



RESEARCH ARTICLE

Effects of acute gamma irradiation on the morphology of *Curcuma alismatifolia* ‘Siam Shadow’ and *C. alismatifolia* ‘Siam Scarlet’

Vachiraporn Pikulthong¹, Sirichanya Inboon¹, Santi Ariya², Narumon Boonman¹, Chanate Wanna¹, Piyada Wongwiwat¹ & Sirirat Phakpaknam^{1*}

¹Department of Science, Faculty of Science and Technology, Suan Sunandha Rajabhat University, Bangkok 10300, Thailand

²National Center for Genetic Engineering and Biotechnology, National Science and Technology Development Agency. Thailand Science Park, Khlong Luang, Pathum Thani 12120, Thailand

*Email: sirirat.ph@ssru.ac.th



ARTICLE HISTORY

Received: 28 October 2023

Accepted: 29 June 2024

Available online

Version 1.0 : 23 October 2024



Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>)

CITE THIS ARTICLE

Pikulthong V, Inboon S, Ariya S, Boonman N, Wanna C, Wongwiwat P, Phakpaknam S. Effects of acute gamma irradiation on the morphology of *Curcuma alismatifolia* ‘Siam Shadow’ and *C. alismatifolia* ‘Siam Scarlet’. Plant Science Today (Early Access). <https://doi.org/10.14719/pst.3049>

Abstract

Curcuma alismatifolia (Pathumma in Thailand), also known as Siam Tulip, is an economically valuable cut flower in Thailand and is continuously bred to enhance its captivating array of colors. This study aimed to investigate the effects of acute gamma irradiation on 2 cultivars of *C. alismatifolia* ‘Siam Shadow’ and *C. alismatifolia* ‘Siam Scarlet’ for *in vitro* propagation and determination of the lethal dose 50 (LD₅₀). The irradiation was conducted at doses of 0, 20, 30, 40 and 80 Gy using young shoots tissue culture. The results of the experiment revealed that the LD₅₀ for ‘Siam Shadow’ could not be determined while ‘Siam Scarlet’ was 29.06 Gy. ‘Siam Shadow’ exhibited chimera leaf formation when exposed to a gamma irradiated dose of 40 Gy, while ‘Siam Scarlet’ exhibited chimera leaf formation when gamma irradiated at doses of 20 Gy *in vitro*. After planting in a greenhouse, ‘Siam Shadow’ was irradiated with 30 Gy of gamma irradiated, had reduced bract numbers and the bracts became smaller. Conversely, ‘Siam Scarlet’ irradiated with 20 Gy of gamma irradiated, produced smaller inflorescences with only 2 bracts, while irradiation with 30 Gy resulted in 2 lobes and spotted leaves at the edges of the leaves. The results indicated that both the *C. alismatifolia* cultivars and gamma irradiation rates had a significant influence on survival rate, number of shoots, plant height, chlorophyll mutants and morphology. As the outcome, it was determined that gamma irradiation with optimum doses of gamma rays could induce new characteristics in *C. alismatifolia*.

Keywords

Acute gamma rays; *Curcuma alismatifolia* ‘Siam Shadow’; *C. alismatifolia* ‘Siam Scarlet’; LD₅₀; Mutation

Introduction

Curcuma alismatifolia Gagnep. is a genus of *Curcuma* belonging to the family Zingiberaceae with a rhizome as an underground stem. This plant thrives and blooms particularly well during the rainy season. Internationally, *C. alismatifolia* is often referred to as Siam Tulip because of its beautiful flowers similar to European tulips. Thailand exports Pathumma (*C. alismatifolia*) with a value range from 3-6 hundred thousand dollars in various forms, including undeveloped rhizomes, growing rhizomes and fresh flowers (1). Because *C. alismatifolia* has unique flowers and a variety of colors, it is used for cut flowers, potted plants and ornamental flowering plants, making *C. alismatifolia* interesting for

many foreign countries. Thailand is an important source of the genetics and diversity of *C. alismatifolia* plants and it is found in all regions throughout the country (1, 2).

C. alismatifolia is Thailand's second-most important economic flowering plant after Orchids. It is an economic plant contained in the National Economic and Social Development Plan for exports (3). According to the producers and developers of KP Holland, *C. alismatifolia* has been exported to the Netherlands market and is worth more than 4 million euros (4). According to the report to date, the trend of production for flowering and ornamental plants in the global market is expected to include products such as flowering plants and exotic local plant species (typical local flowers and plants). Consequently, there is continuous breeding of ornamental plants to meet market demands. As a result, the breeding of ornamental plants is taking place continuously to respond to the demands of the market. The breeding of *C. alismatifolia* takes a long time because *C. alismatifolia* has a dormancy of 2-3 years. The resulting hybrids are often sterile, which is a natural mechanism and they cannot be used for further breeding. As a result, cross-breeding has not been successful (5) and the main obstacle to producing *C. alismatifolia* rhizomes at the export level is the shortage of disease-free *C. alismatifolia* rhizomes. *C. alismatifolia* rhizomes stored for planting in the next season may be inferior in quality or have infectious diseases, which will adversely affect farmers' plots and influence exports (6). Artificial crossbreeding has produced many *Curcuma* variants with superior growth properties and bract color. Inter-specific hybridization faces a significant challenge because of their low fertility, which prevents them from serving as parents for the following stages of reproduction (7). It was reported that the hybrids with near parent relationships had much higher fruit setting rates than those with distant parent relationships (8).

The impact of gamma irradiation on the germination and growth attributes of economically important plants demonstrates the potential of this technique to expedite the breeding process and facilitate the cultivation of plant varieties possessing desired characteristics (9). There were *C. alismatifolia* breeding and improvement by gamma irradiation to find a lethal dose of 50 (LD₅₀). Studies are on the result of gamma irradiation on *C. alismatifolia* hybrid 'Laddawan' in the tissue culture and found that the optimal gamma irradiating doses ranged from 30-60 Gy (1). For *C. alismatifolia* cultivars 'Pink' an irradiation dose of approximately 25 Gy was applied (10); LD₅₀ values of the cultivars were achieved at 21 Gy for 'Chiang Mai Red', 23 Gy for 'Sweet Pink', 25 Gy for 'Kimono Pink', and 28 Gy for 'Doi Tung 554' (11).

Propagation through plant tissue culture techniques enables the rapid generation of large quantities of genetic material within a short period. This method ensures disease-free propagation, reduces planting duration and eliminates the dependency on seasonal factors (12). Therefore, this research examined *C. alismatifolia* breeding by using tissue culture techniques from young shoots and the seedlings were irradiated with gamma-rays, which is the process of inducing genetic mutations in plants and finding the appropriate amount of gamma irradiation to

induce new characteristics. The production of varieties is produced *in vitro* (clean culture) so it is convenient for storage and can be produced out of season, which will increase income for farmers who grow *C. alismatifolia* in the future.

Materials and Methods

Plant materials

Plant samples used in this experiment were 2 cultivars of *C. alismatifolia* 'Siam Shadow' and 'Siam Scarlet' were obtained from *Curcuma* Farmers in San Sai district, Chiang Mai, Thailand (13, 14). These samples were cultured *in vitro* at the National Center for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency (NSTDA), Thailand.

Rhizome sterilization

The buds from the rhizomes of *C. alismatifolia* (adapted from the patent code IPCA01H4/00, C07K) (6) were cut and then disinfected with 70 % ethanol (v/v) for 1 min before discarding the solution. Next, bleach disinfection was performed using a solution containing 20 % (v/v) sodium hypochlorite along with 2 drops of Tween 20, which was shaken at 150 rpm for 20 min and then discarded. The explant was placed on MS medium (15), 30 g sucrose, 2.75 g Kelcogel®, adjusted pH to 5.6 supplemented with Thidiazuron (TDZ) at a concentration of 1 mg/L. The explant was cultured at 25 ± 2 °C with a light intensity of 65 µmol/m²/s⁻¹ for 16 h/day using fluorescent white light.

After culturing for 4 weeks, the young shoots free from microbial contamination were selected and cut in half length-wise. These shoots were cultured on MS medium with plant growth regulators BA at a concentration of 3 mg/L for an additional 4 weeks at 25 ± 2 °C with a light intensity of 65 µmol/m²/s⁻¹ for 16 h/day using fluorescent white light.

LD₅₀: The impact of gamma irradiation on tissue death

Seedlings of 'Siam Shadow' and 'Siam Scarlet' were irradiated with gamma rays at rates of 0, 20, 30, 40 and 80 Gy. The gamma irradiation source was cobalt-60 and consisted of 6 rods provided by Paul Stephens Consultancy Ltd. (at a dose rate of 75 Gy/h, JS 8900 IR-155 at the Thailand Institute of Nuclear Technology (Public Organization) in Ongkharak District, Nakhon Nayok Province, Thailand. Subsequently, the seedlings were placed on MS medium supplemented with BA 3 mg/L cultured at 25 ± 2 °C under 50 % light intensity for 1 week. After irradiation, the number of surviving plantlets was recorded to calculate the LD₅₀, after culturing for 4 weeks. The results of morphological changes were recorded.

Survival rate and mutation after transplanting seedling

The temperature in the tissue culture bottles was adjusted to room temperature for a day. The seedling was placed in a container with planting materials including coconut coir, husk, black husk ashes and potting soil at a ratio of 3:4:2:1. The planting materials were filled to ¾ of its volume to cultivate the seedling in the greenhouse.

Statistical analysis

Statistical analysis was performed using SPSS version 24.0, with a confidence level of 95 %. The experimental design was a factor in a completely randomized design (Factorial in CRD) with 2 factors: factor A: cultivars of *C. alismatifolia* ('Siam Shadow' and 'Siam Scarlet'), factor B: gamma irradiation dose on the number of shoots after irradiation, height of pseudo-stems and survival rate after planting with 3 replications for each treatment, 25 plantlets per replication. The paired difference mean was then compared by Scheffe's test and determined LD₅₀ by Probit analysis.

Results and Discussion

Effects of gamma irradiation on the LD₅₀ and morphology *in vitro*

The buds from the rhizomes of 'Siam Shadow' and 'Siam Scarlet', which were disinfected had survival rates of 92.86 % and 85.72 % respectively, with one shoot/explant and a plant height of 2-3 cm. The explant, which had undergone gamma irradiation for 4 weeks, showed normal growth. Notably, gamma irradiated 'Siam Shadow' seedlings had a higher survival rate than 'Siam Scarlet' seedlings under the same dose of gamma irradiation, with no microbial contamination detected (Fig. 1).

Seedling of 'Siam Shadow', the control and irradiat-

Table 1. Survival rate and number of shoots of 'Siam Shadow' and 'Siam Scarlet' seedlings after gamma irradiation *in vitro*.

Dose rate (Gy)	Survival rate (%)		No. of shoots/explant	
	Siam Shadow	Siam Scarlet	Siam Shadow	Siam Scarlet
Control	100 ± 0.00 ^d	100 ± 0.00 ^d	2.52 ± 0.15 ^e	1.48 ± 0.19 ^d
20	100 ± 0.00 ^d	88 ± 4.90 ^c	2.08 ± 0.10 ^e	1.20 ± 0.06 ^{cd}
30	100 ± 0.00 ^d	64 ± 7.48 ^b	1.44 ± 0.32 ^d	0.88 ± 0.10 ^b
40	100 ± 0.00 ^d	0 ± 0.00 ^a	1.12 ± 0.10 ^{bc}	0.00 ± 0.00 ^a
80	64 ± 4.00 ^b	0 ± 0.00 ^a	0.95 ± 0.30 ^b	0.00 ± 0.00 ^a
cultivars (A)		*		*
dose (B)		*		*
A x B		*		*

* There was a statistically significant difference at the 95 % confidence level. The mean ± standard deviation followed by different superscripts within the same column was statistically different by Scheffe's test.

under the same irradiation level because of radiosensitivity differences. As a result, the amount of gamma irradiation applied to each plant is not the same. The same species, but different varieties, exhibit sensitivity to different irradiation doses (16, 17). It was reported that low-dose impacts caused physiological, biochemical and molecular changes, the majority of which were positive (18). Low gamma-ray levels may or may not have the same effects on seedling growth, but they also increase germination and seedling

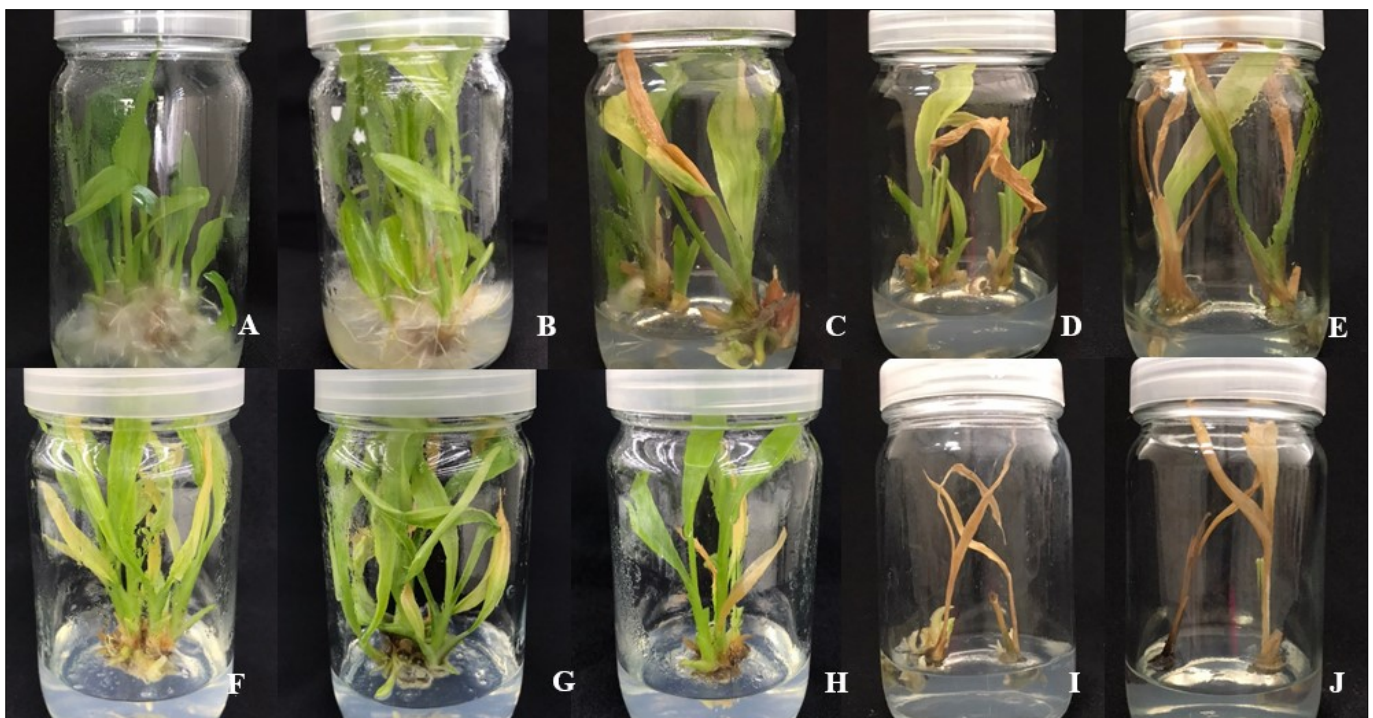


Fig. 1. Growth of 'Siam Shadow' and 'Siam Scarlet' seedlings after gamma irradiation on MS medium supplemented with BA 3 mg/L for 4 weeks. (A-E) Seedling of 'Siam Shadow': Control, 20, 30, 40 and 80 Gy and (F-J) Seedling of 'Siam Scarlet': Control, 20, 30, 40 and 80 Gy.

ed by gamma rays at doses of 20, 30, 40 and 80 Gy exhibited survival rates of 100 %, 100 %, 100 %, 100 % and 64 % respectively. Because the survival values were greater than 50 %, LD₅₀ could not be determined. On the other hand, seedlings of 'Siam Scarlet', including the control and irradiated by gamma rays at doses of 20, 30, 40 and 80 Gy, displayed survival rates of 100 %, 88 %, 64 %, 0 % and 0 % respectively (Table 1). With Probit analysis, LD₅₀ was determined to be 29.06 Gy. The seedling of 'Siam Shadow' had a higher survival rate than the seedling of 'Siam Scarlet'

growth. Gamma-rays, which act as ionizing radiation, can cause cytological, biochemical, physiological and morphological changes within cells and tissues, principally through the production of free radicals. These changes have an impact on how plants grow and develop. On the other hand, they can have stimulatory effects at lower doses (19).

The chances of survival by *C. alismatifolia* are lower as the amount of gamma irradiation increases because

gamma irradiation has a penetrating effect on the object. As the dose increases, the destructive energy is high, resulting in the death of plant tissues such as *Lindernia* spp. (20) *Narcissus tazetta* (21) *Cullen corylifolium* (22) *Curcuma* hybrid 'Laddawan' (23). When plants are exposed to gamma irradiation, it results in chromosomal abnormalities and may lead to a reduction or a halt of cell division (2). High doses of gamma irradiation can completely inhibit the growth of the apical meristem, causing a cessation of cell expansion (24). Growth reductions can result from auxin degradation, auxin production inhibition, assimilatory mechanism failure and changes in the particular activity of enzymes.

The number of shoots for 'Siam Shadow' and 'Siam Scarlet' seedlings on MS medium supplemented with BA 3 mg/L indicated that the control of 'Siam Shadow' and 'Siam Scarlet' had average amounts of 2.52 ± 0.15 and 1.48 ± 0.19 shoots/explant respectively. The seedling of 'Siam Scarlet' at a dose of 40 Gy gamma irradiation had the minimum number of shoots at 0.88 ± 0.10 shoots/explant, while the seedling of 'Siam Scarlet' at a dose of 40 and 80 Gy gamma irradiation had no shoot because the seedling died (Table 1). Following the research conducted the irradiation was made on *C. hybrid* 'Laddawan' seedlings with gamma rays, it was found that the number of shoots decreased with an increase in gamma irradiation dose (1). This finding contrasts with another report (25), who reported that the irradiation doses of 20 and 40 Gy could stimulate tillering of *Globba williamsiana* more than the control and decreased as the dose of irradiation increased. Appropriate doses of irradiation reduce the level of auxin hormone, inhibit the growth of the crest, and promote the growth of the lateral bud (26). The quantities of gamma radiation administered and the specific plant types employed greatly influence both the number of viable plants and several aspects of plant growth (27). According to one report, increases in root and shoot lengths were seen in plants after seeds were exposed to low levels of gamma radiation (28). This growth stimulation is caused by the transcriptional regulation of genes associated with phytohormones, the antioxidant system, late embryogenesis abundant proteins, and cell wall components. However, cultivars of *C. alismatifolia* ('Siam Shadow' and 'Siam Scarlet') and the dose of gamma irradiation have a mutual influence on the renewal of *C. alismatifolia* with a statisti-

cally significant difference at the 95 % confidence level.

In vitro, notable changes in leaf appearance were observed in 'Siam Shadow' at an irradiated dose of 30 Gy. The leaves were light green in color, which indicates an unstable appearance (Fig. 2). This characteristic was not found in the new shoots that emerged. At an irradiated dose of 80 Gy, 'Siam Shadow' had a pale-white characteristics and the white plant was eventually found dead. Also, the characteristics of chimera leaves were observed in the 'Siam Shadow' irradiated by 40 Gy of gamma-ray and 'Siam Scarlet' irradiated by 20 Gy (Fig. 2) which was similar to *Curcuma* hybrid (23) *C. hybrid* 'Laddawan' (1). The irradiated plants exhibited lower chlorophyll content compared to the control, with a further decrease in chlorophyll content as the irradiation dose increased. These chimeric leaves displayed irregular changes. It is important to note that most somatic mutations are not inheritable, meaning that the next generation will no longer exhibit these variations (10). It was reported that gamma-rays interact with atoms or molecules to create free radicals in living cells (29). Depending on the intensity of irradiation, these radicals have been shown to differentially influence the morphology, anatomy, biochemistry and physiology of plants. They can harm or modify critical plant cell components. Among these consequences are modifications to the metabolism and cellular structure of plants.

Effects of gamma irradiation on survival rate and morphology in vivo

Before planting the seedlings of 'Siam Shadow' and 'Siam Scarlet' after gamma irradiation in the greenhouse, they were acclimated to room temperature for one day. After that, agar gel of the tissue culture medium was removed and the seedlings were soaked in an antifungal solution for 1 min. Following this, the seedlings were placed in a container with planting materials including coconut coir, husk, black husk ashes and potting soil at a ratio of 3:4:2:1.

After planting in a greenhouse for 4 weeks, the survival rates of 'Siam Shadow' and 'Siam Scarlet' seedlings were observed except for seedlings of 'Siam Scarlet' that died after gamma irradiation at 40 and 80 Gy. The survival rate of seedlings of 'Siam Shadow' on control, 20, 30 and 40 Gy were found at 100 %, 100 %, 88 % and 84 % respectively. No survival of seedling of 'Siam Shadow' when it was irradiat-

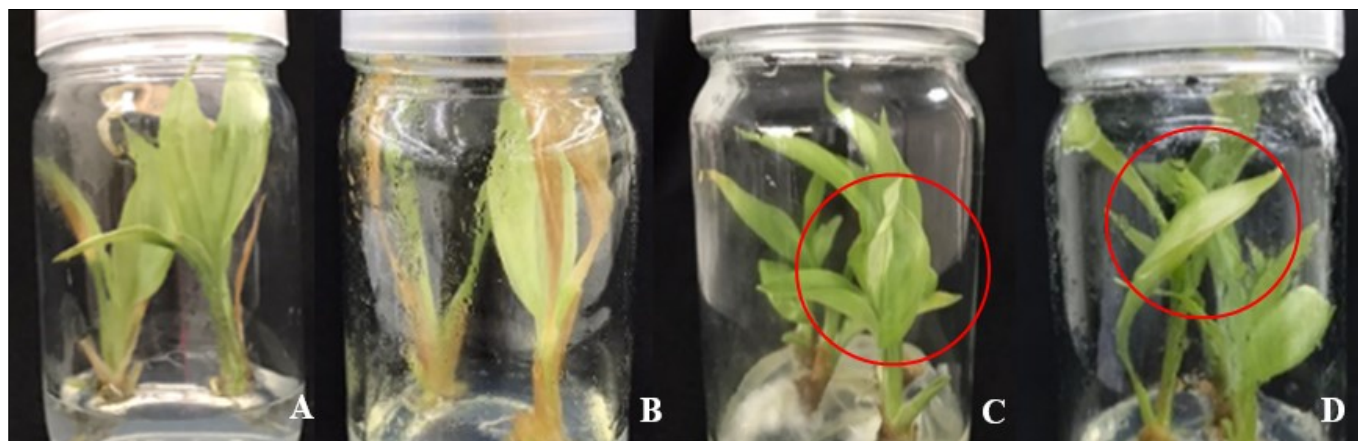


Fig. 2. Seedlings of 'Siam Shadow' and 'Siam Scarlet' after gamma irradiation on MS medium supplemented with BA 3 mg/L for 4 weeks. (A-B) Leaf characteristics of 'Siam Shadow' at doses of 30 and 80 Gy, (C) Leaf characteristics of 'Siam Shadow' at doses of 40 Gy and (D) Leaf characteristics of 'Siam Scarlet' at dose of 20 Gy.

ed at 80 Gy. On the other hand, the survival rate of seedlings of 'Siam Scarlet' in control, 20 Gy and 30 Gy was found at 100 %, 100 % and 62.50 % respectively. These findings align with a study on *Catharanthus roseus* cv. Mediterranean Deep Rose, which was subjected to acute gamma irradiation at a quantitative 50-200 Gy (30). It was found that the survival rate decreased as the irradiation dose increased.

The heights of 'Siam Shadow' and 'Siam Scarlet' seedlings were observed after 4 weeks of planting in the greenhouses. The control 'Siam Shadow' and 'Siam Scarlet' seedlings had the average maximum pseudo-stem heights of 21.91 ± 0.61 and 18.96 ± 0.88 cm respectively (Table 2 and Fig. 3). The height of the stem decreased as the irradiation dose increased corresponding to one reported that the height of the *Gomphrena* Hybrid plant after irradiation

Table 2. Survival rate and height of 'Siam Shadow' and 'Siam Scarlet' seedlings after gamma irradiation *in vivo*.

Dose rate (Gy)	Survival rate (%)		No. of shoots/explant	
	Siam Shadow	Siam Scarlet	Siam Shadow	Siam Scarlet
Control	100 ± 0.00^d	100 ± 0.00^d	21.91 ± 0.61^e	18.96 ± 0.88^d
20	100 ± 0.00^d	100 ± 0.00^d	21.21 ± 0.18^e	13.81 ± 0.75^c
30	88 ± 1.33^c	62.50 ± 4.43^b	14.39 ± 0.71^c	8.02 ± 1.25^b
40	84 ± 2.69^c	0 ± 0.00^a	10.34 ± 0.52^b	0.00 ± 0.00^a
80	0 ± 0.00^a	0 ± 0.00^a	0.00 ± 0.00^a	0.00 ± 0.00^a
cultivars (A)	*			*
dose (B)	*			*
A x B	*			*

* There was a statistically significant difference at the 95 % confidence level. The mean \pm standard deviation followed by different superscripts within the same column was statistically different by Scheffe's test.

had statistically and significantly decreased (31). *Curcuma* Hybrid height decreased as the gamma irradiation dose increased because of the response of plant tissue to gamma irradiation (23). Gamma irradiation, especially at lower doses, has gained widespread recognition for its ability to enhance the growth attributes of numerous economically important plants (32). It has been statistically analyzed that the cultivars of *C. alismatifolia* ('Siam Shadow' and 'Siam Scarlet') and gamma irradiation have mutual influence and affect the survival rate and height of pseudo-stem with a statistical significance at a 95 % confidence level.

After planting in greenhouses, changes in the flower morphology of 'Siam Shadow' were observed at 30 Gy of gamma irradiation. The bracts and flowers separated from the inflorescences and the number of bracts decreased. 'Siam Shadow' at the 40 Gy of gamma irradiation had withered flowers. For 'Siam Scarlet' exposed to 20 Gy of gamma irradiation, small inflorescences and 2 bracts were observed (Fig. 4), consistent with one study (33). The dose of gamma irradiation of 10-30 Gy changed the morphology of the flower, the number of layers of petals and flower color. These abnormalities may manifest themselves through the phenotype after irradiation. This is caused by irradiation affecting gene changes. If one of the genes is abnormal, it can affect the formation of pigments in plants (34).

After planting the 'Siam Scarlet' that had undergone 30 Gy of gamma irradiation, there were 2-lobed leaves and a spotted leaf appearance (Fig. 4A, 4B). Such features were the result of the function to reduce the irradiation, chemical, and mutation. If the cell cannot be repaired however, it will cause a mutation (35). Gamma irradiation will cause mutations or alterations in the genetic material of cells and can



Fig. 3. The seedling of 'Siam Shadow' and 'Siam Scarlet' after gamma irradiation and planted in pots for a period of 4 weeks. (A-E) 'Siam Shadow' seedling: control, 20, 30, 40 and 80 Gy, (F-H) 'Siam Scarlet': control, 20 and 30 Gy.



Fig. 4. Characteristics of 'Siam Shadow' and 'Siam Scarlet' flowers after planting in greenhouses. (A) 2-lobed blade of 'Siam Scarlet' (30 Gy), (B) Spotted leaves of 'Siam Scarlet' (30 Gy), (C) 'Siam Shadow' control, (D) 'Siam Scarlet' control, (E, F, G) Change of 'Siam Shadow' at petals, inflorescences and bracts (30 Gy) and (H) Small inflorescences and 2 bracts of 'Siam Scarlet' (20 Gy).

transmit this change to the offspring cells (36). Irradiation has proven to be successful in mutation breeding across various crops and ornamental plants (10, 37), resulting in morphological changes in the color and shape of the stem, leaves and flowers. This process has led to the generation of novel genetic variations (38, 39), with new characteristics exclusively emerging in the treated plants (40, 41).

Conclusion

Gamma irradiation doses and cultivars of *C. alismatifolia* 'Siam Shadow' and 'Siam Scarlet' mutually influence the induction of shoots, as well as the height of pseudo-stem and survival rate after planting. The seedling of 'Siam Scarlet' LD₅₀ is 29.06 Gy. Changes in leaf morphology were observed and found to have light green leaves and a pale white stem for 'Siam Shadow'. Chimera leaf formation was found *in vitro* when irradiated at 40 Gy of gamma irradiation, while ornamental leaves and flowers separated from the inflorescence decreased. Wilted flowers were found when 'Siam Shadow' was irradiated at 30 Gy. On the other hand, 'Siam Scarlet' that was irradiated at 20 Gy of gamma irradiation was found in the chimera leaves *in vitro*. After planting, the inflorescence was small, with 2 ornamental leaves. At gamma irradiation of 30 Gy, there were 2-lobed leaves and spotted leaf formation. The heights of pseudo-stems were reduced, unlike the control. This study included the guidance of *C. alismatifolia* breeding by irradiation of seedlings obtained from plant tissue culture of *C. alismatifolia* 'Siam Shadow' and 'Siam Scarlet', with the aim of providing initial towards developing new characteristic for farmers.

Acknowledgements

This research was completed with the assistance and valuable guidance of Dr. Tharathorn Teerakathiti. The researchers would also like to express their heartfelt gratitude for the support received from the Thailand Institute of Nuclear Technology (Public Organization).

Authors' contributions

SP conducted the design of the study, carried out the experiments. SI and SA participated in the plant collection, assistant the experiments. CW conducted the design of the study, revised the manuscript. NB revised the manuscript. PW performed the statistical analysis. VP revised the manuscript.

Compliance with ethical standards

Conflict of interest: The authors here with declare no conflict of interest.

Ethical issues: None.

References

1. Tosri C, Chusreeaom K, Limtiyayotin M, Sukin N, Jompuk P. Comparative effect of high energy electron beam and ¹³⁷Cs gamma ray on survival, growth and chlorophyll content in *curcuma* hybrid 'Laddawan' and determine proper dose for mutations breeding. EJFA. 2019;31(5):321-27. <https://doi.org/10.9755/ejfa.2019.v31.i5.1942>
2. Taychasinpitak T, Sreeboonraung S, Wongpiyasatid A, Lekawatana S. Effects of chronic gamma irradiation on mutations

- of weeping fig (*Ficus benjamina* var. *variegata*). *Agricultural Sci J (Thailand)*. 2007;38(2):143-49.
3. Sumyo S, Athalangrong A. [Internet]. Department of Agriculture Extension online; 2017 Sep 15 [cited 2023 Jan 1]. Available from: <https://www.doa.go.th/hort/wpcontent/uploads/2021/11/>
 4. KP Holland. Growing and breeding *Curcuma* online [Internet]. KP Holland online; 2020 Mar 1 [cited 2023 Jan 20]. Available from: <https://www.kpholland.nl/en/breeding/siam-curcuma/>
 5. Keawbua K, Nualmanee R, Ruamrungsri S, Hongpakdee P. Development of *Curcuma* Hybrids cv. 'Doi Tung Red' as Potted Plant by Paclobutrazol Drenching. *Khon Kaen AGR J*. 2018;46(SUPPL.1):375-80.
 6. Teerakathiti T, Phaepoon W, Kongkaew P. The process of using disease-free Pathumma plant pseudo-stems with increasing the number of shoots [Internet]. Ipthailand online; 2013 Aug 31 [cited 2022 Sep 9]. Available from: <https://search.ipthailand.go.th/>
 7. Ketmaro S, Taychasinpitak T, Mongkolchaiyaphruek A, Wongchaochant S. Effect of colchicine on increasing pollen viability in a *Curcuma* hybrid (*Curcuma sparganiifolia* × *C. parviflora*). *Kasetsart J Nat Sci*. 2012;46:363-70.
 8. Ye Y, Zhou Y, Tan J, Zhu G, Liu J, Xu Y. Cross-Compatibility in Interspecific Hybridization of Different *Curcuma* Accessions. *Plants*. 2023;12(10):1961. <https://doi.org/10.3390/plants12101961>
 9. Majeed A, Muhammad Z, Ullah R, Ali H. Gamma irradiation i: effect on germination and general growth characteristics of plants—a review. *Pak J Bot*. 2018;50(6):2449-53.
 10. Abdullah TL, Johari E, Mohd N. Changes in flower development, chlorophyll mutation and alteration in plant morphology of *Curcuma alismatifolia* by gamma irradiation. *Am J Appl Sci*. 2009;6(7):1436-39. <https://doi.org/10.3844/ajassp.2009.1436.1439>
 11. Taheri S, Abdullah TL, Ahmad Z, Abdullah NAP. Effect of acute gamma irradiation on *Curcuma alismatifolia* varieties and detection of DNA polymorphism through SSR marker. *Biomed Res Int*. 2014. 2014;18. <https://doi.org/10.1155/2014/631813>
 12. Phakpaknam S, Yunchum N, Dechkla M, Pikunthong V. *In vitro* micropropagation and microrhizome induction of *Curcuma comosa* Roxb. *RMUTP Research Journal*. 2022;16(2): 13-23. <https://doi.org/10.14456/jrmutp.2022.18>
 13. Sirirugsa P, Larsen K, Maknoi C. The genus *Curcuma* L. (Zingiberaceae): distribution and classification with reference to species diversity in Thailand. Tan BC, Leong-Skornickova J, editors. In: *Proceedings of the 4th International Ginger Symposium*; 2006 Jul 3-6; Singapore. *The Gardens' Bulletin*; 2007. P.203-20.
 14. Cayamarit K, Balslev H, Newman MF, Sangvirotjanapat S. e-Flora of Thailand Volume 16, Part 2: Zingiberaceae: [e-book]. Forest Herbarium (BKK): Royal Forest Department; 2023 [cited 2023 Sep 20]: 145. Available from: <https://botany.dnp.go.th/eflora/floragenus.html?factsheet=Curcuma>.
 15. Murashige T, Skoog F. A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol Plant*. 1962;15:473-97.
 16. Limtiyayothin M, Taychasinpitak T, Jompuk P. Gamma ray-induced *in vitro* mutations in bromeliad: *Tillandsia cyanea*. *Khon Kaen AGR J*. 2018;46(5):983-90.
 17. Kashima S, Thanananta T, Kachonpadungkitt Y, Bodhipadma K. Induced mutation in *Musa acuminata* genomes AAA using gamma ray and determination with HAT-RAPD marker. *TJST*. 2020;9(5):669-79. <https://doi.org/10.14456/tjst.2020.71>
 18. Beyaz R, Kahramanogullari CT, Yildiz C, Darcin ES, Yildiz M. The effect of gamma radiation on seed germination and seedling growth of *Lathyrus chrysanthus* Boiss. under *in vitro* conditions. *J Environ Radio Act*. 2016;162:129-33. <https://doi.org/10.1016/j.jenvrad.2016.05.006>
 19. Beyaz R. Impact of gamma irradiation pretreatment on the growth of common vetch (*Vicia sativa* L.) seedlings grown under salt and drought stress. *Int J Radiat Biol*. 2020;96(2):257-66. <https://doi.org/10.1080/09553002.2020.1688885>
 20. Sittinisaisukand S, Taychasinpitak T, Jompuk P, Chanchula N. Effect of acute gamma irradiation on *in vitro* culture of *Lindernia* spp. *TJST*. 2018;7(2):159-68. <https://doi.org/10.14456/tjst.2018.25>
 21. Gang L, Xiaoying Z, Yiging Z, Qingcheng Z, Xun X, Jiashu C. Effect of radiation on regeneration of *Chinese narcissus* and analysis of genetic variation with AFLP and TAPD markers. *PCTOC*. 2007;88;319-27. <https://doi.org/10.1007/s11240-006-9189-9>
 22. Jan S, Parween T, Hameed R, O Siddiqi T. Effects of presowing gamma irradiation on the photosynthetic pigments, sugar content and carbon gain of *Cullen corylifolium* (L.) Medik. *Chil J Agric Res*. 2013;73(4):345-50. <https://doi.org/10.4067/S0718-58392013000400003>
 23. Veeraborirak P, Taychasinpitak T, Sukprasert P, Piriya-phattarakit A. Effect of gamma irradiation on morphological of character *Curcuma* Hybrid. *Agricultural Sci J (Thailand)*. 2020;9(3):243-50.
 24. Venketeswaran S, Partanen CR. A comparative study of the effects of gamma radiation on organized and disorganized growth of tobacco. *Radiat Bot*. 1966;6(1):13-20. [https://doi.org/10.1016/S0033-7560\(66\)80089-3](https://doi.org/10.1016/S0033-7560(66)80089-3)
 25. Chanchula N. Mutational induction in *Globba williamsianaby* Gamma irradiation. *TJST*. 2013;2(1):45-52. <https://doi.org/10.14456/tjst.2013.7>
 26. Wongpiyasatid A. *Mutation: for plant breeding*. 1st ed. Bangkok: Kasetsart University press; 2017.
 27. Ulukapi K, Ozmen SF. Study of the effect of irradiation (60Co) on M1 plants of common bean (*Phaseolus vulgaris* L.) cultivars and determined of proper doses for mutation breeding. *JRRAS*. 2018;11(2):157-61. <https://doi.org/10.1016/j.jrras.2017.12.004>
 28. Volkova PY, Duarte GT, Soubigou-Taconnat L, Kazakova EA, Pateyron S, Bondarenko VS, et al. Early response of barley embryos to low-and high-dose gamma irradiation of seeds triggers changes in the transcriptional profile and an increase in hydrogen peroxide content in seedlings. *J Agron Crop Sci*. 2020;206(2):277-95. <https://doi.org/10.1111/jac.12381>
 29. Wi SG, Chung BY, Kim JS, Kim JH, Baek MH, Lee JW, et al. Effects of gamma irradiation on morphological changes and biological responses in plants. *Micron*. 2007;38(6):553-64. <https://doi.org/10.1016/j.micron.2006.11.002>
 30. Padungsil N, Taechasinpitak T, Wongchowchan S, Chanchula N. Mutational induction in *Catharanthus roseus* L. by acute gamma irradiation. *TJST*. 2015;4(1):95-103. <https://doi.org/10.14456/tjst.2015.4>
 31. Taktuea S, Taychasinpitak T, Sukprasert P, Piriya-phattarakit A, Balla N. Effect of acute gamma irradiation on *Gomphrena* Hybrid *in vitro*. *TJST*. 2018;7(6):605-13. <https://doi.org/10.14456/tjst.2018.59>
 32. Kim JH, Baek MH, Chung BY, Wi SG, Kim JS. Alterations in the photosynthetic pigments and antioxidant machineries of red pepper (*Capsicum annum* L.) seedlings from gamma-irradiated seeds. *J Plant Biol*. 2004;47:314-21. <https://doi.org/10.1007/BF03030546>
 33. Kongprakhon P, Jitbumrong N. Utilization of gamma-rays induced morphology changes in miniature rose (*Rosa chinensis*

- Jacq. var. *minima* Voss). Khon Kaen AGR J. 2017; 45 (SUPPL.1):1296-302.
34. Pangjai W, Huehne-Srifah P. Gamma-ray induced morphological changes in rain lily (*Zephyranthes* spp.). Khon Kaen AGR J. 2021;49(3):643-55.
35. Lamseejan S. Plant Mutation. Bangkok: Kasetsart University Press;1997.
36. Chanchula N, Jala A. Mutational Induction in *Pinguicula* sp. by Gamma Irradiation. TJST. 2014;3(2):76-81. <https://doi.org/10.14456/tjst.2014.6>
37. Muhallilin I, Aiyah SI, Sukma D. The diversity of morphological characteristics and chemical content of *Celosia cristata* plantlets due to gamma ray irradiation. Biodiversitas Journal of Biological Diversity. 2019;20(3):862-66. <https://doi.org/10.13057/biodiv/d200333>
38. Jo YD, Kim SH, Hwang JE, Kim YS, Kang HS, Kim SW, et al. Construction of mutation populations by gamma-ray and carbon beam irradiation in chili pepper (*Capsicum annuum* L.). Hortic Environ Biotechnol. 2016;57:606-14. <https://doi.org/10.1007/s13580-016-1132-3>
39. Shin JM, Kim BK, Seo SG, Jeon SB, Kim JS, Jun BK, et al. Mutation breeding of sweet potato by gamma-ray radiation. Agric Nat Resour. 2011;6(6):1447-54.
40. Lamseejan S, Peeranuch J, Arunee W, Surin D, Prapanpongse K. Gamma-rays induced morphological changes in chrysanthemum. (*Chrysanthemum morifolium*). Nat Sci. 2000;34(3):417-22.
41. Topoonyanont, N, Jaikanta S, Boonmanee P. *Curcuma alismatifolia* Gagnep. micropropagation in twin-flasks temporary immersion bioreactor. In: van den Ende JE, Krikke AT, den Nijs APM, editors. ISHS Board: In: Proceedings of the X International Symposium on Flower Bulbs and Herbaceous Perennials 886; 2008 Apr 20-24; Netherlands. Lisse; Acta Horticulturae, 2008. p. 267-71. <https://doi.org/10.17660/ActaHortic.2011.886.37>