



# RESEARCH ARTICLE

# Photosynthetic parameters change in *Lycopersicon esculentum* leaves under nutrient deficiencies

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# Abstract

Lycopersicon esculentum leaves cultivated hydroponically for 24 and 48 hrs with various specific mineral deficits had their photosynthetic characteristics examined. After 24 hrs of K<sup>+</sup>, NO<sub>3</sub><sup>-</sup>, and PO<sub>4</sub><sup>2-</sup> deficiency, a substantial induction of net photosynthetic rate was observed. The net photosynthetic rate of  $SO_{4^{2-}}$ ,  $Mg^{2+}$ ,  $Fe^{2+}$ ,  $NO_{3^{-}}$ ,  $Ca^{2+}$  and  $PO_{4^{2-}}$  deficits was significantly induced by the 48 hr exposure. After 24 hrs of deficiencies in  $SO_4^{2-}$ ,  $Mg^{2+}$ ,  $Fe^{2+}$ ,  $NO_3^{-}$ , Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup>, stomata conductance was dramatically increased. Deficiencies in  $SO_4^{2-}$ ,  $Fe^{2+}$ ,  $NO_3^{-}$ ,  $Ca^{2+}$  and  $PO_4^{2-}$  were continuously induced over 48 hrs. After 24 hrs of SO<sub>4</sub><sup>2-</sup>, Fe<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup> deficiencies, intercellular CO<sub>2</sub> concentration shows a considerable induction. After 48 hrs of K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, Mg<sup>2+</sup> and NO<sub>3</sub>-deficits, this behavior remained strongly induced. Water use efficiency considerably decreased in response to these changes after 24 hrs of SO<sub>4</sub><sup>2-</sup>, Fe<sup>2+</sup>, NO<sub>3</sub><sup>-</sup> and PO<sub>4</sub><sup>2-</sup> deficiencies and this effect continued after 48 hrs of Mg<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup> deficiencies. Deficits in K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, Mg<sup>2+</sup>, Fe<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup> for 24 hrs dramatically increased transpiration rate, which was modified by those deficiencies. A 48 hr exposure to  $NO_{3^{-}}$ ,  $Ca^{2+}$  and  $PO_{4^{2-}}$ deficiency dramatically increased the transpiration rate. After 48 hrs, an SO<sub>4</sub><sup>2-</sup> deficit drastically decreased the transpiration rate. The findings indicate that after a short term of exposure, it may be possible to diagnose a specific mineral shortage and determine which mineral influenced the parameters of photosynthesis in such a way that the selected parameters responded in a manner that was consistent with the duration of exposure.

#### **Keywords**

Abiotic stresses, acclimation, duration of exposure, mineral deficiency, photosynthetic parameters

# Introduction

Plant nutrition, whether in excess or deficiency, has a significant impact on plant growth and productivity. As a result, mineral nutrient research concentrated on the symptoms, signals, and target molecular levels of regulation as responses to changes in soil nutrient content. The findings mainly suggested that changes in nutrient concentrations will have a significant impact on plant production and subsequently how this will affect their agricultural application, which was described as a generalized dose-response curve (1, 2).

Furthermore, the importance of minerals in biochemical and physiological functions demonstrates the severity of their limitation (3). Photosynthetic carbon assimilation is an aspect of photosynthesis that is regulated

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by abiotic factors such as the availability of minerals. In response to macro- or microelement deficit, investigations are on the physiological status of the photosynthetic machinery such as decreased PSII photochemical efficiency and limitation of the activity of PSI depending on parameters derived from chlorophyll fluorescence in tomato and maize leaves (1). Early studies have been conducted to record how nutritional limitations alter photosynthetic characteristics in plant leaves (4 - 6). Through a variety of mechanisms, such as leaf gas exchange, chlorophyll fluorescence and photosynthetic machinery components such as chlorophyll contents, PSII photochemistry, PSI content and Rubisco carboxylation activity (2, 6 - 10). Lightindependent photosynthetic processes are the most affected physiological process when plant circumant mineral availability varies. Further research into plant leaf gas exchange studies to measure the condition of photosynthesis in response to nutritional limitations is still needed. As part of a continuing series of investigations into the impact of mineral shortages on photosynthetic regulation in L. esculentum leaves, this study looks at how gas exchange changes over time in response to specific nutritional deficits.

#### **Materials and Methods**

#### Plant material and growth conditions

In 9 cm Petri plates with 2 layers of filter paper and 6 ml of sterile, distilled water, tomato seeds (*L. esculentum* Mill.) were put. The dishes were then incubated at 25 °C in the dark. After that, the seedlings were moved to a soil mixture of peat moss, perlite and vermiculite at a ratio of 2:1:1 and allowed to grow for two weeks under controlled monitored conditions at growth chamber (14 hrs of light with 80  $\mu$  mol quanta m<sup>-2</sup> s<sup>-1</sup> at 21 °C and then 10 hrs of darkness at 20 °C and ~55% relative humidity).

Plantlets that were 14 days old were transplanted into 100 ml jars with the following nutritional solution: KNO<sub>3</sub>, 10 mM and Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O, 0.5 mM CuSO<sub>4</sub>·5H<sub>2</sub>O, 2  $\mu$ M ZnSO<sub>4</sub>·7H<sub>2</sub>O, 0.115 mM H<sub>3</sub>BO<sub>3</sub>, and 0.1 mM H<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O. 5 mM MgSO<sub>4</sub>·7H<sub>2</sub>O, 2.5 mM KH<sub>2</sub>PO<sub>4</sub>, 0.05 mM KCl, and 05.0 mM MnSO<sub>4</sub>, 0.05 mM Fe–Na<sub>2</sub>EDTA (1, 8). The implementation of a specific mineral nutrient shortage followed standard instructions (1). The deficiency of a particular mineral nutrient was created and the replacement with equivalent moles is shown in Table 1. For each deficiency of a specific mineral nutrient treatment, photosynthetic measurements were performed on 8 fully developed plant leaf samples after 24 and 48 hrs of the stress application.

#### Photosynthesis measurements

A CIRAS-3 portable photosynthesis device (PP Systems, Amsbury, MA, USA) was used to evaluate the net photosynthetic rate (*A*), stomatal conductance (*Gs*), intercellular CO<sub>2</sub> concentration (*Ci*), photosynthetic water use efficiency (*WUE*), and transpiration rate (*E*). The CIRAS-3 automatically recorded data every 5 s. On the CIRAS-3, an automatic control device was used to maintain the CO<sub>2</sub> concentration (380  $\mu$  mol mol<sup>-1</sup>), relative humidity (60%), and leaf temperature (28 °C) under a photon flux density of 100  $\mu$  mol m<sup>-2</sup>s<sup>-1</sup>.

#### Statistical analysis

For all experiments, samples were analyzed and all the assays were carried out in 3 independent experiments (n=4). The results were expressed as mean $\pm$  SD. One-way ANOVA, followed by Tukey's honest significance test, was carried out at a 95% confidence level ( $p \le 0.05$ ) to compare means of parameters and interactions that were statistically different. Statistical analysis of the data was carried out using GraphPad Prism 8.0 (GraphPad Software Inc., San Diego, CA, USA).

#### Results

#### The net photosynthetic rate ( $\mu$ mol CO<sup>2</sup> m<sup>2</sup> s<sup>-1</sup>)

The net photosynthetic rate (*A*) of *L. esculentum* Mill leaves was affected in different manners by the nutritional deficiencies that resulted from the treatments employed in this study (Fig. 1). The net photosynthetic rate was significantly increased in tomato leaves grown in cultures deficient in  $SO_4^{2^-}$ ,  $Fe^{2^+}$ ,  $NO_3^-$  and  $Ca^{2^+}$  as the duration of specific deficiency increased (48 hr) by 1.3-, 1.3-, 1.5-, and 1.4- fold respectively. Tomato leaves grown in cultures deficient in K<sup>+</sup> were significantly induced 1.3- fold early (24 hr). Tomato plants grown in cultures deficient in  $PO_4^{2^-}$  showed a higher significantly induced net photosynthetic rate of 1.5-fold (24 hr) and 1.4- fold (48 hr). Tomato plants grown in cultures deficiently reduced net photosynthetic rate 0.8- fold (24 hr) compared with control.

Table 1. Treatment of specific mineral deficiencies and substitution of one salt with another, each with the final concentration specified in the materials and methods

Deficient nutrient	Replacement	Purpose of replacement
Complete medium	All minerals are	Presents
- Mg/ MgSO <sub>4.</sub> 7H <sub>2</sub> O	NaSO <sub>4</sub>	To keep the supply of sulfur
- Ca/Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O	NaNO <sub>3</sub>	To keep the supply of nitrogen
- Fe/ Fe-Na <sub>2</sub> EDTA		
- P/ KH <sub>2</sub> PO <sub>4</sub>	KCI	To keep the supply of potassium
- K/ KH <sub>2</sub> PO <sub>4</sub>	NaNO <sub>3</sub>	To keep the osmoticum
- N/ KNO3 and Ca(NO3)2.4H2O	KCl and CaCl <sub>2</sub>	To keep the supply of potassium and calcium
- S/ MgSO <sub>4.</sub> 7H <sub>2</sub> O	MgCl <sub>2</sub>	To keep the supply of magnesium



**Fig. 1.** Net photosynthetic rate (*A*) of leaves of *L. esculentum* Mill. leaves subjected to specific mineral deficiencies compared with that of plants grown in complete nutrient solution. Data represent mean values  $\pm$  SD. Different letters denote statistically different means (Tukey's test; *P*≤0.05). Bars bearing different letters indicate significance difference.

#### Photosynthetic water use efficiency (mmol CO<sup>2</sup>/mol H<sub>2</sub>O)

The photosynthesis/transpiration relation of *L. esculentum* Mill plants during different nutritional disorders presented in Fig. 2. Photosynthetic Water Use Efficiency (WUE) was significantly induced in tomato leaves grown in cultures deficient in Mg<sup>2+</sup> and Ca<sup>2+</sup> as the duration of specific deficiency increased (48 hr) by 1.5- and 1.2- fold respectively compared with control. Tomato plants grown in cultures deficient in NO<sub>3</sub><sup>-</sup> showed the lowest significantly reduced WUE to 0.3- fold (24 hr) and 0.4- fold (48 hr) compared with control.



**Fig. 2.** Water use efficiency (WUE) of *L. esculentum* Mill. leaves subjected to specific mineral deficiencies compared with that of plants grown in complete nutrient solution. Data represent mean values  $\pm$  SD. Different letters denote statistically different means (Tukey's test; *P*≤0.05). Bars bearing different letters indicate significance difference.

### Stomatal conductance (mol CO<sup>2</sup> m<sup>2</sup> s<sup>-1</sup>)

In terms of stomatal conductance (*Gs*) tomato leaves showed a specific sensitivity toward mineral deficiency similar to that of the net photosynthetic rate behavior (Fig. 3). Tomato plants grown in cultures deficient in NO<sub>3</sub><sup>-</sup> had intrinsically more open stomata than other mineralsdeficient plants, in respect to time of exposure, which induced 3.6- fold and 4.9- fold after 24 hr and 48 hr respectively, compared to the control. While cultures deficient in PO<sub>4</sub><sup>2-</sup> showed a lower than NO<sub>3</sub><sup>-</sup>effect that significantly induced stomatal conductance reached 2.2- fold and 1.5fold after 24 hr and 48 hr respectively. Ca<sup>2+</sup> deficient tomato plants exhibit slight increases reaching 1.3- fold and 1.2-fold after 24 hr and 48 hr respectively. Tomato leaves grown in cultures deficient in  $SO_4^{2^-}$  and  $Fe^{2^+}$  induced stomatal conductance as a specific duration of exposure (24 hr) that increased by 2.9- and 3- fold respectively.



**Fig. 3.** Stomatal conductance (*Gs*) of *L. esculentum* Mill. leaves subjected to specific mineral deficiencies compared with that of plants grown in complete nutrient solution. Data represent mean values  $\pm$  SD. Different letters denote statistically different means (Tukey's test; *P*≤0.05). Bars bearing different letters indicate significance difference.

# Transpiration Rate (mmol $H_2O m^2 s^{-1}$ )

Interestingly, the transpiration rate improved significantly in plants grown in cultures deficient in K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, Mg<sup>2+</sup>, Fe<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup> reached 2- fold, 3.3- fold, 1.6- fold, 3.2fold, 3.5- fold, 1.5- fold and 3.5- fold after 24 hr of treatment respectively. Then transpiration rate dramatically decreased in plants deficient in K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, Mg<sup>2+</sup> and Fe<sup>2+</sup> after 48 hr of treatment. Plants were grown in cultures deficient in NO<sub>3</sub><sup>-</sup>, Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup> they showed significant improvement reached 3.8-, 1.3- and 1.7- fold after 48 hr of treatment respectively compared to the control as shown in Fig. 4.



**Fig. 4.** Transpiration rate (*E*) of *L. esculentum* Mill. leaves subjected to specific mineral deficiencies compared with that of plants grown in complete nutrient solution. Data represent mean values  $\pm$  SD. Different letters denote statistically different means (Tukey's test; *P*≤0.05). Bars bearing different letters indicate significance difference.

#### Internal CO<sup>2</sup> concentration (µmol mol<sup>-1</sup>)

Intercellular CO<sub>2</sub> concentration (Ci) improved significantly

in plants grown in cultures deficient in K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, Mg<sup>2+</sup>, Fe<sup>2+</sup>, NO<sub>3</sub><sup>-</sup>, Ca<sup>2+</sup> and PO<sub>4</sub><sup>2-</sup> reached 1.3- fold, 1.5- fold, 1.3- fold, 1.6- fold, 1.6- fold, 1.2- fold and 1.3- fold after 24 hr of treatment respectively. Then transpiration rate dramatically decreased significantly in plants deficient in K<sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, Mg<sup>2+</sup> reached to 0.5-foldd for Fe<sup>2+</sup> and Ca<sup>2+</sup> reached 0.7- fold and 0.8- fold after 48 hr of treatment respectively compared to the control as shown in Fig. 5.



**Fig. 5.** Internal  $CO_2$  Concentration (*Ci*) of *L*. esculentum Mill. leaves subjected to specific mineral deficiencies compared with that of plants grown in complete nutrient solution. Data represent mean values  $\pm$  SD. Different letters denote statistically different means (Tukey's test; *P*≤0.05). Bars bearing different letters indicate significance difference.

# Discussion

The metabolome and enzymatic regulation that indicate plant plasticity to deal with the specific variations in plant tissue mineral content are among the levels of responses that are involved in plants' acclimation to mineral shortages (8, 9). Additionally, depending on the type, duration, and severity of the abiotic stress, different mechanisms, such as rapid and long-term acclimation, express the sensitivity through photosynthetic performance in plants (2, 11-15).

In vivo gas exchange analysis is the main tool in diagnosing the photosynthesis performance of plants. Long-term N-deficiency treatment had wide effects on plant and leaves performance (16 - 18). Leaf senescence is accelerated by N-deficiency through protein degradation and photosynthetic pigments contents reduction (16, 19). Short-time exposure for N-deficiency did not exhibit leaf senescence characteristics of L. esculentum Mill. (8, 9). Moreover, leaf gas exchanges still exhibit significant improvement in most parameters while there was a lower reduction of WUE which may to the reduction of root surface area with diverse N-deficiency levels. For that, if the photosynthesis was affected by N- limitation is due to reduced carboxylation efficiency rather than to stomatal limitation (20 -23).

A vital nutrient for photosynthetic metabolism is potassium. On photosynthetic characteristics, the Kdeficiency stress time extension (days) had a negative impact (24 - 26). Although the physiological and biochemical functions of  $K^+$  in plants have been established, the researchers reached the conclusion that K deficiency has a negative effect on stomatal function. However, neither the early (24 hr) net photosynthetic rate nor the transpiration rate decreased in this study. Furthermore, I draw the conclusion that stomatal conductance was not one of the elements limiting  $CO_2$  absorption. Therefore, even with the *Ci* reduced, I propose that the K<sup>+</sup> internal homeostasis may control the mechanisms for carboxylation efficiency either by reducing rubisco activity or by impairing photosynthesis's photochemistry (24, 26).

Sulfur plays a key orchestral role in photosynthesis by regulating the production of antioxidants, the photosynthetic mechanism, and stomatal conductance (27). Long-term sulfur restriction causes a reduction in photosynthesis (28, 29). Photosystems were more negatively impacted by Fe deficiency (30). Most leaf gas exchange parameters significantly improved after a short-term exposure for S-deficient and Fe-deficient plants, however, there was a less dramatic decrease in WUE (24 hr). Even though A was unaffected by extending the exposure time, the other parameters were significantly reduced after 48 hrs, suggesting that S- and Fe-deficient plants behave similarly in terms of how they affect photosynthetic parameters, possibly because both are necessary components of iron-sulfur complexes in other proteins and the photosynthetic apparatus in the chloroplast (2, 30, 31).

Magnesium is vital in various physiological and biochemical plant processes as part of structures or enzymes activators. Mg-deficiency induced alterations of photosynthesis parameters that are not associated with stomatal conductance since the parameters changes do not flow in the same relationship as the negative relationship between *A* and  $C_i$  as shown in the results. These findings might be explained by Mg roles as part of Chl structures or photosynthesis and respiration enzymes activators as found previously (32 - 36).

Calcium plays many different roles in plants; enzyme activities, a second messenger, leaf senescence, cell membrane, tolerance to stress and photosynthesis (37, 38). Long-term Ca deficiency lowers photosynthetic performance due to lower stomatal conductance (18). In this study, tomato leaves photosynthetic performance was inducing after short-term Ca deficiency, which could be the alleviation of calcium deficiency symptoms increased with fractioning of calcium (39).

Phosphorus (P) is a key component of macromolecules, plays critical roles in the metabolism of macromolecules, signal transduction, photosynthesis and has a pivotal impact on stress resistance. Prolonged P deficiency lowers photosynthetic performance (40). Short-time exposure to P deficiency exhibits significant improvement in most parameters of photosynthesis. There are reports on the lowers photosynthetic performance of P-deficient plants to lower stomatal conductance (18). The P deficiency induction of these parameters in this study might indicate that the P deficiency exhibit calcium deficiency effects that increase with fractioning of phosphorus (41).

# Conclusion

During the analysis of photosynthesis, the selected parameters showed specific responses to the selected mineral. In addition, the response was developed according to the duration of exposure. As a result, it was possible to diagnose a specific mineral deficit instantly and determine which mineral had an impact on the parameters of photosynthesis. Short-term exposure to a specific mineral deficit influenced the selected parameters of photosynthesis that responded in a manner consistent with the duration of exposure. Furthermore, the study of the photosynthesis parameters changes to specific mineral deficiency needs to be investigated with different plant species and genotypes to use it as an instant diagnosis tool.

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# **Authors contributions**

KA designed the project, performed the research, supervised and discussed the results and wrote the article.

# **Compliance with ethical standards**

**Conflict of interest**: Author does not have any conflict of interests to declare.

#### Ethical issues: None.

#### References

- Osman KT. Plant nutrients and soil fertility management. In: Soils. Springer, Dordrecht. 2013;129-59. http:// dx.doi.org/10.1007/978-94-007-5663-2
- Kalaji HM, Oukarroum A, Alexandrov V, Kouzmanova M, Brestic M, Zivcak M, Goltsev V. Identification of nutrient deficiency in maize and tomato plants by *in vivo* chlorophyll a fluorescence measurements. Plant Physiology and Biochemistry. 2014;81:16-25. https://doi.org/10.1016/j.plaphy.2014.03.029
- de Bang TC, Husted S, Laursen KH, Persson DP, Schjoerring JK. The molecular-physiological functions of mineral macronutrients and their consequences for deficiency symptoms in plants. New Phytologist. 2021; 229(5): 2446-69. https:// doi.org/10.1111/nph.17074
- Longstreth DJ, Nobel PS. Nutrient influences on leaf photosynthesis: Effects of nitrogen, phosphorus and potassium for *Gossypium hirsutum* L. Plant Physiology. 1980;65(3):541-43. https://doi.org/10.1104/pp.65.3.541
- Ciompi S, Gentili E, Guidi L, Soldatini GF. The effect of nitrogen deficiency on leaf gas exchange and chlorophyll fluorescence parameters in sunflower. Plant Science. 1996;118(2):177-84. https://doi.org/10.1016/0168-9452(96)04442-1
- Lima J, Mosquim P, Da Matta F. Leaf gas exchange and chlorophyll fluorescence parameters in *Phaseolus vulgaris* as affected by nitrogen and phosphorus deficiency. Photosynthetica. 1999;37:113-21. https://doi.org/10.1023/ A:1007079215683
- Terry N. Effects of sulfur on the photosynthesis of intact leaves and isolated chloroplasts of sugar beets. Plant Physiology. 1976;57(4):477-79. https://doi.org/10.1104/pp.57.4.477

- Alsharafa KY. Mineral deficiencies influence on tomato leaves: Pigments, hydrogen peroxide and total phenolic compounds contents. Plant Omics. 2017;10:78-87. https://doi.org/ 10.21475/poj.10.02.17.pne386
- Alsharafa KY. Mineral deficiencies effect on resistance-related enzymes activities in tomato leaves. Journal of Plant Nutrition. 2018; 41(18): 2320-29. https:// doi.org/10.1080/01904167.2018.1509997
- Tränkner M, Tavakol E, Jákli B. Functioning of potassium and magnesium in photosynthesis, photosynthate translocation and photoprotection. Physiologia Plantarum. 2018;163(3):414-31. https://doi.org/10.1111/ppl.12747
- Oelze ML, Vogel MO, Alsharafa K, Kahmann U, Viehhauser A, Maurino VG, Dietz KJ. Efficient acclimation of the chloroplast antioxidant defence of *Arabidopsis thaliana* leaves in response to a 10-or 100-fold light increment and the possible involvement of retrograde signals. Journal of Experimental Botany. 2012; 63 (3):1297-1313. https://doi.org/10.1093/jxb/err356
- Alsharafa K, Vogel MO, Oelze ML, Moore M, Stingl N, König K et al. Kinetics of retrograde signalling initiation in the high light response of *Arabidopsis thaliana*. Philosophical Transactions of the Royal Society B: Biological Sciences. 2014; 369(1640): 20130424. https://doi.org/10.1098/rstb.2013.0424
- Alkhsabah IA, Alsharafa KY, Kalaji HM. Effects of abiotic factors on internal homeostasis of *Mentha spicata* leaves. Applied Ecology and Environmental Research. 2018;16:2537-64. http:// dx.doi.org/10.15666/aeer/1603\_25372564
- Kalaji HM, Rastogi A, Živčák M, Brestic M, Daszkowska-Golec A, Sitko K, Cetner MD. Prompt chlorophyll fluorescence as a tool for crop phenotyping: an example of barley landraces exposed to various abiotic stress factors. Photosynthetica. 2018;56 (3):953-61. https://doi.org/10.1007/s11099-018-0766-z
- Al-Sammarraie ON, Alsharafa KY, Al-Limoun MO, Khleifat KM, Al-Sarayreh SA, Al-Shuneigat JM, Kalaji HM. Effect of various abiotic stressors on some biochemical indices of *Lepidium sativum* plants. Scientific Reports. 2020; 10(1): 21131. https:// doi.org/10.1038/s41598-020-78330-1
- Huang ZA, Jiang DA, Yang Y, Sun JW, Jin SH. Effects of nitrogen deficiency on gas exchange, chlorophyll fluorescence and antioxidant enzymes in leaves of rice plants. Photosynthetica. 2004;42(3):357-64. https://doi.org/10.1023/ B:PHOT.0000046153.08935.4c
- Zhao D, Reddy KR, Kakani VG, Reddy, VR. Nitrogen deficiency effects on plant growth, leaf photosynthesis and hyperspectral reflectance properties of sorghum. European Journal of Agronomy. 2005; 22(4): 391-403. https://doi.org/10.1016/ j.eja.2004.06.005
- Chen CT, Lee CL, Yeh DM. Effects of nitrogen, phosphorus, potassium, calcium, or magnesium deficiency on growth and photosynthesis of *Eustoma*. HortScience. 2018; 53(6):795-98. https://doi.org/10.21273/HORTSCI12947-18
- Boussadia O, Steppe K, Zgallai H, El Hadj SB, Braham M, Lemeur R, Van Labek, MC. Effects of nitrogen deficiency on leaf photosynthesis, carbohydrate status and biomass production in two olive cultivars 'Meski' and 'Koroneiki'. Scientia Horticulturae. 2010; 123(3):336-42. https://doi.org/10.1016/ j.scienta.2009.09.023
- Zhu Y, Fan X, Hou X, Wu J, Wang T. Effect of different levels of nitrogen deficiency on switchgrass seedling growth. The Crop Journal. 2014; 2(4): 223-34. https://doi.org/10.1016/ j.cj.2014.04.005
- Sumi A, Sugata S, Yahiro I, Odawara M. Effect of fertilizer and fixed nitrogen on the water use efficiency of genge (*Astragalus sinicus* L.). Plant Production Science. 2015; 18(1):104-08. https:// doi.org/10.1626/pps.18.104

- 22. Wang X, Wang L, Shangguan Z. Leaf gas exchange and fluorescence of two winter wheat varieties in response to drought stress and nitrogen supply. PLoS One. 2016; 11(11): e0165733. https://doi.org/10.1371/journal.pone.0165733
- Alou IN. van der Laan M, Annandale JG, Steyn JM. Water and nitrogen (N) use efficiency of upland rice (*Oryza sativa* L.× *Oryza* glaberrima Steud) under varying N application rates. Nitrogen. 2020; 1(2): 151-66. https://doi.org/10.3390/nitrogen1020013
- Wang XG, Zhao XH, Jiang CJ, Li CH, Shan CONG, Di WU, Wang CY. Effects of potassium deficiency on photosynthesis and photoprotection mechanisms in soybean (*Glycine max* (L.) Merr.). Journal of Integrative Agriculture. 2015;14(5):856-63. https://doi.org/10.1016/S2095-3119(14)60848-0
- Helena Ramirez-Solet C, Magnitskiy S, Melo Martinez SE, Alvarez -Florez F, Marina Melgarejo L. Photosynthesis, biochemical activity, and leaf anatomy of tree tomato (*Solanum betaceum* Cav.) plants under potassium deficiency. Journal of Applied Botany and Food Quality. 2021; 94: 75-81. https:// doi.org/10.5073/JABFQ.2021.094.009
- Kusaka M, Kalaji HM, Mastalerczuk G, DĄBROWSKI P, Kowalczyk K. Potassium deficiency impact on the photosynthetic apparatus efficiency of radish. Photosynthetica. 2021; 59(1): 127-136. https://doi.org/10.32615/ps.2020.077
- Fatma M, Iqbal N, Gautam H, Sehar Z, Sofo A, D'Ippolito I, Khan NA. Ethylene and sulfur coordinately modulate the antioxidant system and ABA accumulation in mustard plants under salt stress. Plants. 2021;10(1):180. https://doi.org/10.3390/ plants10010180
- D'Hooghe P, Escamez S, Trouverie J, Avice JC. Sulphur limitation provokes physiological and leaf proteome changes in oilseed rape that lead to perturbation of sulphur, carbon and oxidative metabolisms. BMC Plant Biology. 2013; 13(1): 1-15. https://doi.org/10.1186/1471-2229-13-23
- Samborska IA, Kalaji HM, Sieczko L, Borucki W, Mazur R, Kouzmanova M, Goltsev V. Can just one-second measurement of chlorophyll a fluorescence be used to predict sulphur deficiency in radish (*Raphanus sativus* L. *sativus*) plants?. Current Plant Biology. 2019;19:100096. https://doi.org/10.1016/j.cpb.2018.12.002
- Roosta HR, Estaji A, Niknam F. Effect of iron, zinc and manganese shortage-induced change on photosynthetic pigments, some osmoregulators and chlorophyll fluorescence parameters in lettuce. Photosynthetica. 2018;56(2):606-15. https:// doi.org/10.1007/s11099-017-0696-1
- Ohnishi M, Furutani R, Sohtome T, Suzuki T, Wada S, Tanaka S et al. Photosynthetic parameters show specific responses to essential mineral deficiencies. Antioxidants. 2021;10(7):996. https://doi.org/10.3390/antiox10070996
- Jin XL, Ma CL, Yang LT, Chen LS. Alterations of physiology and gene expression due to long-term magnesium-deficiency differ between leaves and roots of *Citrus reticulata*. Journal of Plant Physiology. 2016;198:103-15. https://doi.org/10.1016/ j.jplph.2016.04.011

- Li CP, Qi YP, Zhang J, Yang LT, Wang DH, Ye X et al. Magnesiumdeficiency-induced alterations of gas exchange, major metabolites and key enzymes differ among roots, and lower and upper leaves of *Citrus sinensis* seedlings. Tree Physiology. 2017; 37(11): 1564-81. https://doi.org/10.1093/treephys/tpx067
- 34. Sitko K, Gieroń Ż, Szopiński M, Zieleźnik-Rusinowska P, Rusinowski S et al. Influence of short-term macronutrient deprivation in maize on photosynthetic characteristics, transpiration and pigment content. Scientific Reports. 2019;9(1):14181. https://doi.org/10.1038/s41598-019-50579-1
- Ye X, Chen XF, Deng CL, Yang LT, Lai NW, Guo JX, Chen LS. Magnesium-deficiency effects on pigments, photosynthesis and photosynthetic electron transport of leaves and nutrients of leaf blades and veins in *Citrus sinensis* seedlings. Plants. 2019; 8 (10):389. https://doi.org/10.3390/plants8100389
- Jaghdani SJ, Jahns P, Tränkner M. Mg deficiency induces photooxidative stress primarily by limiting CO<sub>2</sub> assimilation and not by limiting photosynthetic light utilization. Plant Science. 2021; 302: 110751. https://doi.org/10.1016/j.plantsci.2020.110751
- Elkelish AA, Alnusaire TS, Soliman MH, Gowayed S, Senousy HH, Fahad S. Calcium availability regulates antioxidant system, physio-biochemical activities and alleviates salinity stress mediated oxidative damage in soybean seedlings. Journal of Applied Botany and Food Quality. 2019;92:258-66. https:// doi.org/10.5073/JABFQ.2019.092.036
- Aslam S, Gul N, Mir MA, Asgher M, Al-Sulami N, Abulfaraj AA, Qari S. Role of jasmonates, calcium, and glutathione in plants to combat abiotic stresses through precise signaling cascade. Frontiers in Plant Science. 2021;1172. https:// doi.org/10.3389/fpls.2021.668029
- Gao H, Wu X, Zorrilla C, Vega SE, Palta JP. Fractionating of calcium in tuber and leaf tissues explains the calcium deficiency symptoms in potato plant overexpressing CAX1. Frontiers in Plant Science. 2020;10:1793. https://doi.org/10.3389/ fpls.2019.01793
- Meng X, Chen WW, Wang YY, Huang ZR, Ye X, Chen LS, Yang LT. Effects of phosphorus deficiency on the absorption of mineral nutrients, photosynthetic system performance and antioxidant metabolism in *Citrus grandis*. Plos One. 2021;16(2): e0246944. https://doi.org/10.1371/journal.pone.0246944
- Carstensen A, Herdean A, Schmidt SB, Sharma A, Spetea C, Pribil M, Husted S. The impacts of phosphorus deficiency on the photosynthetic electron transport chain. Plant Physiology. 2018;177(1):271-84. https://doi.org/10.1104/pp.17.01624

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