



RESEARCH ARTICLE

# Evaluation of antibacterial and antifungal activities of *Pistacia* atlantica and *Pistacia khinjuk*

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## Abstract

Medicinal plants are renowned for their various therapeutic properties, including antibacterial and antifungal activities. This study aimed to investigate the antibacterial and antifungal activities of Pistacia atlantica and Pistacia khinjuk. Hydroalcoholic extracts of P. atlantica and P. khinjuk were prepared to assess their antibacterial and antifungal activities. Standard strains of Bacillus cereus, Pseudomonas aeruginosa, Escherichia coli, Staphylococcus aureus, Klebsiella pneumonia, Aggregatibacter actinomycetemcomitans, Porphyromonas gingivalis, and Aspergillus flavus were utilized for the evaluation of antibacterial and antifungal activities. The inhibitory effects of the extracts on the growth of bacterial and fungal strains were evaluated using minimum inhibitory concentration (MIC) and minimal bactericidal concentration (MBC) through a 96-well microplate analysis following CLSI guidelines. Our findings revealed that the MICs and MBCs of P. atlantica and P. khinjuk for bacterial strains ranged from 0 to 64 mg/mL. Additionally, the MIC and MFC values for fungal strains ranged from 16 to 64 mg/ mL. The results indicated that Pseudomonas aeruginosa and Klebsiella pneumonia were the most sensitive bacterial strains to P. atlantica. Furthermore, Aspergillus flavus was identified as the most sensitive fungal strain to P. atlantica. In conclusion, these findings suggest that P. atlantica and P. khinjuk possess antibacterial and antifungal effects. The paper argues that these plant extracts could be used as a supplementary treatment alongside conventional antibacterial and antifungal drugs.

## Keywords

Antibacterial effect; antifungal activity; medicinal plants; *Pistacia atlantica*; *Pistacia khinjuk* 

# Introduction

Infectious diseases rank among the world's most prevalent health issues, imposing a substantial financial burden on society (1). Treatment of infectious diseases with existing drugs often encounters challenges such as drug resistance, prompting the need to discover new antibiotic compounds (2). With the escalating resistance of bacteria and fungi to antimicrobial agents and the high side effects associated with synthetic antimicrobial chemicals, researchers are increasingly exploring herbal extracts and natural antibacterial agents for the treatment of infectious diseases (3). Throughout history, people and traditional healers have utilized medicinal plants for disease treatment, with many ailments successfully addressed by herbs (4). Abun-

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dant evidence indicates that 70-80% of people worldwide rely on medicinal plants for primary healthcare (5). Medicinal plants offer a unique and renewable resource for discovering new therapeutically active biomolecules, thanks to the structural and biological diversity of their compounds (6). It is estimated that compounds isolated from medicinal plants contribute to 25% of commonly used drugs (7). Numerous studies have demonstrated that medicinal plants possess diverse biological properties, including antibacterial, anti-inflammatory, and antipyretic effects (8, 9). The antimicrobial properties of medicinal plants have proven effective against a wide range of microorganisms (10). Several studies argue that the presence of phytochemical constituents, such as polyphenolic compounds, is responsible for the various healing properties of medicinal plants (11, 12).

Over the years, extensive research has been conducted on the therapeutic significance of plant species in the genus Pistacia (13). It is now widely acknowledged that one of the most well-known medicinal plants of the Anacardiaceae family is Pistacia atlantica (P. atlantica). Traditionally, *P. atlantica* has been used for various purposes, including therapeutic effects on the stomach, intestine, brain, heart, lung, and kidneys (14). Numerous biological properties have been attributed to P. atlantica, encompassing antioxidative, anti-microbial, anti-inflammatory, anti-cancer, anti-diabetic, and wound-healing properties (13). Different parts of P. atlantica have been found to contain a diverse range of beneficial phytoconstituents, such as phenolic compounds, flavonoids, and tannins (13, 14). Pistacia khinjuk (P. khinjuk) is also a significant member of the genus Pistacia. In folk medicine, P. khinjuk has proven effective in treating issues such as cardiovascular disease, dental pain, dyspepsia, and cancer (15). There is substantial evidence regarding the wound healing, antibacterial (16), antioxidant (17), and antihyperlipidemic (18) effects of P. khinjuk. Detailed phytochemical analysis of P. khinjuk has revealed the presence of tannins, phenolics, and glycosides (19). To the best of our knowledge, previous studies have explored the anti-microbial and antifungal effects of the leaves of P. atlantica and P. khinjuk. However, few studies have focused on the anti-microbial and antifungal activities of the fruits of these plants. Furthermore, this study aimed to evaluate the antibacterial and antifungal activities of the fruits of P. atlantica and P. khinjuk for the first time on the pathogens studied in this research.

# **Materials and Methods**

#### **Chemicals and Reagents**

All of the required chemicals, including ethanol, Muller Hinton broth, Muller Hinton agar, and potato dextrose agar (PDA), were purchased from Merck (Germany) Company.

## **Collection of Plant Materials**

In May 2020, fruits of *P. atlantica* and *P. khinjuk* were collected from the Khargūshān Mountains at the end of the Khorramabad-Andimeshk freeway and the Bisim Moun-

tains in Khorramabad County, Lorestan Province, Iran. Subsequently, the plant samples were identified by botanists from the Lorestan Agriculture and Natural Resources Research Center in Khorramabad, Iran.

#### **Preparation of Extract**

Fresh fruits of the plants were collected and air-dried under shaded conditions for ten days at room temperature to prepare hydroalcoholic extracts of *P. atlantica* and *P. khinjuk*. After drying, the fruits were pulverized, and hydroalcoholic extracts were obtained using the maceration method. The resulting powder was dissolved in a hydroalcoholic mixture (50% distilled water and 50% ethanol) in a ratio of 1:3 (w/v) at 70°C. Hydroalcoholic solutions, combining alcohol and water, are known for their enhanced efficacy in extracting medicinal substances from herbs. The suspension was maintained at 30 °C for 72 hours and later filtered through filter paper. The solvent was then removed using a rotary evaporator (IKA-RV10, Germany) (20).

# **Bacterial Strains**

The bacterial strains, including *Bacillus cereus* (ATCC: 14579), *Pseudomonas aeruginosa* (ATCC: 27853), *Escherichia coli, Staphylococcus aureus* (ATCC: 12600), and *Klebsiella pneumonia* (ATCC: 700603), were received from the National Cell Bank of Iran (NCBI, Pasteur Institute, Tehran).

# **Fungal Strains**

The standard strain of *Aggregatibacter actinomycetemcomitans*, *Porphyromonas gingivalis*, and *Aspergillus flavus* were received from the National Cell Bank of Iran (NCBI, Pasteur Institute, Tehran).

#### **Determination of Antibacterial Activity**

To assess antibacterial properties, we determined the Minimal Inhibitory Concentration (MIC) and Minimal Bactericidal Concentration (MBC) using the microbial broth dilution method with 96-well plates, following the Clinical and Laboratory Standards Institute (CLSI) guidelines. The stocks of P. atlantica and P. khinjuk extracts were prepared in Muller Hinton broth. Initially, 50 mL of sterile Muller Hinton broth was added to each well. Serial dilutions were then created by adding 100 mL of the stock, resulting in concentrations ranging from 50 mg/mL to 0.097 mg/mL. Each well's extract concentration was half that of the previous well. Subsequently, 50 mL of a 24-hour microbial culture (equivalent to 1.5 × 105 CFU/mL) was introduced to each well. After shaking, the plates were incubated at 37°C for 24 hours. Post-incubation, the wells were examined for turbidity, indicating bacterial growth. The MIC was identified as wells without turbidity. For MBC determination, 5 mL from clear wells was plated on Muller Hinton Agar medium and observed for microbial growth after 24 hours. The MBC was considered the well with the lowest extract concentration where no bacterial growth occurred on the corresponding plate. The control contained diluted extract plus medium, while the negative control included microbial suspension plus medium. Tetrazolium salt was added to each well, and the MIC was determined by the color change to red-pink. Each test was conducted in triplicate (21).

#### **Determination of Antifungal Activity**

To assess the antifungal activity of P. atlantica and P. khinjuk extracts, the broth microdilution method was employed to determine the Minimum Inhibitory Concentration (MIC) and Minimum Fungicidal Concentration (MFC) in accordance with CLSI guidelines. In the initial step, a 96well microplate was prepared, with each well receiving 100 µL of Potato Dextrose Agar (PDA) medium. To observe the antifungal effects of extracts, two-fold serial dilutions ranging from 50 mg/ml to 0.097 mg/ml were meticulously prepared in the microplates. Afterward, the concentrations of P. atlantica and P. khinjuk extracts were added to separate rows in the 96-well plate. Additional rows were assigned for control (PDA medium + fungi suspension), negative control (PDA medium + extracts), and blank (distilled water). A fungus suspension (1.5 × 105 CFU/mL) was prepared in 100 µL of PDA medium and added to the wells, excluding those designated for the blank and negative control. Subsequently, the microplates were incubated at 35 °C for 48 to 72 hours. All experiments were conducted in triplicate. Following the incubation period, careful scrutiny of the microplates was performed to identify turbidity due to fungal growth. The last dilution of extracts without turbidity in each well was determined as the MIC.For MFC determination, 20 µL from the pre-MIC wells was cultured in PDA medium and incubated at 35°C for 24 hours (22).

# **Statistical Analysis**

All values were presented as the mean ± standard deviation (SD). Data analysis was conducted using SPSS version 22.0 and Microsoft Office Excel 2019. Each assay was repeated three times for the samples, and the results were averaged. ANOVA and t-test methods were employed to compare the means of two groups. All P values < 0.05 were considered statistically significant.

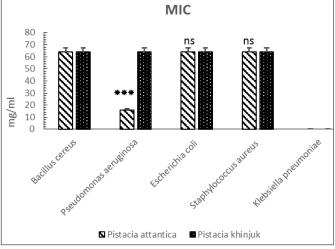
# Results

# Antibacterial Activity

The antibacterial effects of the hydro-alcoholic extracts of P. atlantica and P. khinjuk were illustrated in Fig. 1 and 2. The results of the antibacterial tests showed that the hydro-alcoholic extracts of P. atlantica and P. khinjuk had antibacterial effects. Based on the data in Fig. 1, the MIC values of the hydro-alcoholic extract of P. atlantica were obtained as 64, 16, 64, 64, and 0 mg/ml for *Bacillus cereus*, Pseudomonas aeruginosa, Escherichia coli, Staphylococcus aureus, and Klebsiella pneumonia, respectively. Furthermore, the MIC values of the hydro-alcoholic extract of P. khinjuk were found to be 64, 64, 64, 64, and 0 mg/mL for Bacillus cereus, Pseudomonas aeruginosa, Escherichia coli, Staphylococcus aureus, and Klebsiella pneumonia, respectively (Fig. 1). Additionally, the MIC values of the hydroalcoholic extract of P. atlantica were significantly lower than the MIC values of the hydro-alcoholic extracts of P. khinjuk for Pseudomonas aeruginosa (p < 0.0001). However, there was no significant change in the MIC values of the hydro-alcoholic extract of P. atlantica and P. khinjuk for

Bacillus cereus, Escherichia coli, Staphylococcus aureus, and Klebsiella pneumonia.

Looking at Fig. 2, it is apparent that the MBC values of the hydro-alcoholic extract of *P. atlantica* were obtained as 64, 16, 64, 64, and 0 mg/ml for *Bacillus cereus*, *Pseudo*-



**Fig. 1.** Determined MICs of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for selected bacterial strains. **\*\*\***Significant change at p < 0.0001. **ns**: Non-significant change.

monas aeruginosa, Escherichia coli, Staphylococcus aureus, and Klebsiella pneumonia, respectively. In addition, the MBC values of the hydro-alcoholic extract of *P. khinjuk* were found to be 64, 64, 64, 64, and 0 mg/ml for Bacillus cereus, Pseudomonas aeruginosa, Escherichia coli, Staphylococcus aureus, and Klebsiella pneumonia, respectively (Fig. 2). Furthermore, the MBC values of the hydroalcoholic extract of *P. atlantica* were significantly lower than the MBC values of the hydro-alcoholic extract of *P. khinjuk* for Pseudomonas aeruginosa (p < 0.0001). However, there was no significant change in the MBC values of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for Bacillus cereus, Escherichia coli, Staphylococcus aureus, and Klebsiella pneumonia.

#### Antifungal Activity

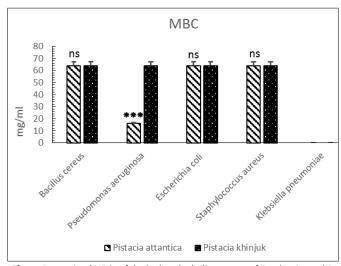
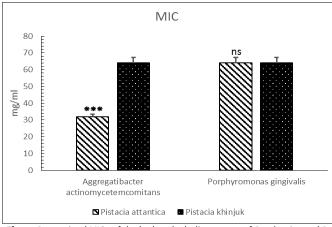
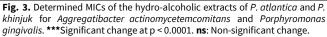


Fig. 2. Determined MBCs of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for selected bacterial strains. **\*\*\***Significant change at p < 0.0001. **ns**: Non-significant change.

The results, as seen in Fig. 3, indicate that the hydroalcoholic extract of *P. atlantica* exhibited potent antifungal activity against *Aggregatibacter actinomycetemcomitans* (MIC value = 32 mg/ml) and *Porphyromonas gingivalis* (MIC value = 64 mg/ml). Moreover, our results revealed an antifungal effect of the hydro-alcoholic extract of *P. khinjuk* against *Aggregatibacter actinomycetemcomitans* (MIC value = 64 mg/ml) and *Porphyromonas gingivalis* (MIC value = 64 mg/ml) and *Porphyromonas gingivalis* (MIC value = 64 mg/ml) (Fig. 3). The results showed a significant difference in the MIC value of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for *Aggregatibacter actinomycetemcomitans* (p < 0.0001). However, a non-significant difference was observed in the MIC value of the hydroalcoholic extracts of *P. atlantica* and *P. khinjuk* for *Porphyromonas gingivalis*.

We found that the MFC value of the hydro-alcoholic





extract of *P. atlantica* was obtained at 64 mg/ml for *Aggregatibacter actinomycetemcomitans* and *Porphyromonas gingivalis*. Furthermore, the MFC value of the hydroalcoholic extract of *P. khinjuk* was found to be 64 mg/ml for *Aggregatibacter actinomycetemcomitans* and *Porphyromonas gingivalis* (Fig. 4). We also found that there was no significant difference between the MFC values of the hydroalcoholic extracts of *P. atlantica* and *P. khinjuk* for *Aggregatibacter actinomycetemcomitans* and *Porphyromonas gingivalis*.

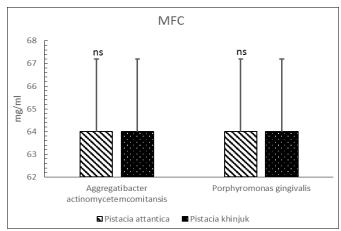
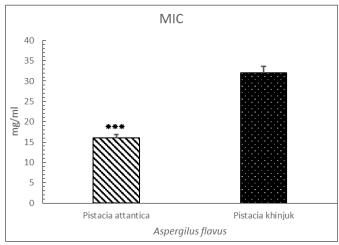


Fig. 4. Determined MFCs of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for *Aggregatibacter actinomycetemcomitans* and *Porphyromonas gingivalis*. ns: Non-significant change.

As shown in Fig. 5, our finding determined the MIC values of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* as 16 and 32 mg/ml for *Aspergillus flavus*, respectively. Furthermore, our results indicated that the MIC value of the hydro-alcoholic extract of *P. atlantica* was significantly lower than the MIC value of the hydro-alcoholic extract of *P. khinjuk*.



**Fig. 5.** Determined MICs of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for *Aspergillus flavus*. **\*\*\***Significant change at p < 0.0001. **ns**: Non-significant change.

It is evident from the data in Fig. 6 that the MFC value of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* was 32 mg/ml for *Aspergillus flavus*. Furthermore, it has been observed that there was no significant difference between the MFC values of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for *Aspergillus flavus*.

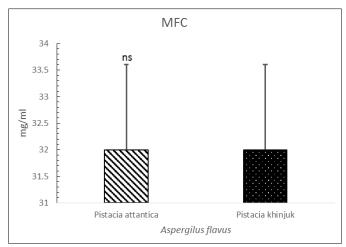


Fig. 6. Determined MFCs of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* for *Aspergillus flavus*. **ns**: Non-significant change.

# Discussion

Several reports have shown the antibacterial and antifungal activities of *P. atlantica* and *P. khinjuk*. The present study was designed to determine the antibacterial and antifungal effects of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk* against selected bacterial and fungal strains. The current study found that the hydro-alcoholic extract of *P. atlantica* possesses antibacterial property. Our findings revealed that selected bacterial strains, including *Bacillus cereus*, *Pseudomonas aeruginosa*, *Escherichia coli*, and *Staphylococcus aureus*, displayed sensitivity

to the extract of *P. atlantica*. Similar finding was reported by Rigane and colleagues (2016). They observed a similar antibacterial effect on Gram-positive and Gram-negative bacteria post-treatment with ethanolic and aqueous extracts of *P. atlantica* (23). Comparison of our findings with those of other studies confirms the antibacterial effect of P. atlantica (24, 25). This result may be attributed to the high content of phytochemical constituents responsible for the antibacterial effect of P. atlantica. It has been established that chemical compounds, such as  $\alpha$ -pinene and D-limonene, are present in abundance in *P. atlantica* (26). There is ample evidence regarding the antibacterial role of these compounds (27). The results of our study indicated an antibacterial effect of the hydro-alcoholic extract of P. khinjuk. Our findings in the present study are consistent with those of Azadpour and colleagues (2015), who reported the antibacterial effect of the methanolic extract of P. khinjuk against both Gram-positive and Gram-negative bacteria (28). Our experiments align with the results of a previous study conducted by Taghizadeh et al. They documented the inhibitory effect of P. khinjuk essential oil on bacterial strains. (15). There are several possible explanations for this result. For example, P. khinjuk is a rich source of phytoconstituents such as  $\alpha$ -pinene,  $\beta$ - pinene and Dlimonene which have a wide range of biological capacities, including antibacterial effects (27, 29). In our study, the antifungal analysis demonstrated the inhibitory effect of the hydro-alcoholic extract of *P. atlantica* on the growth of fungal strains. A considerable amount of literature has been published on the antifungal effects of *P. atlantica*. For instance, Falahati and colleagues showed that the methanolic extract of P. atlantica was effective against Candida strains (30). Many organisms live in and on our body; they are usually harmless or even useful, but some of them may cause disease under certain conditions (31-35). For the treatment of infectious diseases, medicinal plants and natural substances can be a suitable choice (36-38) because they are rich in secondary medicinal compounds such as phenols, flavonoids, anthocyanins, tannins, and plant antioxidants (39-42). The results of this study are consistent with the findings of our study. Our findings broadly support the work of other studies in this area. In a similar study conducted by Shialy et al., the ethanolic extract of P. atlantica possessed antifungal effects against fungal strains, including Aspergillus flavus, with a MIC value ranging from 6.25 to 25 mg/ml (43). The findings of this study are consistent with the results of our research. P. atlantica is recognized as a medicinal plant with a variety of pharmacological effects (20). Indeed, numerous types of biologically active compounds have been identified in P. atlantica, which are attributed to its therapeutic effects, including antifungal ones (44). In another part of our study, we identified the antifungal effect of P. khinjuk against fungal strains, including Aspergillus flavus. This result aligns with previous findings in the literature. For example, our results support the findings of Tahvilian *et al.*, who reported the antifungal activity of the essential oil of P. khinjuk against Candida albicans. They demonstrated a potent antifungal effect of P. khinjuk on Candida albicans (45). P.

khinjuk is a medicinal herb with several therapeutic effects, including antifungal properties (28). It has been observed that there is a relationship between the presence of phytochemical constituents in *P. khinjuk* and the biological properties of this plant (15). Indeed, the results of the current study revealed that bacterial strains, especially Pseudomonas aeruginosa, were more sensitive to P. atlantica than P. khinjuk due to the lower MIC value. In addition, fungal strains, including Aspergillus flavus and Aggregatibacter actinomycetemcomitans, showed a lower MIC value to P. atlantica compared to P. khinjuk and were subsequently more sensitive to P. atlantica than P. khinjuk. Bioactive components of *P. atlantica* including *a*-pinene and  $\beta$ -pinene, may be attributed to this antimicrobial effect (16). These findings suggest the therapeutic potential of P. atlantica and P. khinjuk in the pharmaceutical industry. Traditional herbalists use plants to treat diseases caused by microbial contamination (46-49). The problem of antibiotic resistance has made it necessary to try to identify effective antibacterial compounds against pathogenic bacteria resistant to existing antibacterial compounds (50-52).

# Conclusion

The present study aimed to investigate the antibacterial and antifungal activities of *P. atlantica* and *P. khinjuk*. The findings clearly indicate the antibacterial and antifungal effects of the hydro-alcoholic extracts of *P. atlantica* and *P. khinjuk*. Taken together, these results suggest that *P. atlantica* and *P. khinjuk* could be considered as alternative antifungal agents to inhibit the growth of bacterial and fungal strains. A future study exploring the other biological properties of these plants would be very interesting.

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# **Authors contributions**

All authors contributed equally in the preparation well as revision of the manuscript and approved the final version.

# **Compliance with ethical standards**

**Conflict of interest**: No conflict of interest was stated by the author(s).

Ethical issues: None.

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