



Immunomodulatory and anti-inflammatory effects of active compounds of *Cassia angustifolia* Vahl. leaf extract

Walaa Najm Abood

RESEARCH ARTICLE

Department of Microbiology, College of Vet. Medicine, University of Diyala, Baquba 32001, Iraq

*Email: walaaabood@gmail.com

ARTICLE HISTORY

Received: 03 December 2023 Accepted: 02 January 2024

Available online Version 1.0 : 02 April 2024 Version 2.0 : 14 April 2024

() Check for updates

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/ journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/ index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an openaccess article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (https://creativecommons.org/licenses/ by/4.0/)

CITE THIS ARTICLE

Abood WN. Immunomodulatory and antiinflammatory effects of active compounds of *Cassia angustifolia* Vahl. leaf extract. Plant Science Today. 2024; 11(2): 373–377. https:// doi.org/10.14719/pst.3159

Abstract

Cassia angustifolia medicinal plant used for the treatment many diseases such as respiratory diseases, skin inflammation, and has the ability of antibacterial and anticancer properties. In this work was investigated the Immunmodulatory and anti-inflammatory effect of *C. angustifolia*. Methods: The ethanol leaf extract of C. angustifolia was utilized to examine gene expression related to angiogenesis cytokines in treated RAW 264.7 macrophage cell with C. angustifolia leaf extract. The results demonstrated increasing significantly in the treated macrophage RAW 264.7 viability accompanied by an improvement in angiogenesis cytokines expression with the inhabited Nitric Oxide production as dose depended manner. GCMS analysis was identified 11 active components within the extract, each exhibiting biological activity as antioxidant, antitumor and anti-inflammatory, Notable compounds include hexadecanoic acid, 2 Pentadecanone, Phthalic acid, oxalic acid, carbonic acid, Tricosane, Undecanal and many others. Conclusion: Ethanol leaf extract of C. angustifolia exhibits the immunmodulatory and anti-inflammatory effect through inhibition NO production and increasing angiogenesis cytokines expression.

Keywords

anti-Inflammatory bioactive compounds; Cassia angustifolia ; immunomodulation

Introduction

Traditional plants were widely use in recent years to treat a lot of diseases instead chemical drugs with fewer side effects that may be effective throughout the use of chemical drugs for a long time. *C. angustifolia* is one of the traditional medicines with the common name senna in Makkah the city in the Saudi Arabia country was discovered and used in the different place in the world for pharmacological purpose (1).

C. angustifolia is used in the pharmacological treatment for many diseases as respiratory inflammation, skin infection, piles, and heart disease (2). The research application used *C. angustifolia* extract for antifungal and antibacterial (3, 4). Other study was applied *C. angustifolia* leaf extract for treatment hemorrhoid and colon inflammation (5). In the pharmacological purpose different parts from *C. angustifolia* plant are used in the medicine as leaf, roots, and seeds. That diagnosed have a lot of active compounds like flavonoid, phenol, alkaloids, naphthopyrone glycosides and investigated for their biological activities: anti-inflammation, anti-oxidants, hepatoprotective, and antimicrobial (6-8).

The immune response is the interacting between innate and specific response to immune process. Phytochemical have an effect on the immune responses through enhanced modulation in the one or more components of immune response (9). Immunomodulation compounds like alkaloids, phenols, polysaccharides, and lectins (10). Many plants were investigated for their immunomodu-latory effect as *Aloe vera*, *Allium sativum, Curcuma longa, Centella asiatica, Piper longum*, and *Ocimum sanctum* [11]. One of the modulation effect enhanced apoptosis process (12).

In previous study was showed the Immunotoxicity of Senegalia greggii seed extract by effect on the CD8 Tcells, CD4 Tcell, the level of TGF-beta and TNF-alpha and effect on the inflammatory process (13). A lot of countries dependent on the herbal medicine in the cure of many diseases like China, Japan, Iraq, Egypt, and India, which play an important role in the modulation of immune system (14). Phytochemical polysaccharide in the medicinal plants act as anti tumor (15). Methanol extract of Enicostema axillare effect on both cell mediated and humoral immune process by decreasing pro inflammatory mediators in the macrophages (16). Given the significant role of the important C. angustifolia in medicine used in the different field, this study was aimed investigated the immunomodulatory and anti inflammatory influence ethanol leaf extract of C. angustifolia.

Materials and Methods

Plant extraction

C. angustifolia also common known as (Senna alexandrina Mill) leaf Figure 1, was purchased under voucher number (EM3.12). The ethanol extract was done as described by Abood *et al.* <u>(17)</u>.

Immunomodulation effect of C. angustifolia



Fig. 1. Cassia angustifolia (Senna alexandrina Mill).

The immunomodulatory effect of ethanol leaf extract of C. angustifolia was investigated on the effect RAW 264.7 macrophages cells via measuring cell viability after treatment with different doses from extract at different time exposure. The final concentration of extract was 125, 250,500 and 1000 μ g/mL, the method was done as described by Abood (9).

Effect of C. angustifolia on the production of Nitric oxide

The impact of *C. angustifolia* on the production of Nitric oxide (NO) from macrophage RAW 264.7 cell was investigated by exposed the cell to graded concentration doses 25, 50, 100, 200 μ g/mL from *C. angustifolia* ethanol leaf extract by applied Griess method (18). Stimulation of RAW 264.7 cell at the number 1x10⁶ cells / well by incubated with LPS (100 μ g/mL) with extract in the plate at 24h at 37°C with 5% CO₂. The cells were treated with LPS be as positive control. A dilution of NaNO2 was prepared as standard curve. Aspirated culture media and centrifuged at 4000 r.p.m for 10 min. Griess reagent was added to the an equal amount from supernatant and incubated at room temperature in the dark place at 30 min, at the end of the time read the absorbance was at 546 nm.

Anti-inflammatory effect of C. angustifolia

To investigated anti inflammatory influence leaf extract of *C. angustifolia* through initiated angiogenesis process to produce many angiogenesis expression proteins and cytokines in the excited macrophage RAW 264.7 cell line (from Raybio C-series angiogenesis, USA).

Method involves treating the cells in the 100 μ g/mL C. angustifolia leaf extract, 24h. Membrane for Antibody array placed in the tray then two mL block buffer added buffer for thirty min, the end of the incubation time removed it by aspiration. One mL from media to treated cell is added to antibody array membrane, incubated to three hours at room temperature in the end of incubated time was aspirated. Membrane was washed in the 1X wash buffer 1 by 2 mL for 5 min at room temperature, replicated wash buffer three times. Repeated wash step by wash buffer 2 as the same step for wash buffer 1. Added 1 mL from biotinylated antibody on the membrane well, incubated 2 h at room temperature and removed at the end of incubation. Repeated the wash step. HRP-streptavidin 2 mL in the 1X concentration is inserting on membrane then incubated to 2h at room temperature and removed at end of incubation, repeat wash step. Then transfer membrane to the plastic sheet, 500 μ L from detection buffer added and a chemilluminescece imaging system was employed to obtain the results.

Diagnosis of active compounds

Leaf *C. angustifolia* extract was applied for gas chromatography mass spectrum analysis, is done by MSDCHEM device. Phytochemical analysis is done through inoculation $0.5 \,\mu$ L extract, and divided ratio was 1:50, at 250 temperature for injection and back eluent. Flow column 1mL/min. Helium used as carrier gas and the temperature oven at 60 to 246. Collected rate is 4 C/ min.

Detection active compounds through spectrum for the compound and compared with willy and NIST/EPA/NIH mass spectral library.

Analysis of result

Data were presented as (mean \pm SD), statistical analysis of result was done using T-test and significant with P value < 0.05. Analysis for gene expression of angiogenesis and pro-

tein array was done through C-Series angiogenesis analysis manufacture software.

Results

Immunomodulatory effect for C. angustifolia leaf extract

Immunomodulatory effect for ethanol *C. angustifolia* leaf extract on the treated RAW 264.7 cell line was showed increased in the viability of the cells significantly at P value < 0.05 to dose dependent manner compare to normal not treated cells as presented in the Figure 2.

Effect of C. angustifolia on the production of Nitric oxide (NO)

The impact of *C. angustifolia* on the production of Nitric oxide from treated macrophage RAW 264.7 cell showed the production of Nitric oxide was decreased significant at P



Fig. 2. The immunomodulatory effect of *Cassia angustifolia* leaf extract for treating RAW 264.7 cell. *Significant at *P* < 0.05.

value < 0.05 in the stimulated RAW 264.7 cells for differ doses from leaf extract at depended manner compared with non-stimulated RAW 264.7 cell, presented in the Figure 3.

Anti-inflammatory effect of C. angustifolia

The application of *C. angustifolia* leaf extract to stimulate angiogenesis process to produce many angiogenesis expression proteins on the stimulated macrophage RAW 264.7 cell with *C. angustifolia* leaf extract showed different



Fig. 3: The effect of *Cassia angustifolia* on the production of Nitric oxide from treated macrophage RAW 264.7 cell showed the production of Nitric oxide (μ M). *Significant at *P* < 0.05.

excite to the angiogenesis expression proteins, increased in the fold of expression for the angiopoietin-1, angiopoietin-2, angiostatin, GM-CSF, I-309, IL-10, IL-1 alpha, IL-1 beta, MMP-1, MMP-9, PECAM-1, Tie-2, TNF-alpha, μ PAR, and VEGFR2. While down expression in the angio-genesis expression proteins for Endostatin, GCSF, MCP-4, IL-2, IL-4, I- **Table 1:** Angiogenesis expression proteins on the stimulated macrophage

 RAW 264.7 cell with ethanol leaf extract of *Cassia angustifolia*

375

Protein	Symbols gene	Fold ex- pression	Fold ex- pression in treated cell with C. angustifolia	Percent- age Fold changes
Angiopoietin-1	ANGPT1	2762.675	6770.344	59.19447
Angiopoietin-2	ANGPT2	4834.975	7834.035	38.28244
Angiostatin	CCL1	559277.1	600187.1	6.816214
Endostatin	COL18A1	668347.1	673143.6	0.712558
GCSF	CSF3	33569.43	29067.93	-15.4861
GM-CSF	CSF2	31413.93	33800.4	7.060493
I-309	(TCA-3/CCL1)	99337.08	124980.3	20.51779
IL-10	IL10	108170.6	145961.7	25.8911
IL-1 alpha	IL1A	8907.825	14250.52	37.49123
IL-1 beta	IL1B	10479.43	12006.72	12.72033
IL-2	IL2	2.775	0.05821	-4667.22
IL-4	IL4	2.775	0.05821	-4667.22
I-TAC	(CXCL11)	2.775	0.05821	-4667.22
MCP-3	(MARC/CCL7)	2.775	0.05821	-4667.22
MCP-4	(CCL13)	44840.08	40529.03	-10.6369
MMP-1	MMP1	48422.58	50173.64	3.490018
MMP-9	MMP9	42384.58	49700.98	14.72084
PECAM-1	(CD31)	42624.58	66146.85	35.56069
Tie-2	TEK	24275.58	33289.26	27.07685
TNF alpha	TNF	29879.58	35775.79	16.48103
uPAR	PLG	278274.9	333709.8	16.61171
VEGFR 2	K D R	319074.9	351153	9.135184
VEGFR 3	FLT4	29899.58	28418.83	- 5.21044

Phytochemical analysis of Cassia angustifolia

Phytochemical analysis of Cassia angustifolia was done by applied to the GC-MS analysis and the analysis was diagnosis eleven active compounds listed in the Table 2.

Discussion

Previous studies have extensively documented the diverse biological activities of *C. angustifolia*. This current study

Table 2: Phytochemica	l analysis (GM-CS) of	Cassia angustifolia	leaf extract
-----------------------	-----------------------	---------------------	--------------

Active compound	Formula	RT	Area%
Oxalic acid	$C_{16}H_{30}O_4$	17.15	3.10
2-Pentadecanone	C ₁₈ H ₃₆ O	19.77	4.59
Carbonic acid	C ₂₂ H ₄₄ O ₃₃	19.77	4.59
Methoxyacetic acid	$C_{18}H_{36}O_3$	20.11	1.98
Tricosane	C ₂₃ H ₄₈	20.11	1.98
Hexadecanoic acid	$C_{17}H_{34}O_2$	20.88	7.04
Heptadecanoic acid	$C_{19}H_{38}O_2$	23.89	2.44
Hexadecyl propyl ether	C ₁₉ H ₄₀ O	24.51	1.41
Undecanal	$C_{11}H_{22}O$	26.45	2.47
Octadecanoic acid	$C_{20}H_{40}O_2$	26.45	2.47
Phthalic acid	$C_{16}H_{22}O_4$	29.66	31.48

ABOOD

contributes evidence supporting of C. angustifolia leaf extract as modulation for immune response by increasing the RAW cells viability through effect on the Nitric oxide production and effect on the expression of angiogenesis protein expression. The angiogenesis process is important in the inflammation through effect in the different roles. Angiopoietin was act synergetic with VEGF to stimulate angiogenesis and enhanced remodeling and controlled of the formation blood vessel (19). Some medicinal plants were investigated for NO modulation like *Linum persicum*, Dionysia termeana, Salvia mirzayanii and Ocimum gratissimum and proven effect to decreased NO production with significant inhibited TNF-alpha and IL-1 acting as antiinflammatory agents (20, 21). Angiostatin is plasminogen has potential inhibitor endothelial cell proliferation and metastasis of tumor (22). The outcome provide an evidence of the anti inflammatory potential of *C. angustifolia*. The Endostatin has anti cancer effect through tumor regression caused inhibition angiogenesis. The role of GCSF was important in the existence, differentiation and proliferation of neutrophil from heamopoietic, down expression of GCSF was effect on the maturation of neutrophil and synergetic effect on the inflammation process (23, 24). C. angustifolia leave extract was leading to increase the production of GM-CSF in the treated cells. The GM-CSF is effect on the development inflammation process, immunomodulator (25).

The effect of C. angustifolia leave extract on the modulation production of inflammatory cytokines as IL-10, IL-1 alpha and beta, and TNF-alpha, the modulation caused increasing or decreasing influence for promote inflammation and limiting immune response to pathogens and stimulating macrophage, give T cell development and differentiation (26-28). Allium cepa (onion) was consumed through the world, it has many phytochemical as sapoines, quercetin, falvonoids, cepanens, phenolic compounds, and organ sulfurs all the antioxidant and immunomodulatory effect by reduced Th cell cytokines but increasing CD4 cells cytokines (29). Chemokines I-TAC is a mediator to the T cells migration and activation and it is role influence by INF-gamma level (30). Ocimum basilicem used for treatment respiratory tract infection have anti inflammatory and immunomodulatory potential by increasing INF- gamma and decreasing IL4 in the treated rats (31). In the many of previous studies were showed the pharmacological potential of medicinal plants as anti inflammatory and immunomodulatory agents may be used as drug for treatment many diseases (32).

Additional evidence for immunomodulation of *C.* angustifolia is increasing in the fold expression of MMP family, PECAM-1, TNF alpha, VEGFR2 and μ PAR. These inflammations proteins have lot of important and crucial role in the immune response and defense, through play role for recruitment of leukocyte, cytokine production, inflammation process, adhesion of platelet (33, 34). The macrophage plasminogen activators is μ PAR (urokineas) that control for phagocytosis to the chronic inflammation leads to immunomodulation and development of immune response by supportive matrix generation and control to

the cells migration, adhesion, proliferation the process in the inflammatory response (35).

Conclusion

C. angustifolia has an important and potential role in the immune response through play the role in the effect on the up and down expression and fold change for angio-genesis proteins. And may be depend in the future drug developed and discover a new immunomodulator therapy as antitumor, immunoregulator and anti-diabetic.

Compliance with ethical standards

Conflict of interest: The author declared no conflict of interest.

Ethical issues: None.

References

- Khan MA, Ahmad M, Zafar M, Sultana S, Marwat SK, Shaheen S, Leghari MK, Jan G, Ahmad F, Nazir A. Medico-botanical and chemical standardization of pharmaceutically important plant of *Tricholepis chaetolepis* (Boiss) Rech. f. Journal of Medicinal Plants Research. 2011;5(8):1471-77.
- Sultana S, Ahmad M, Zafar M, Khan MA, Arshad M. Authentication of herbal drug Senna (*Cassia angustifolia* Vahl.): A village pharmacy for Indo-Pak subcontinent. African Journal of Pharmacy and Pharmacology. 2012;6(30):2299-308. https:// doi.org/10.5897/AJPP12.446
- Akrayi HF. Antibacterial effect of aqueous extracts of spices and herbs against bacteria isolated from frozen meat. Medical Journal of Islamic World Academy of Sciences. 2014;22(1):30-35. https://doi.org/10.12816/0008168
- 4. Al-Marzoqi AH, Hadi MY, Hameed IH. Determination of metabolites products by *Cassia angustifolia* and evaluate antimicobial activity. Journal of Pharmacognosy and Phytotherapy. 2016 Feb 29;8(2):25-48. https://doi.org/10.5897/JPP2015.0367
- Naqishbandi A. Plants used in Iraqi traditional medicine in Erbil-Kurdistan region. Zanco Journal of Medical Sciences. 2014;18 (3):811-15. https://doi.org/10.15218/zjms.2014.0038
- Singh S, Singh SK, Yadav A. A review on *Cassia* species: Pharmacological, traditional and medicinal aspects in various countries. American Journal of Phytomedicine and Clinical Therapeutics. 2013 Jun 30;1(3):291-312.
- Hussein H, Al-Khafaji NM, Al-Mamoori AH, Al-Marzoqi AH. Evaluation of *in vitro* antibacterial properties of the crude phenolic, alkaloid and terpenoid extracts of *Cassia senna* L. against human gram-negative pathogenic bacteria. Plant Archives. 2018;18(1):354-56.
- Agarwal S, Pandey V. Antioxidant enzyme responses to NaCl stress in *Cassia angustifolia*. Biologia Plantarum. 2004;48(4):555-60. https://doi.org/10.1023/B:BIOP.0000047152.07878.e7
- Porwal O, Ozdemir M, Kala D, Anwer ET. A review on medicinal plants as potential sources of natural immunomodulatory action. Journal of Drug Delivery and Therapeutics. 2021;11(6):324-31. https://doi.org/10.22270/jddt.v11i6.5125
- Parbat AY, Malode GP, Shaikh AR, Panchale WA, Manwar JV, Bakal RL. Ethnopharmacological review of traditional medicinal plants as immunomodulator. World Journal of Biology Pharmacy and Health Sciences. 2021;6(2):43-55. https:// doi.org/10.30574/wjbphs.2021.6.2.0048

- 11. Mukherjee PK, Nema NK, Bhadra S, Mukherjee D, Braga FC, Matsabisa MG. Immunomodulatory leads from medicinal plants. 2014.
- 12. Amirghofran Z, Bahmani M, Azadmehr A, Javidnia K, Miri R. Immunomodulatory activities of various medicinal plant extracts: Effects on human lymphocytes apoptosis. Immunological Investigations. 2009;38(2):181-92. https:// doi.org/10.1080/08820130902817051
- 13. Abood WN, Al-Obaidy EN. Immunotoxicity of Senegalia greggii seed extract. HIV Nursing. 2022 Nov 4;22(2):3167-71.
- Shantilal S, Vaghela JS, Sisodia S. Review on immunomodulation and immunomodulatory activity of some medicinal plant. Eur J Biomed. 2018;5(8):163-74.
- 15. Wong CK, Leung KN, Fung KP, Choy YM. Immunomodulatory and anti-tumour polysaccharides from medicinal plants. Journal of International Medical Research. 1994;22(6):299-312. https://doi.org/10.1177/030006059402200601
- Saravanan S, Babu NP, Pandikumar P, Raj MK, Paulraj MG, Ignacimuthu S. Immunomodulatory potential of Enicostema axillare (Lam.) A. Raynal, a traditional medicinal plant. Journal of Ethnopharmacology. 2012;140(2):239-46. https://doi.org/10.1016/ j.jep.2012.01.010
- Abood WN, Fahmi I, Abdulla MA, Ismail S. Immunomodulatory effect of an isolated fraction from *Tinospora crispa* on intracellular expression of INF-γ, IL-6 and IL-8. BMC Complementary and Alternative Medicine. 2014;14:1-12. https:// doi.org/10.1186/1472-6882-14-205
- Vašíček O, Lojek A, Číž M. Serotonin and its metabolites reduce oxidative stress in murine RAW264. 7 macrophages and prevent inflammation. Journal of Physiology and Biochemistry. 2020;76 (1):49-60. https://doi.org/10.1007/s13105-019-00714-3
- Koblizek TI, Weiss C, Yancopoulos GD, Deutsch U, Risau W. Angiopoietin-1 induces sprouting angiogenesis *in vitro*. Current Biology. 1998;8(9):529-32. https://doi.org/10.1016/S0960-9822(98) 70205-2
- Amirghofran Z, Malek-Hosseini S, Golmoghaddam H, Kalantar F, Shabani M. Inhibition of nitric oxide production and proinflammatory cytokines by several medicinal plants. Iranian Journal of Immunology. 2011;8(3):159-69.
- Djafoua YM, Mouokeu RS, Tume C, Kamtchueng MO, Kuiate JR. Immunomodulatory activity of methanol leaf extracts of Cameroonian medicinal plants. Journal of Complementary and Integrative Medicine. 2015;12(4):267-75. https://doi.org/10.1515/ jcim-2014-0023
- 22. Soff GA. Angiostatin and angiostatin-related proteins. Cancer and Metastasis Reviews. 2000 Jun;19:97-107. https:// doi.org/10.1023/A:1026525121027
- Shichiri M, Hirata Y. Antiangiogenesis signals by endostatin. The FASEB Journal. 2001;15(6):1044-53. https://doi.org/10.1096/ fsb2fj991083com

- Roberts AW. G-CSF: A key regulator of neutrophil production, but that's not all. Growth Factors. 2005;23(1):33-41. https:// doi.org/10.1080/08977190500055836
- Hamilton JA. GM-CSF in inflammation. Journal of Experimental Medicine. 2020;217(1):e20190945. https://doi.org/10.1084/ jem.20190945
- Miller MD, Krangel MS. The human cytokine I-309 is a monocyte chemoattractant. Proceedings of the National Academy of Sciences. 1992;89(7):2950-54. https://doi.org/10.1073/ pnas.89.7.2950
- Saraiva M, O'garra A. The regulation of IL-10 production by immune cells. Nature Reviews immunology. 2010;10(3):170-81. https://doi.org/10.1038/nri2711
- Liao W, Lin JX, Leonard WJ. IL-2 family cytokines: New insights into the complex roles of IL-2 as a broad regulator of T helper cell differentiation. Current Opinion in Immunology. 2011;23 (5):598-604. https://doi.org/10.1016/j.coi.2011.08.003
- 29. Marefati N, Ghorani V, Shakeri F, Boskabady M, Kianian F, Rezaee R, Boskabady MH. A review of anti-inflammatory, antioxidant and immunomodulatory effects of Allium cepa and its main constituents. Pharmaceutical Biology. 2021;59(1):285-300. https://doi.org/10.1080/13880209.2021.1874028
- Cole KE, Strick CA, Paradis TJ, Ogborne KT, Loetscher M, Gladue RP, Lin W, Boyd JG, Moser B, Wood DE, Sahagan BG. Interferoninducible T cell alpha chemoattractant (I-TAC): A novel Non-ELR CXC Chemokine with potent activity on activated T cells through selective high affinity binding to CXCR3. The Journal of Experimental Medicine. 1998;187(12):2009-21. https:// doi.org/10.1084/jem.187.12.2009
- Eftekhar N, Moghimi A, Mohammadian Roshan N, Saadat S, Boskabady MH. Immunomodulatory and anti-inflammatory effects of hydro-ethanolic extract of *Ocimum basilicum* leaves and its effect on lung pathological changes in an ovalbumininduced rat model of asthma. BMC Complementary and Alternative Medicine. 2019;19(1):1-11. https://doi.org/10.1186/s12906-019-2765-4
- Bisht D, Rashid M, Arya RK, Kumar D, Chaudhary SK, Rana VS, Sethiya NK. Revisiting liquorice (*Glycyrrhiza glabra* L.) as antiinflammatory, antivirals and immunomodulators: Potential pharmacological applications with mechanistic insight. Phytomedicine Plus. 2022;2(1):100206. https://doi.org/10.1016/ j.phyplu.2021.100206
- Van Lint P, Libert C. Chemokine and cytokine processing by matrix metalloproteinases and its effect on leukocyte migration and inflammation. Journal of Leucocyte Biology. 2007;82 (6):1375-81. https://doi.org/10.1189/jlb.0607338
- Romer LH, McLean NV, Yan HC, Daise M, Sun J, DeLisser HM. IFNgamma and TNF-alpha induce redistribution of PECAM-1 (CD31) on human endothelial cells. Journal of Immunology (Baltimore, Md.: 1950). 1995;154(12):6582-92. https://doi.org/10.4049/ jimmunol.154.12.6582