (Thunb.) Ohwi & Ohashi) is an underutilized and unexplored indigenous legume with high potential for commercial production. In this study, pollen quality, viability, germination rate and incompatibility among selected six rice bean (V. umbellata) accessions from Barili, Cebu, Philippines were evaluated to determine the barriers and effective pollination habit for increased productivity while retaining the important traits, including high tolerance in poor soils, superior climatic resilience and resistance to pest and diseases. Results of acetocarmine calorimetric assay showed that rice beans' (V. umbellata) pollens are highly viable, with accessions VU 004 (56.33 ± 4.91%) and VU 007 (54.34 ± 4.53%) having the optimum viability rate. Brewbaker and Kwack medium treated with 0.2 g.l⁻¹ and 0.3 g.l⁻¹ boric acid (H₃BO₃) enhanced the germination rate in vitro (11.56 ± 5.53% and 9.47 ± 6.50% respectively). Bud (14.96 ± 1.53%) and post-anthesis pollens (10.28 ± 0.94%) have optimum germination rate in 0.2 g.l⁻¹ boric acid media, while anthesis pollens are suitable in media supplemented with sucrose and boric acid alone (12.20 ± 1.50%) and with 0.1 g.l⁻¹ myo-inositol supplementation (8.49 ± 1.86%). Pollination test revealed that rice bean accessions have high self-compatibility (50.76 ± 3.45%) and low cross-compatibility (26.57 ± 2.49%). The findings provide an important background in understanding the pollen quality and intraspecific interaction among indigenous rice bean (V. umbellata) accessions in Barili, Cebu to improve production and hybridization.

Keywords
Pollen grains, indigenous crop, pollen viability, rice bean, self- and cross incompatibility

Introduction
In plant breeding, pollen grains play a vital role in developing a high-quality variety. Pollen carries the male gametes responsible for successful genetic exchange and ensures the survival of the species (1). Effective pollination is an essential requirement for fruit development and seed setting in plants (2). Therefore, an insight into the pollen biology are important in order to improve crop productivity (3); such that knowledge in pollen germinability, viability and the compatibility is a convenient and reliable aspect in future breeding programs (4).
Pollination in higher plants enables seed development through pollen-pistil interaction (5). However, compatibility and receptibility barriers are problems in the success of the process. Pollen incompatibility is a pre-fertilization barrier that hinders successful pollination due to: (a) rejection of pollen grains from a male donor by the female receptor flower, (b) failure of pollen grain germination, (c) failure of the pollen tube to reach the ovary and (d) unsuccessful fusion of male and female gametes (6). Incompatibility in plants can be classified as self-incompatibility and cross-incompatibility. Self-incompatibility (SI) occurs when pollen is recognized and rejected by the stigma of plants of the same cultivars (7). On the other hand, cross-incompatibility (CI) is the rejection of pollen by pistil due to lack of recognition of the pollen from the foreign origin (8). Self-incompatibility restricts inbreeding due to self-discrimination of pollen by pistil, while cross-incompatibility (CI) limits hybrid zygote formation between species of different gene pool (9, 10).

Interspecific incompatibility in Vigna species is influenced by the parental genotype and meiotic irregularities (6). It was reported that the crossability of interspecific hybrids among Vigna radiata, Vigna mungo and Vigna umbellata and identified fertilization barriers in some crosses, which resulted to embryo degeneration and abnormal seed development (11). They further identified incompatibility barriers among self- and cross pollinated Vigna species such as delayed entry of pollen tube to the ovules and delayed and failure in nuclear division of endosperm. Reports are on the low crossability and pollen fertility between V. radiata and V. mungo crosses due to genotypic and ecological differences and meiotic abnormalities (12). One of the determining factors in successful fertilization and germination is pollen viability (13). Pollen viability is vital in seed setting (14), especially in leguminous and grain crops such as Vigna spp. It was reported that over 90% genetically viable pollens of Vigna unguiculata however the development in in vitro medium resulted to low germination (15). In bambara groundnut (Vigna subterreneo (L.) Verdc.), rapid loss of pollen viability and germi-nability was reported due to several environmental factors including temperature, humidity and organic solvents used (16).

Like all other legumes, Vigna species is essential in biological nitrogen fixation. In addition, some Vigna species, especially the wild and exotic has resistance to pests and diseases (17, 18). Rice bean (Vigna umbellata (Thunb.) Ohwi & Ohashi), for instance, has been reported to exhibit complete resistance to bruchids, small beetles attacking vignas (6). The most common and widely spread disease among Vigna species, with major economic significance, is the Yellow Mosaic Disease (YMD) caused by Begomoviruses (19). Significant evidences were reported on the high degree of resistance of rice bean (V. umbellata) to the disease compared to other Vigna species (20, 21). Rice bean (V. umbellata) has also superior nutritional value comparable to other legume grains (22). Previous studies reported that the crop is rich in amino acids, vitamins, dietary fiber, minerals such as calcium, iron and zinc and antioxidants such as polyphenols and tannins (23, 24).

Although rice bean (V. umbellata) has superior traits and nutritional content, the crop is a less known and regarded as underutilized legume (24). In addition, underutilized crops are species grown extensively in the past or have a potential in the future either for agronomic, genetic or economic reasons but are currently grown in a limited area. Rice bean (V. umbellata) is among these crop species that did not receive major attention for improvement and thus scientific knowledge about them is limited (22).

Result of initial fragmented studies on its traits provides insights into its potential as a major agricultural crop and as an important genetic resource for breeding. Information on the mode of transfer and retention of these essential genes to other Vigna sp. (interspecific crossing) or of the same species (intraspecific breeding) is a prerequisite to a crop improvement programme. However, crossability barriers in Vigna sp. hinder the development of interspecific hybrids due to the difference in gene pool, resulting in low fertilization and number of seeds (25). Although several attempts have been conducted to overcome these barriers, reproductive and floral biology has yet been fully explored among intraspecific crosses of rice bean (V. umbellata). Understanding the pollination habit and reproductive mechanism among crop varieties are essential to increase the chances of successful fertilization (2, 26). Since pollen grain carries the traits (27), investigating the pollen biology and mechanism provides essential knowledge in the development of inbreds and hybrids with superior quality. Hence, the objective of this study was to evaluate the pollen quality and incompatibility among native accessions of Vigna umbellata (Thunb.) Ohwi & Ohashi for improved production.

Materials and Methods

Plant materials, experimental design and crop management

A total of six rice bean accessions obtained from Barili, Cebu, Philippines and maintained by the Center for Studies in Biotechnology in Cebu Technological University Barili Campus was used in this study. The accessions were assigned with code (VU 001 (Brgy. Balao), VU 002 (Brgy. Kangdampas-San Jose), VU 004 (Brgy. Pangpang), VU 005 (Brgy. Poblacion-market), VU 006 (Brgy. Budbud) and VU 007 (Brgy. Kangdampas-Centro)).

Healthy seeds were used as planting material and propagated in an experimental pot laid out in a Randomized Complete Block Design (RCBD) with three replications and 10 samples per replication.

The experimental setup was maintained following farmer’s practice. Soil planting medium, applied with 25 g chicken dung per pot, was prepared seven days before...
planting. Cultural practices such as weeding using bolo, watering every 4:00 PM and training of vines to climb in their respective stakes were done manually. The setup was covered with a plastic barrier prior to flowering to avoid pollen contamination from other accessions. Flowers were assessed for pollen viability and germination (in vitro and in vivo) assays.

**Pollen viability test**

Viable pollen count was estimated using acetic acid cal- lorimetric assay. Around 32,046 pollen samples were collected in three different plants of the same accession (six accessions) and were spread to a glass slide. A drop of 2% acetic acid solution was added to the slide and was examined under a binocular compound light microscope. pollen is considered viable if it appeared red color and nonviable if colorless after staining (28). Pollen viability was computed using the standard formula (29):

\[
\text{Pollen viability} = \left( \frac{\text{number of viable pollen}}{\text{total number of pollen}} \right) \times 100
\]

**In vitro pollen germination test**

Pollen collection was done in 3 different flowering stages: pre-anthesis (bud phase), anthesis (flower fully open) and post-anthesis (6 hours after anthesis). Each sample was germinated in a liquid Brewbaker and Kwack (BK) media (30) with different concentrations of boric acid (H3BO3) and varying concentrations of myoinositol (Table 1). Then it was incubated at a controlled temperature (27 °C) for 24 hrs under dark condition. Around 53,873 pollen samples were examined in a binocular compound light microscope (Olympus CX31, Japan) and were considered germinated if the diameter of the pollen tube is equal to or larger than the pollen diameter (31). All setup was replicated three times and the germination rate was estimated using the formula (29):

\[
\text{Pollen germination rate} = \left( \frac{\text{number of germinated pollen}}{\text{total number of pollen}} \right) \times 100
\]

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Composition</th>
</tr>
</thead>
<tbody>
<tr>
<td>T0</td>
<td>Distilled water (control)</td>
</tr>
<tr>
<td>T1</td>
<td>100 g.L⁻¹ sucrose + 0.1 g.L⁻¹ H3BO3 + 0.3 g.L⁻¹ CaNO3 + 0.4H2O + 0.2 g.L⁻¹ MgSO4 + 0.2H2O + 0.1 g.L⁻¹ KNO3 (Standard BK (30) medium)</td>
</tr>
<tr>
<td>T2</td>
<td>100 g.L⁻¹ sucrose + 0.2 g.L⁻¹ H3BO3 + 0.3 g.L⁻¹ CaNO3 + 0.4H2O + 0.2 g.L⁻¹ MgSO4 + 0.2H2O + 0.1 g.L⁻¹ KNO3</td>
</tr>
<tr>
<td>T3</td>
<td>100 g.L⁻¹ sucrose + 0.3 g.L⁻¹ H3BO3 + 0.3 g.L⁻¹ CaNO3 + 0.4H2O + 0.2 g.L⁻¹ MgSO4 + 0.2H2O + 0.1 g.L⁻¹ KNO3</td>
</tr>
<tr>
<td>T4</td>
<td>100 g.L⁻¹ sucrose + 0.5 g.L⁻¹ H3BO3 (modified Gauliche et al. (28) medium)</td>
</tr>
<tr>
<td>T5</td>
<td>100 g.L⁻¹ sucrose + 0.5 g.L⁻¹ H3BO3 + 0.1 g.L⁻¹ myo-inositol</td>
</tr>
<tr>
<td>T6</td>
<td>100 g.L⁻¹ sucrose + 0.5 g.L⁻¹ H3BO3 + 0.2 g.L⁻¹ myo-inositol</td>
</tr>
</tbody>
</table>

**Data collection and statistical analysis**

All data were recorded and photographed. Data were analyzed using IBM SPSS Statistics version 20. For each quantitative variable, data were subjected to analysis of variance (p < 0.05). Individual treatment means were analyzed using post-hoc test (Duncan’s Multiple Range Test) and expressed with standard error of means (SEM).

**Results and Discussion**

**Pollen viability assay**

Pollen viability is defined as the ability of the pollen grains to germinate (36). Viability is often associated with chromosome arrangement (37) and meiotic regularities (38, 39). Acetocarmine stains the cytoplasm and the chromatin material in the nuclei of viable pollen (40). Viable pollen retains the stain, which appears pink to deep red, while sterile pollen does not retain any stain and appears transparent (Fig. 1). The sterility of pollen has been observed in dicots...
having irregular chromosome segregation, abnormal sporad formation and unequal-sized fertile pollen grains (41). Cytomixis on meiotic behavior also affects the viability of pollen as observed in wild Himalayan poppy (Meconopsis aculeata Royle) (42). The current study revealed that V. umbellata pollen has a tricolporate pollen – having three distinct pores, which is common to all members of Papilionoideae (43-45).

The acetocarmine calorimetric assay revealed that accessions VU 004 (56.33 ± 4.91%) and VU 007 (54.34 ± 4.53%) had the highest pollen viability rate while VU 005 (28.33 ± 4.03%) obtained the lowest rate (Fig. 2). The average pollen viability rate for all accessions is 44.90 ± 3.14%. The overall rate is 49.82% lower compared to Yard Long Bean (Vigna unguiculata subsp. sesquipedalis) (46).

Pollen viability is directly associated with pollen quality and is connected with reproductive biology, determining the effective seed set after pollination (47). The significantly high pollen viability rate in accessions VU 004 and VU 007 increases the chance of effective pollination and higher fertilization rate, indicating strong sexual reproduction and successful seed formation. It was reported that stronger pollen viability is an effective measure for seed setting enhancement (48). Whereas, low pollen viability increases the failure in pollination, which further results in low seed set and reduced breeding efficacy (49).

**Optimization of pollen germination medium (PGM) in rice bean (V. umbellata)**

*In vitro* pollen germination test is another method of testing the viability of pollen in artificial media. The medium is generally composed of sugar and ionic salts, such as calcium nitrate, magnesium sulfate and boric acid (1, 28, 30, 50). Organic supplements such as polyethylene glycol, vitamins, amino acids and plant growth hormones were also incorporated to increase the rate of pollen germination (51-53). BK (30) medium is the most widely used pollen germination...
media (PGM) which was tested and was found to be suitable in 86 plant species (30, 54).

In the current study, the response of pollen grains of different rice bean (V. umbellata) accessions to boron (T1 – T5) and myo-inositol (T6 – T9) were assessed. A relatively lower overall germination rate (6.89 ± 7.09%) was observed among accessions in the artificial media (Table 2). The T5 showed promising results with 11.56 ± 5.53% average germination rate for all accessions, with VU 007 having the highest rate (15.96 ± 2.72%). Accessions VU 001, VU 002, VU 005, and VU 007 showed promising pollen germination response in BK (30) media. On the other hand, VU 004 and VU 006 both have optimum pollen germination response in BK (30) media supplemented with 0.2 g.l⁻¹ (T5) and 0.3 g.l⁻¹ (T6) boric acid and PGM containing sucrose and boric acid alone (T6). The effectiveness of BK (30) media has been reported by previous study on black gram (Vigna mungo L. VAR. DPU-88-31) with 73.00% average pollen germination (55). Whereas around 44.25 to 55.00% pollen germination was observed among Momordica sp. germinated media containing sucrose and boric acid alone (3).

**Effects of boron and myo-inositol on pollen development in pollen germination media (PGM)**

Bursting and lysis of pollen tube and pollen grains were also observed in different media (Fig. 3). In addition, T1, T3, and T5, where bursting were observed, has significantly higher pollen germination rate. Similar results were observed in pollen germination in mango, where bursting was observed in medium treated with 1.0 ml.l⁻¹ boric acid (56). The treatment also resulted in 37.34% higher germination rate compared to medium without boric acid. In comparison, medium treated with high concentrations of boron resulted in shorter, swelled and increase in diameter of pollen tubes in Malus domestica trees (57). In the present study, 50% increase in boron, in the form of boric acid (T5), also increases the pollen germination rate at around 10.21%. However, an 80% (T4) increase of the salt caused 17.73% decrease in germination rate. Similar findings were reported in Picea meyeri PGM, treated with 0.001 to 0.01% boron acid, where germination rates ranged from 38 to 61% (58).

The role of myo-inositol in pollen tube development has been reported in previous studies (59, 60). Higher plants utilize uronosyl and pentosyl, a derivative of myo-inositol present in the cell wall, for polysaccharide formation and synthesis of exudates found on the stigma and style (61-63). While in this study, myo-inositol inhibits germination by 54.68 to 98.71% (T5–T6), in addition to no germination observed in some rice bean (V. umbellata) accessions. The lower germination result can be attributed to the regulating capacity of inositol polyphosphate kinase reported in Arabidopsis (AtIPK2a). Previous study revealed that the reduction in AtIPK2a transcript levels resulted in enhanced pollen germination and pollen tube growth of transgenic Arabidopsis thaliana, under nonoptimal low Ca²⁺ concentrations culture medium (64).

**Effects of stages in flowering in in vitro pollen germination**

To determine the most efficient phase of rice bean for pollen germination, pollen grains from different flowering stages (bud phase, anthesis and post-anthesis) were germinated in the artificial media. Germinated pollen grains developed longer pollen tube compare to the diameter of the ungerminated pollen itself (Fig. 4).

Individual treatment composition showed variable effects of pollen germination at different flowering stages. BK (30) medium with 0.2 g.l⁻¹ boric acid (T5) exhibited a relatively higher germination rate with 34.68%, combining the

### Table 2. Pollen germination rate among rice bean (V. umbellata) accessions in different medium compositions.

<table>
<thead>
<tr>
<th>TR</th>
<th>ACCESSION (± SEM)</th>
<th>Average Germination Rate (± SEM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>EA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TM</td>
<td>VU 001</td>
<td>VU 002</td>
</tr>
<tr>
<td>T1</td>
<td>1.18 ± 1.8 abc</td>
<td>1.55 ± 0.69 c</td>
</tr>
<tr>
<td>T2</td>
<td>9.84 ± 1.30 a</td>
<td>7.18 ± 1.12 ab</td>
</tr>
<tr>
<td>T3</td>
<td>2.97 ± 1.90 a</td>
<td>11.59 ± 1.31 a</td>
</tr>
<tr>
<td>T4</td>
<td>4.90 ± 1.68 abc</td>
<td>6.94 ± 1.52 ab</td>
</tr>
<tr>
<td>T5</td>
<td>5.19 ± 2.57 ab</td>
<td>7.27 ± 1.33 ab</td>
</tr>
<tr>
<td>T6</td>
<td>2.68 ± 1.52 c</td>
<td>3.68 ± 1.94 bc</td>
</tr>
<tr>
<td>T7</td>
<td>0.00</td>
<td>0.64 ± 0.64 c</td>
</tr>
</tbody>
</table>

![Fig. 3](image-url) Normal (A) and burst/lysed (B-F) pollen tube and pollen grains as a response to varying media formulation in rice bean (V. umbellata) using a compound light microscope at 400x total magnification.

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germination rate in all phases (Fig. 5). The same treatment was also observed to have a higher germination rate in bud pollens (14.96 ± 1.53%) and post-anthesis pollens (10.28 ± 0.94%). Boron, in the form of boric acid, plays an essential role in in vitro pollen germination by regulating the pollen tube growth through its ability to stimulate the plasma membrane H⁺-ATPase (65). Contrasting results were obtained in in vitro pollen germination in Chinese fir (Cunninghamia lanceolata L.) using the same concentration of boric acid, which completely inhibited pollen germination (66). Further studies need to be done that not using the optimal concentration of boric acid hinders pollen germination. Based on the current findings and previous reports, the optimum boron requirement varied in every species (57).

On the other hand, anthesis pollens have the highest germination rate in modified Gaaliche et al. (28) media alone (T₄) (12.20 ± 1.50%) and with 0.1 g L⁻¹ myo-inositol (T₃) supplementation (8.49 ± 1.86%). Myo-inositol has been incorporated in several growth media and has the ability to stimulate pollen tube growth in some plant species and cell wall biosynthesis (60, 67, 68). The effects of PGM composition in pollen germination at different flowering stages has also been reported in passion fruit (Passiflora spp.), having 24.08 to 54.87% higher in anthesis compared to post and pre-anthesis (69).

**Incompatibility in rice bean (V. umbellata)**

Compatibility refers to the ability of the stigma to recognize and accepts the pollen grains. The process is highly selective and a factor determining the quantitative and qualitative productivity aspects of agronomic crops (70). In the current study, each accession was cross-pollinated in vivo, and germination rate and pollen penetration in style were determined. The result showed 26.57 ± 2.49% germination rate among accession crosses. Highest and significant mean rate (p ≤ 0.05) was observed in three cross-pairs (male x female) namely VU 002 x VU 004 (55.12 ± 11.35%), VU 004 x VU 007 (54.53 ± 6.79%), and VU 002 x VU 005 (51.83 ± 5.49%) (Table 3). In addition, VU 004 x VU 007 has the highest pollen penetration observed in style. The table also showed no germination observed in some crosses (VU 001 x VU 004, VU 005 x VU 002, VU 006 x VU 004, VU 006 x VU 005, and VU 007 x VU 008). Previous reports mentioned that failure germination could be caused by pollen tubes that are unable to penetrate the stigma and style, embryo abortion during embryogenesis, and the most common in legumes, embryo degeneration (33). The intra-specific incompatibility observed in some rice bean (V. umbellata) accession is due to swelling of pollen tubes (Fig. 6). Swollen tube was also observed in wild and cultivated sesame (Sesamum indicum) (71). Delayed pollen tube growth along with other structural abnormalities like twisting, swelling, high branching, bi-furcated tip and variation in callose form were also noticed in the interspecific crosses of Abelmoschus spp. (72).

![Fig. 4](https://plantsciencetoday.online) Germinated pollen (pollen tube length is greater than pollen diameter) in different media during bud phase (A), anthesis (B), and post-anthesis (C) using a compound light microscope at 400x total magnification.

![Fig. 5](https://plantsciencetoday.online) Pollen germination rate among rice bean (V. umbellata) accessions in different stages of flowering.

<table>
<thead>
<tr>
<th>PARENT COMBINATION</th>
<th>Female (Pollen Receptor)</th>
<th>Male (Pollen Source)</th>
<th>GERMINATION RATE (± SEM) IN STIGMA</th>
<th>POLLEN PENETRATION IN A SINGLE STYLE</th>
</tr>
</thead>
<tbody>
<tr>
<td>VU 001</td>
<td>VU 002</td>
<td>46.24 ± 11.53</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 001</td>
<td>VU 004</td>
<td>0.00 ± 1.31</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>VU 001</td>
<td>VU 005</td>
<td>6.30 ± 6.30</td>
<td>±</td>
<td></td>
</tr>
<tr>
<td>VU 001</td>
<td>VU 006</td>
<td>2.29 ± 2.29</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>VU 001</td>
<td>VU 007</td>
<td>3.99 ± 3.99</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>VU 002</td>
<td>VU 001</td>
<td>3.62 ± 3.62</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>VU 002</td>
<td>VU 004</td>
<td>55.12 ± 11.35</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 002</td>
<td>VU 005</td>
<td>51.83 ± 5.49</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 002</td>
<td>VU 006</td>
<td>43.49 ± 2.37</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 002</td>
<td>VU 007</td>
<td>41.75 ± 3.27</td>
<td>++</td>
<td></td>
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<tr>
<td>VU 004</td>
<td>VU 001</td>
<td>47.58 ± 1.12</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 004</td>
<td>VU 002</td>
<td>42.63 ± 9.68</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>VU 004</td>
<td>VU 005</td>
<td>42.56 ± 6.41</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>VU 004</td>
<td>VU 006</td>
<td>34.39 ± 6.20</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>VU 004</td>
<td>VU 007</td>
<td>54.53 ± 6.79</td>
<td>+++</td>
<td></td>
</tr>
<tr>
<td>VU 005</td>
<td>VU 001</td>
<td>48.39 ± 7.97</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 005</td>
<td>VU 002</td>
<td>0.00 ± 1.31</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>VU 005</td>
<td>VU 004</td>
<td>6.67 ± 6.67</td>
<td>±</td>
<td></td>
</tr>
<tr>
<td>VU 005</td>
<td>VU 006</td>
<td>8.23 ± 8.23</td>
<td>++</td>
<td></td>
</tr>
<tr>
<td>VU 005</td>
<td>VU 007</td>
<td>49.60 ± 5.45</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>VU 006</td>
<td>VU 001</td>
<td>34.18 ± 18.77</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>VU 006</td>
<td>VU 002</td>
<td>3.24 ± 3.24</td>
<td>±</td>
<td></td>
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<tr>
<td>VU 006</td>
<td>VU 004</td>
<td>0.00 ± 1.31</td>
<td>-</td>
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<tr>
<td>VU 006</td>
<td>VU 005</td>
<td>0.00 ± 1.31</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>VU 006</td>
<td>VU 007</td>
<td>48.87 ± 10.10</td>
<td>±</td>
<td></td>
</tr>
<tr>
<td>VU 007</td>
<td>VU 001</td>
<td>37.98 ± 10.04</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>VU 007</td>
<td>VU 002</td>
<td>27.81 ± 1.11</td>
<td>±</td>
<td></td>
</tr>
<tr>
<td>VU 007</td>
<td>VU 004</td>
<td>50.00 ± 1.31</td>
<td>±</td>
<td></td>
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<tr>
<td>VU 007</td>
<td>VU 005</td>
<td>5.65 ± 5.65</td>
<td>±</td>
<td></td>
</tr>
<tr>
<td>VU 007</td>
<td>VU 006</td>
<td>0.00 ± 1.31</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>Average</td>
<td></td>
<td>26.57 ± 2.49</td>
<td>±</td>
<td></td>
</tr>
</tbody>
</table>
average germination rate of self-pollinated accessions is 47.66% higher than the cross-pollinated accessions. Like cowpea (*Vigna unguiculata* (L.) Walp.), rice bean (*V. umbellata*) is self-pollinated with somatic chromosome number of 22 (73, 74), resulting in higher self-compatibility and pollen tube growth. Similar results were found out in the preceding studies on *in vivo* pollen tube growth of self-pollinated *Vigna* spp. with normal and high pollen grain germination (75).

**Conclusion**

Based on the acetocarmine calorimetric assay findings, the pollen grains of indigenous accessions of rice beans (*V. umbellata*) are highly viable, with accessions VU 004 and VU 007 having the optimum viability rate. In general, BK medium treated with 0.2 g.1⁻¹ and 0.3 g.1⁻¹ boric acid enhances the germination rate *in vitro*. Bud and post-anthesis pollens have optimum germination rate when supplemented at 0.2 g.1⁻¹ boric acid media, while anthesis pollens are suitable in myo-inositol supplemented media. Rice bean (*V. umbellata*) accessions have lower interspecific compatibility but have higher self-compatibility.

**Supplementary data**

**Table 1**. Details of rice bean (*V. umbellata*) accessions used in the study.

**Fig. 1**. Geographical location of indigenous rice bean (*V. umbellata*) accession collections.

**Fig. 6**. Normal (A) and swollen (B-D) pollen tube (pointed by an arrow) in incompatible inter-specific crosses (male x female) of rice bean (*V. umbellata*) using a compound light microscope at 400x total magnification.

No significant difference was observed among self-pollinated accessions (Table 4); however, the highest germination rate was observed in VU 001 (64.01 ± 1.83%). The

**Table 4.** Self-compatible pollen-stigma interaction and style penetration in rice bean (*V. umbellata*) accessions.

<table>
<thead>
<tr>
<th>PARENT COMBINATION</th>
<th>GERMINATION RATE (+ SEM) IN STIGMA</th>
<th>POLLEN PENETRATION IN A SINGLE STYLE</th>
</tr>
</thead>
<tbody>
<tr>
<td>VU 001 VU 001</td>
<td>64.01 ± 1.83</td>
<td>±</td>
</tr>
<tr>
<td>VU 002 VU 002</td>
<td>47.44 ± 4.92</td>
<td>±</td>
</tr>
<tr>
<td>VU 004 VU 004</td>
<td>45.33 ± 13.32</td>
<td>±</td>
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<tr>
<td>VU 005 VU 005</td>
<td>43.94 ± 7.96</td>
<td>–</td>
</tr>
<tr>
<td>VU 006 VU 006</td>
<td>54.85 ± 10.98</td>
<td>±</td>
</tr>
<tr>
<td>VU 007 VU 007</td>
<td>48.98 ± 8.67</td>
<td>±</td>
</tr>
<tr>
<td><strong>Average</strong></td>
<td><strong>50.76 ± 3.45</strong></td>
<td><strong>±</strong></td>
</tr>
</tbody>
</table>

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**Authors contributions**

MLDP, JAP and MCN designed the study, led and supervised the experimental setups, performed data analysis and checked the manuscript. JHR assisted the experiments, collected data and drafted the manuscript.

**Compliance with ethical standards**

**Conflict of interest**: Authors do not have any conflict of interests to declare.

**Ethical issues**: None.

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