



REVIEW ARTICLE

Andean tubers: Traditional medicine and other applications

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Abstract

Tubers are a staple food in Andean highland communities, valued for their rich carbohydrate content with starch being the predominant component. Additionally their consumption is preferred for various health benefits, including anti-inflammatory effects, pain relief, digestive system improvement, diabetes management and wound healing, among others. Several studies have focused on Andean tubers such as *Oxalis tuberosa*, *Tropaeolum tuberosum*, *Ullucus tuberosus*, *Arracacia xanthorrhiza* and *Smallanthus sonchifolius*, revealing different compounds with pharmacological effects on humans and strengthening the immune system. Therefore, the objective of this review was to compile information from different databases such as Scopus and Web of Science on research conducted in the last decade on these tubers, highlighting their health benefits and potential for technological innovation for the benefit of the Andean communities that produce these crops. These tubers have been little marketed outside their production areas due to a lack of awareness. Although their applications have been limited, they have shown positive results, making them promising sources of health-beneficial phytochemicals. They could be used in technological developments to benefit health, particularly in addressing diseases that cause the most deaths.

Keywords

Andean tubers; *Arracacia xanthorrhiza*; *Oxalis tuberosa*; physicochemical; *Smallanthus sonchifolius*; *Tropaeolum tuberosum*; *Ullucus tuberosus*

Introduction

For over 3000 years, Andean tubers have been a staple food in the diet of the Inca, Quechua and Aymara peoples, serving as important sources of energy, vitamins and minerals. These tubers are characterized by their high productivity, due to their wide adaptability to diverse climatic and orographic conditions, such as altitudes ranging from 1800 to 3800 m above sea level, extreme daily temperature variations and low rainfall (300 mm per year). This resilience allows them to be cultivated without using agricultural inputs such as pesticides and fertilizers (Table 1) (3). In the Andes, the main tubers include Oca (*Oxalis tuberosa*), Mashua (*Tropaeolum tuberosum*), Papalisa (*Ullucus tuberosus*), Potato (*Solanum* ssp. L.), Arracacha (*Arracacia xanthorrhiza*) and Yacon (*Smallanthus sonchifolius*). Among these tubers, Potato is the most well-known and widely cultivated, followed by the Oca, which is the only tuber that has been successfully cultivated outside the Andes,

found in Mexico and New Zealand. The other tubers remain endemic to the Andean highlands (4).

Table 1. Agronomic performance of Andean tubers.

Tuber	Sowing density	Yield kg ha ⁻¹	Reference
<i>Oxalis tuberosa</i>	25000 – 50000 plants ha ⁻¹	5080 - 5223	(1)
<i>Tropaeolum tuberosum</i>	> 30000 plants ha ⁻¹	7000 - 7500	(1)
<i>Ullucus tuberosus</i>	35700 – 41600 plants ha ⁻¹	2000 - 10000	(1)
<i>Arracacia xanthorrhiza</i>	50000-125000 plants ha ⁻¹	- 10000	(1)
<i>Smallanthus sonchifolius</i>	12000-26000 plants ha ⁻¹	20000 - 50000	(2)

The Andean highlands, as depicted in Fig. 1, span across 7 countries: Venezuela, Colombia, Peru, Ecuador, Bolivia, Argentina and Chile. The Andes form the second-longest mountain range globally, stretching approximately 7000 km, surpassed only by the Himalayas. Peru, Argentina and Ecuador host the highest peaks of this mountain chain. Additionally, the region contains around 183 volcanoes, numerous hot springs and mineral deposits (5, 6). The vegetation at the highest latitude is found in this mountainous region. Rainfall is frequent up to the forest limit (3000 m above sea level), with temperatures ranging approximately from -3 to 7 °C and an average altitude ranging from 3000 to 4500 m above sea level. This ecological zone is referred to as the "altiandino" zone, marking the limit of vegetation (7).



Fig. 1. Map of the Andes region.

These tubers are traditionally consumed by Andean communities and have been used to alleviate digestive problems, inflammation, pain, kidney damage and more. People report feeling better after consuming these tubers in their usual forms, such as infusions, soups, purees, drinks etc. There are also few scientific "in vivo" studies

that demonstrate why these tubers produce positive health effects, suggesting that the presence of secondary

metabolites might be responsible. A previous study has evaluated the biological activity of Oca and Yacon on the intestinal health of a group of rats (8) and obtained good results, probably due to the presence of anthocyanins and fructo-oligosaccharides respectively. Similarly, researchers have investigated the anti-inflammatory activity of alkalamides, bioactive compounds in yacon and found a positive anti-inflammatory response on TNF-α and NF-κB (9).

However, the lack of knowledge about the primary and secondary metabolites of Andean tubers (some of these metabolites have been associated with antibiotic, antioxidant, insecticidal, nematocidal, anticancer and diuretic properties) has limited the enhancement of their properties in scientific and technological developments. This limitation hinders the ability to add value to these tubers and thereby support the economic sustainability of Andean communities (10, 11). Based on the above, this review analyses the contributions and systematic information documented in bibliographic databases such as Scopus and Web of Science on the uses of Andean tubers in traditional medicine, their physicochemical composition and recent developments or applications in health. The focus is on 5 Andean Tubers: Oca (*Oxalis tuberosa*), Mashua (*Tropaeolum tuberosum*), Papalisa (*Ullucus tuberosus*), Arracacha (*Arracacia xanthorrhiza*) and Yacon (*Smallanthus sonchifolius*).

Materials and Methods

The methodology was based on a systematic and exhaustive review of the scientific literature available in the mentioned databases. Keywords such as tubers, Andeans, *Oxalis tuberosa*, *Tropaeolum tuberosum*, *Ullucus tuberosus*, *Arracacia xanthorrhiza*, *Smallanthus sonchifolius*, phytochemicals and medicinal use were used to ensure the relevance and quality of the selected studies, resulting in 497 articles (Fig. 2). Documents with a publication date before 2003, those repeated in both databases and those not primarily focused on Andean tubers and their technological developments were excluded.

In addition, a detailed analysis of the selected studies was conducted, extracting relevant information on the traditional medicinal uses of Andean tubers, their physicochemical composition and recent advances and applications in health. Of the initial selection, 68 articles were analyzed. The synthesis of the information was carried out systematically, reviewing the summary, title, analysis of results and conclusions to ensure high relevance.

This allowed the findings to be organized into thematic categories to facilitate the presentation and understanding of the results.

noids or phytosterols, vitamins, minerals, glucosinolates and fatty acids. These important biochemical compounds have gradually evolved to protect and promote plant

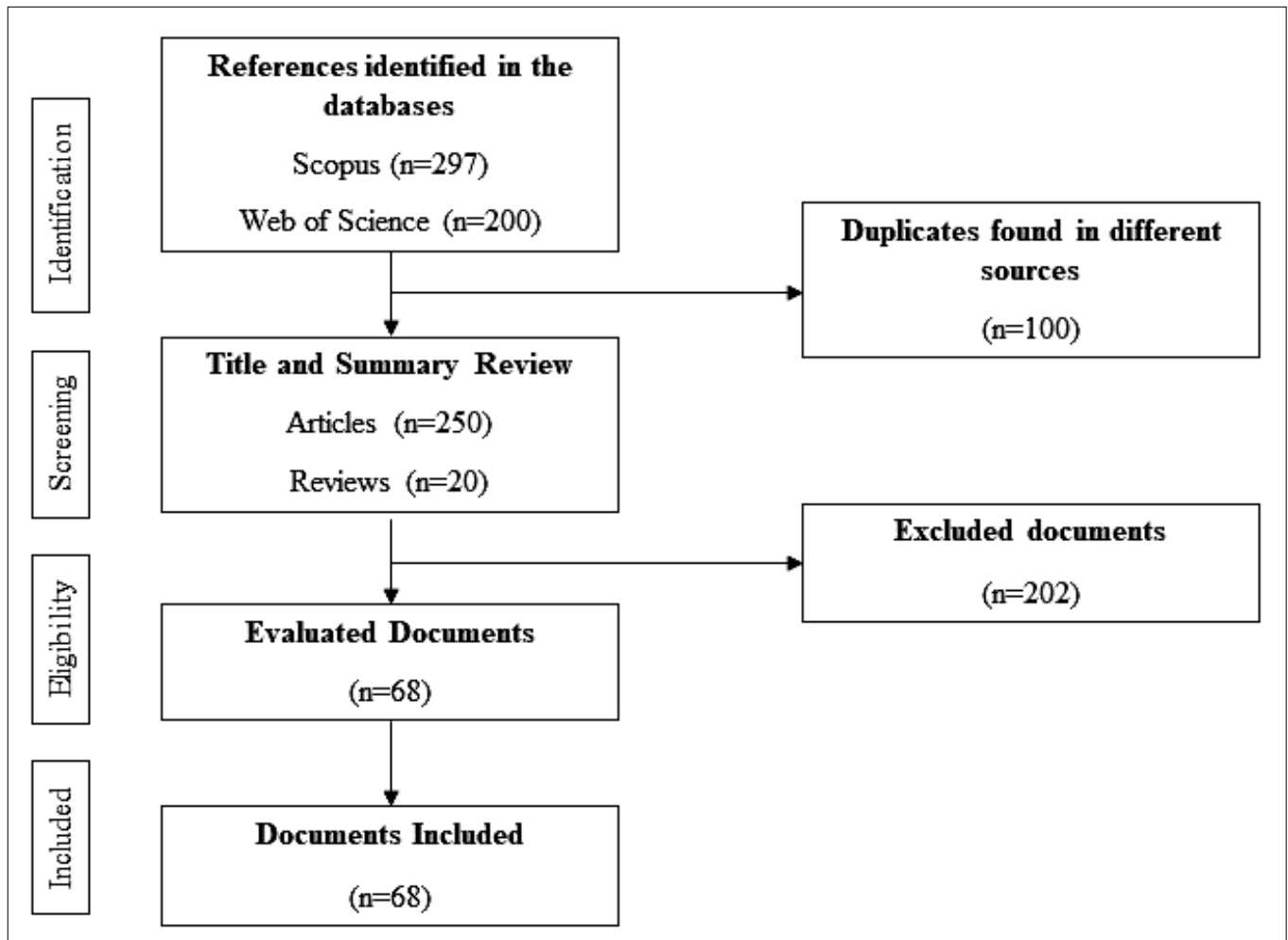


Fig. 2. The methodological process used for the review of Andean tubers.

Results and Discussion

Uses in traditional medicine

Table 2 reports the common names of Andean tubers, their forms of consumption and their applications in traditional medicine by different indigenous communities. These communities have consumed the tubers in different ways according to their traditions and for their health benefits, possibly due to their composition. Each tuber contains secondary metabolites with biological and antioxidant properties that are of interest to the scientific, food and pharmaceutical fields. These properties present opportunities to develop various alternatives and to use resources that have been produced for years but are gradually being lost due to limited knowledge about them. Thus, few scientific studies have demonstrated *in vivo* how the composition of these tubers helps to prevent and/or reduce diseases that can cause death.

Physico-chemical composition

The analysis of the chemical composition of a plant matrix reveals the substances present and their proportions, including proteins, fats, vitamins, minerals, carbohydrates, toxins and bioactive compounds. The latter include primary and secondary metabolites such as polyphenols, carote-

growth and reproduction and have been characterized as being of agroindustrial and food interest (32).

Table 3 describes some secondary metabolites, highlighting their biological activities. For instance, polyphenols accumulate in the outer layers of plant tissues and seeds as a protective mechanism against environmental stressors. These compounds are recognized for their antioxidant activity and various biological benefits, which help protect against oxidative reactions. Consequently, they have garnered attention across scientific areas such as biochemistry, medicine and pharmaceuticals (50). Similarly, fructo-oligosaccharides have been integrated into the food industry because they provide texture to dairy products and moisture to soft baked goods like sucrose, but with fewer calories. In addition to their nutritional properties, they exhibit bioactive characteristics such as prebiotic, hypocholesterolaemic and hypolipidemic properties. These attributes suggest they can reduce the risk of colon cancer, improve cognitive function, have brain-protective effects and reduce blood glucose, phospholipids, serum cholesterol and triglycerides (51). Carotenoids and glucosinolates are also known to have positive health effects due to their biological properties.

Table 2. Medicinal uses and consumption methods of Andean tubers.

Tuber	Common name	Consumption method	Traditional Medicinal Use	Traditional medicine preparation mode	Reference
<i>Oxalis tuberosa</i>	Papa de Oca, Oca, Cuiba, Quiba, Ibia, Apilla, Uncha, Papa extranjera, Agrillo, Oca de la chacra, Oca Rosada, Papa roja.	Soups, stews, sweets, seasoning.	Prostate inflammation, inflammatory diseases, diuretics, gallbladder damage, gastritis, rheumatism, sexual potency, diarrhea, constipation, skin ulcers, postpartum recovery and acne treatment.	Juices, Syrup, Infusions, Tea.	(12-17)
<i>Tropaeolum tuberosum</i>	Mashua, Isaño, Año, Año, Cubio	Fresh, boiled, fried, baked, stewed, soups, flour.	Treating kidney diseases, bladder and kidney stones, skin ulcers, diuretics, and prostate inflammation.	Juices, Syrup, Infusions, Tea	(4, 18-20)
<i>Ullucus tuberosus</i>	Ulluco, Papa Lisa, Lisa, Melloco, Olluco, Chugua, Ruba	Fresh, cooked, soups, salads.	Treating burns, scar prevention.	Juices, Syrup, Infusions, Tea	(21-24)
<i>Arracacia xanthorrhiza</i>	Arracacha, Zanahoria Peruana, Chirivía, Zanahoria Blanca	Soups, bread, frying, purees, cakes, flour, sweets, roasted.	Digestive system issues, gastritis.	Juices, Syrup, Infusions, Tea	(25-27)
<i>Smallanthus sonchifolius</i>	Yacon	Infusion, juices, puree, sweets, jams, syrup, flour, cookies, smoothies, wines.	Obesity, diabetes, colon cancer, kidney conditions, skin rejuvenation.	Juices, Syrup, Infusions, Tea	(29-31)

Table 3. Characteristics of secondary metabolites found in plant matrices.

Secondary metabolites	Type	Characteristics	
Polyphenols	Phenolic acid	Modulate the digestion of food and the absorption of nutrients, due to its ability to unleash and inhibit enzymes of digestion and intestinal transport (33).	<i>O. tuberosa</i> (13) <i>Rhizophora mucronate</i> (41) <i>Dioscorea</i> spp (42)
	Flavonoid	Antioxidant, anti-inflammatory, antitumoral, cardiovascular protection, antidiabetic and immunomodulatory activities. It exerts its antidiabetic effect by stimulating the secretion of insulin, inhibiting the absorption of glucose, reducing the production of free radicals and improving antioxidant activity (34).	<i>O. tuberosa</i> (13) <i>Sedum rubens</i> (43)
	Lignan	Exhibits pharmacological effects such as hepatoprotective, anti-inflammatory, antioxidant, antitumor and antipathological effects (35).	<i>Linum usitatissimum</i> (44)
	Estilbenes	Provides benefits for human health, such as protection against cancer, inflammation, cardiovascular diseases and diabetes, thanks to resveratrol as a fundamental component (36).	
Carotenoids	Carotene	Natural coloring with antioxidant activities and provitamin A, which is susceptible to oxidation and easy isomerization under conditions of light, heat or acids. Therefore, it is not directly used in food and medicine (37).	<i>A. xanthorrhiza</i> (26) Sweet potato (45)
	Xanthophylls	Lutein, zeaxanthin and β -cryptoxanthin are xanthophylls that are important for brain and eye development and for stimulating the immune and antioxidant systems (38).	<i>Zea mays</i> (46)
Glucosinolates		The properties of this substance include antioxidant and immune-strengthening effects. It also inhibits the production of carcinogens, which can cause damage to DNA and cells, leading to tumor growth (39).	<i>Tropaeolum tuberosum</i> (47) <i>Raphanus sativus</i> (48)
Fructooligosaccharides		The non-digestible bifidogenic oligosaccharide is fermented by intestinal microbes that produce short-chain fatty acids, hydrogen gas derived from intestinal microbes and other metabolites. It does not increase blood glucose levels, stimulates insulin secretion, suppresses adipose tissue accumulation and has a beneficial effect on glucose homeostasis (40).	<i>Smallanthus sonchifolius</i> (47) <i>Allium cepa</i> (49)

Oxalis tuberosa

Ibia (*Oxalis tuberosa*), also known as Oca, is a tuber with a color range from white/cream, yellow, orange, pink, red and violet, with firm white to peach flesh and buds all over its surface. Its average size is 66.22 mm long and 18.04 mm in diameter (12, 51) (Fig. 3). Its potential uses include weed control and soil fertility enhancement. Its nutritional composition is approximately 1.1-9 % protein, 13.2 % carbohydrates, with starch (60-50 %), sugars such as sucrose and glucose (21 % and 3.6 % respectively), 0.6 % fat, 1-5.1 % fiber, minerals and organic acids as major components, thus allowing its use in industrial processes such as baking, dehydration and alcohol extraction by fermentation (35). The *in vitro* digestibility of starch has been reported to be around 91.78 %. Additionally, its phytochemical composition includes anthocyanins such as 3, 5-O-diglucosides of petunidin, peonidin, malvidin, 3-O-diglucosides of delphinidin, petunidin as well as 3, 5-O-diglucoside of malvidin (13, 52, 53).

fiber and some secondary compounds such as polyphenols, anthocyanins, carotenoids and glucosinolates (GLS), with glucoabrietin being found in the highest proportion (96–99 %) (54). These compounds are present in higher content compared to other vegetables such as cauliflower, broccoli and maca, with concentrations reported from 0.27 to 54.2 $\mu\text{mol/g}$, depending on the cultivar, post-harvest time and cultivation conditions (55). Products of GLS hydrolysis, including isothiocyanates, thiocyanates, thiones, epithionitriles and nitriles possess anticancer and anti-inflammatory activity. Phenethyl isothiocyanate in particular, has been reported to have an anti-tumor effect, suppressing the migration and invasion of lung cancer cells. Therefore, the presence of GLS in this species may be responsible for its medicinal uses (56, 47).

Ullucus tuberosus

Ullucus tuberosus, commonly known as Ulluco, is a tuber with periderm that varies in color from yellow, orange, red, magenta to purple, while its pulp is white or yellow.

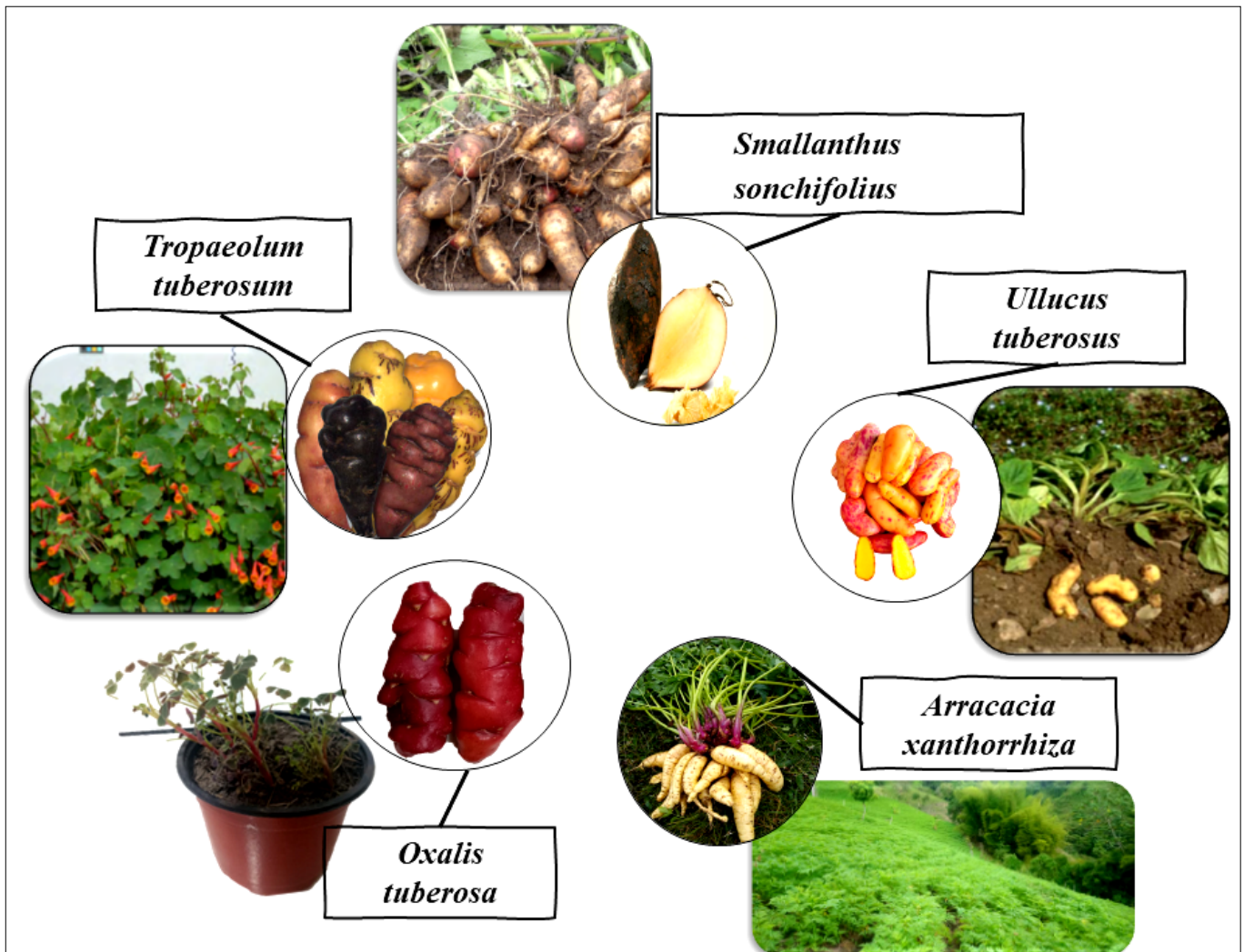


Fig. 3. Some examples of Andean tubers (source: 87-90).

Tropaeolum tuberosum

Tropaeolum tuberosum, commonly known as Mashua, is a tuber from Andean highlands. Its skin colors range from white or yellow to red or purple (Fig. 3). It can measure between 5-15 cm long and 3-6 cm wide at its distal or apical end (10). It contains approximately 35 % fresh starch,

It typically measures about 1 cm in diameter and up to 6 cm in length on average (21, 57) (Fig. 3). Nutritionally, it is considered a good source of carbohydrates, with approximately 65 % of its dry weight corresponding to starch. It also contains proteins, fiber, vitamin C and betalains. Betalains are associated with antioxidant, anti-inflammatory,

antimalarial and antitumor activities and their consumption have also been recommended for possible protection against stress-related conditions and the few sources of betalains in the daily diet (58). Ulluco is consumed fresh, although applications have also been developed for starch extraction for industrial purposes (59).

Arracacia xanthorrhiza

Commonly known as Arracacha, this plant is native to South America and is cultivated mainly in Brazil, Colombia, Venezuela, Ecuador, Peru and Bolivia. Its tubers vary in color from white to yellow and purple, measuring up to 4 cm in thickness and 20 cm in length (Fig. 3). Nutritionally, it is considered a rich source of carbohydrates, providing 25 g/100 g, with approximately 80 % of this content being starch. The yellow tubers, due to their color, contain carotenoids which are precursors of vitamin A. Additionally, Arracacha contains niacin and minerals such as calcium, iron and phosphorus making it a potential nutraceutical food suitable for children, the elderly or convalescent individuals. It has a slightly sweet nutty flavor and is highly digestible (25-27).

Smallanthus sonchifolius

The Andean crop commonly known as Yacon has a skin that varies from tan to dark brown and its pulp can range from white and yellow to purple and orange. The tuber can grow up to 40 cm in length and weigh up to 2 kg (28) (Fig. 3). Yacon has a sweet taste because it is a rich source of β -(2 \rightarrow 1) fructo-oligosaccharides (FOS), ranging from 6.4-65 g/100 g dry matter. These include ketose, nystose and fructofuranosyl nystose. It also contains phenolic compounds, mainly chlorogenic acid, ranging from 7.9 - 30.8 mg/g dry matter and other derivatives of caffeic acid. These compounds provide several health benefits, such as prebiotic, anti-diabetic, antioxidant and antimicrobial effects (30, 60). Since the fructan in Yacon are not hydrolyzed by human digestive enzymes, they are considered as dietary fibers that pass through the gastrointestinal tract unchanged, attributing immunomodulatory properties to Yacon by increasing the production of short-chain fatty acids (61).

Technological developments and applications

Oxalis tuberosa

A snack based on maize semolina with sweet Potato, mashua and three varieties of goose flour was developed to promote its consumption throughout the year, not just during the production season (62). Textural analysis showed a relationship between porosity and hardness and a change in optical properties, possibly due to the loss of reducing sugars during Maillard reactions with proteins. The extrusion process reduced the water content of the mixture by approximately 5.6 %. The snacks exhibited high hygroscopicity, which posed a challenge for packaging and marketing the products. Additionally, the antioxidant capacity of *O. tuberosa* juice and *Gaultheria glomerata* (Cav.) fruit in a functional beverage has been determined (63). It was observed that a mixture of 300 mL of goose juice, 800 mL of Gaultheria juice, 1000 mL of treated water and

220 g of refined sugar provides 40.13 kcal/100 g of total energy. The protein content was 1.38 %, fat 1.08 %, moisture 99.5 %, ash 1.82 % and carbohydrate content 6.22 %. The antioxidant profile showed a total polyphenol content of 1.825 g gallic acid (GA)/100 g and antioxidant activity of 89.56 μ mol of Trolox/100 g. Therefore, the drink made from these 2 species is a viable nutritional option for people who engage in rigorous and frequent physical activity.

By adding 6.6 % and 13.2 % red Potato flour (*O. tuberosa*) (PR) to gluten-free English bread, the texture of the product improved without affecting its physical characteristics (64). Additionally, the dough became more extensible and less hard and the volume increased. The crumb was more uniform and the protein content increased to 5 %. The inclusion of red Potato flour (*O. tuberosa*) in extruded products made from corn semolina resulted in a decrease in fat content from 8.86 % to 5.60 %, a decrease in the glycaemic index compared to the control and an increase in ash content from 1.81 % to 2.30 %.

Similarly, in extruded products made from corn semolina and red Potato flour (HPR), the inclusion of HPR altered the water absorption index values (3 %). The extrudates' water solubility index increased from 6.18 % to 20.19 % and their fracturability increased from 201.58 N to 495.68 N (65) when the HPR was used at 100 %. The luminosity of the extrudates decreased from 44.37 % to 25.06 %. Additionally, the extrudates showed a higher total phenol content of 2.88 mg AG/g dry sample compared to the control's 0.44 mg AG/g dry sample. Therefore, the use of HPR allows for the development of extrudates with favorable physicochemical characteristics and antioxidant properties. In an extrudate obtained from a mixture of Oca (*O. tuberosa*) flour (HO) and oats, it was observed that increasing the fiber content decreased the glycaemic index to 65.04 % and the starch hydrolysis percentage to 67.29 %, thus enabling a controlled release of sugar into the bloodstream (66). Moreover, the inclusion of HO in the extrudates increased their antioxidant activity. Therefore, this food can be considered functional due to its health benefits.

It has been investigated that prebiotic activity exhibited by three Andean tubers, Oca, Papalisa and Potato, as fermentation substrates for *Lactobacillus brevis* strain CJ25, which is used in the production of Potato Cheese (67). This study noted a pH reduction by 0.7 units, enhancing food safety by inhibiting the growth of pathogenic microorganisms. Moreover, *L. brevis* CJ25 demonstrated survival under simulated conditions of 3 h at pH 2.5 and resistance to bile acids. Hence, fostering such fermentations could positively impact the Andean regional economy and the community health. Furthermore, research indicates that Wistar rats fed with Yacon and Oca experienced an increase in *Lacto bacilli* and Bifidobacteria, correlating with a decrease in pH from 7.62 to 6. The antioxidant activity in the cecum of rats fed with Andean tubers increased at about 33 days compared to the control diet. The total content of phenolic compounds was about 137.82 mg AG/100 g tuber. These findings suggest a beneficial effect of Yacon and Oca diets on the intestinal health (11).

Tropaeolum tuberosum

In evaluating the biotransformation of GLS present in mashua using lactic acid bacteria and its *in vivo* absorption in rats, it was found that the bacteria utilize all the GLS. In rats, the GLS were absorbed and metabolized, with low contents detected in stool (0.02 %) and urine (0.59 %) and they were present in plasma up to 3 h after consumption (56). This suggests that *T. tuberosum* is a beneficial product for human health.

Ethanol extracts of *Moringa oleifera*, *T. tuberosum* and *Annona cherimola* have demonstrated antioxidant and antiproliferative activity against colorectal cancer (CRC) cell cultures grown as monolayers. These extracts inhibited colon cancer cells and showed synergistic activity with the chemotherapeutic agent 5-FU, providing new therapeutic options for the prevention and treatment of colorectal cancer (68). When two alkaloids identified in *T. tuberosum* were evaluated in human cancer cell lines, slight cytotoxic activity was observed with IC₅₀ values between 27.45 ± 0.80 µM and 31.07 ± 0.87 µM. Significant anticancer potential was observed with IC₅₀ values between 1.26 ± 0.57 µM and 1.37 ± 0.09 µM, which led to an increase in the rate of apoptosis of these cells (69). Additionally, isolated macamides from *T. tuberosum* exhibited anti-glycation and anti-inflammatory effects. In skin cells, these compounds inhibited the production of TNF-α with IC₅₀ values of 9.38-10.06 µM and the formation of BSA-MGO at concentrations of 9.38-5.30 µM. Significant cross-link cleavage was also observed, with modification rates ranging from 6.58 to 18.8 % (70). Furthermore, isolated alkaloids from this tubers showed anti-TNF-α activity with IC₅₀ values of 1.56-3.12 µM and suppressed NF-κB with IC₅₀ values of 0.02-3.54 µM. These compounds can be used for the development of new synthetic analogs as anti-inflammatory agents (11).

Ullucus tuberosus

The color stability of Ulluco extracts has been studied to utilize the betalains present in this tuber as a natural colorant. The findings indicate that Ulluco extract is effective as a colorant across a wide pH range (21). Specifically, when added to highly acidic foods, the color intensity of Ulluco extract was lower but remained more stable over time. At pH 4, the extract exhibited a significantly lower color intensity (C * ab ~ 40), but the stability of the red tone (hab) over time was higher, making betalains from Ulluco appealing to the food industry for applications requiring long-term color stability. By understanding the color variations, it is possible to obtain a specific color for a product. Moreover, when Ulluco betalains were added to foods with a pH between 5 and 6, the antioxidant capacity increased by 1.3 times compared to those with a pH of 4. This antioxidant capacity was further enhanced when the food is subjected to a cooking process at 80 °C. Additionally, storing the food at 4 °C preserved the maximum possible amount of initial betalains and maintained their antioxidant capacity (71). This research suggests that Ulluco betalains are not only valuable for their coloring properties but also for their potential health benefits, making them an attractive natural additive in food products.

Interesting results have been reported regarding the use of aqueous extracts of *U. tuberosus* in wound healing, as shown in *in vitro* models. Furthermore, levels of cell proliferation were increased. These extracts were found to be non-toxic to fibroblasts even at high concentrations. Additionally, collagenase activity increased by 12 %, suggesting that it could be an effective agent for treating scars. Enhanced migration and procollagen production facilitated tissue regeneration without scarring, while an increase in MMP-1 promoted wound healing during the phases of cellular remodeling and differentiation (22).

The consumption of this tuber can be incorporated into low-fat yogurt as Ulluco flour (FU), which fortifies the yogurt due to its starch content and enhances stability and viscosity. With the addition of FU to the yogurt, acid production during fermentation increased, achieving the final pH of 4.5 in 5.5 h compared to 7 h for the control. After 20 days of storage, the yogurt exhibited pseudoplastic rheological behaviour and sensory attributes showed greater acceptability with 0.9 % FU. This suggests that FU can be considered a valuable gelling and nutritional agent (72).

Arracacia xanthorrhiza

Instant powdered drinks have emerged as an alternative for those who require quick and easy food consumption. Therefore, extruded flours from roots and tubers were used in the development of this type of product, as the starch present in these cultures has a higher content of amylopectin than amylose. Thus, obtaining suspensions with improved thickening and stabilizing characteristics as well as viscoelastic fluid consistency and stability over a wide pH range, is ideal for the development of instant beverages (73). An instant dairy drink has been formulated using extruded Arracacha flour and the addition of folic acid. The resulting product has good sensory acceptance and is characterized by a high content of protein (13.33 %), soybean (10.94 %), almonds (22.02 %) and dietary fiber (3.12 %). The starch digestibility rate was 43.61 ± 0.12 %, progressively increasing to an average value of 56.80 ± 0.11 % at 60 min, indicating intermediate digestion of the mixture (74). When evaluating the effect of yellow Arracacha on an experimental model of ovarian polycystic disease, it was found that it regulated sexual hormone function, including follicle-stimulating hormone (FSH), estradiol, luteinizing hormone (LH) and testosterone due to the presence of polyphenols in its composition (75).

Smalanthus sonchifolius

The prebiotic effect of Yacon has been reported in a guinea pig model (30). The study observed FOS contents ranging from 6.4 to 65 g/100 g dry matter, total phenolic compounds from 7.9 to 30.8 mg of chlorogenic acid and antioxidant capacity between 23 to 136 µmol Trolox equivalent/g dry matter. In addition, the growth of bifidobacteria and *Lacto bacilli* was promoted and high levels of short-chain fatty acids were generated in the cecal material. This resulted in improved cell density and crypt formation in the tissue of the cecum, which are beneficial effects for colon health. The identified FOS were AJC 5189, AKW 5075 and

AMM 5163, which may be of interest to the nutraceutical industry.

During the production of Yacon syrup, positive effects on postprandial glucose and insulin response have been observed through various mechanisms. These include increased viscosity and gel formation in the intestine, reduced glucose diffusion, altered intestinal transit time and prolonged carbohydrate absorption (76). The hypoglycaemic activity of *S. sonchifolius* was evaluated in rodents with streptozotocin-induced diabetes, revealing a reduction in glucose levels and an increase in plasma insulin levels following administration. Additionally, its relatively low-calorie content has been investigated for its antidiabetic activity. Tests of α -amylase and α -glucosidase activity inhibition yielded positive results, suggesting that the bioactive compounds present in this tuber may be useful in treating this disease (29).

Yacon has been associated with positive effects on intestinal and bone health. Studies on rats have reported that Yacon, either alone or in combination with kefir, reduced intraluminal pH, improved intestinal permeability (by decreasing the excretion of lactulose and mannitol) and increased the balance of calcium and osteocalcin, a biomarker of bone formation (77). Similarly, positive effects of Yacon flour (YF) on oxidative stress, inflammation and endotoxemia have been reported in rats with induced colorectal cancer (CRC), where YF increased levels of fecal secretory immunoglobulin A and decreased lipopolysaccharides, tumor necrosis factor alpha and interleukin-12. Additionally, the production of short-chain fatty acids acetate, propionate and butyrate showed interactions with nuclear factor kappa B (NF- κ B), Toll-like receptor 4 (TLR4) and NADPH oxidase through *in silico* analysis. Therefore, treatment with YF could be a promising food to reduce inflammation caused by CCR (78).

Significant improvements in growth performance were observed when black rockfish were fed with Yacon juice, including increased weight gain, specific growth rate, feed intake, feed efficiency and protein efficiency ratio. Activities of plasma digestive enzymes (amylase, trypsin and lipase), lysozyme and antioxidant enzymes (superoxide dismutase, catalase and glutathione) were mostly higher in the groups of fish fed with Yacon juice (79). Similarly, the potential of Yacon to prevent diabetic nephropathy has been evaluated through anti-inflammatory, antioxidant and anti-fibrotic mechanisms. It has also been shown to reduce the proliferation of mesangial cells and levels of transforming growth factor beta-1 (TGF- β 1). In one study, the effects of different concentrations of Yacon (5, 10 and 50 μ g/mL) on cellular models of diabetic nephropathy were examined. Results indicated that Yacon could decrease the gene expressions of SMAD2, SMAD3 and SMAD4 proteins while increasing the expression of SMAD7. This suggests that Yacon has therapeutic potential against cancer (80).

Table 4 shows some of the commercial products developed from these Andean tubers since the Andean associations began an entrepreneurial journey to initiate

the trade of their products and look for economic alternatives.

Table 4. Prepared products from Andean tubers.

Tuber	Products	Reference
<i>Oxalis tuberosa</i>	-	
<i>Tropaeolum tuberosum</i>	Milk drink	(81)
<i>Ullucus tuberosus</i>	Whole Ullucus in brine	(82)
<i>Arracacia xanthorrhiza</i>	Flour	(83)
<i>Smallanthus sonchifolius</i>	Tea bags, dietary supplements, honey, chips, flour	(84-86)

Conclusion

The Andean tubers *Oxalis tuberosa*, *Tropaeolum tuberosum*, *Ullucus tuberosus*, *Arracacia xanthorrhiza* and *Smallanthus sonchifolius* contain various phytochemicals of interest with the potential to be employed in the development of products and processes that will generate added value, thereby contributing to the economic sustainability of the indigenous communities where these crops are grown. Despite limited commercial applications, the existing scientific research indicates positive health effects from these tubers, largely due to their antioxidant capacity and nutritional composition. This underscores the need for further research. Currently, trade in these tubers is restricted and their economic return is low because their potential benefits are not widely known. Developing products and processes based on these Andean tubers could not only provide economic benefits but also can promote the conservation of biodiversity and preserve the cultural heritage of the associated indigenous communities.

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Authors' contributions

ARCL is the main author who conducted the research work, with the assistance and guidance of JFSD, JAAV, MRSS, ACFG, SDNF, SYSB and RRH, to carry out the design, data acquisition and proper writing of the manuscript. All authors read and approved the final manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interests to declare.

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References

- Sanchez-Portillo S, Salazar-Sánchez MDR, Solanilla-Duque JF, Rodríguez-Herrera R. Andean tubers, morphological diversity and agronomic management: A review. *Plant Sci Today*. 2023;10(sp2):98-105.

2. Manrique I, Gonzales R, Valladolid A, Blas R, Lizárraga L. Producción de semillas en yacón (*Smallanthus sonchifolius* (Poepp. & Endl.) mediante técnicas de polinización controladas. *Ecología Aplicada*. 2014;13(2):135-45. <https://doi.org/10.21704/rea.v13i1-2.464>
3. Goldner MC, Pérez OE, Pilosof AMR, Armada M. Comparative study of sensory and instrumental characteristics of texture and color of boiled under-exploited Andean tubers. *LWT*. 2012;47(1):83-90. <https://doi.org/10.1016/j.lwt.2011.12.031>
4. Condori B, Mamani P, Botello R, Patiño F, Devaux A, Ledent JF. Agrophysiological characterization and parametrisation of Andean tubers: Potato (*Solanum* sp.), Oca (*Oxalis tuberosa*), isaño (*Tropaeolum tuberosum*) and Papalisa (*Ullucus tuberosus*). *European Journal of Agronomy*. 2008;28(4):526-40. <https://doi.org/10.1016/j.eja.2007.12.002>
5. Visscher AM, Vanek S, Huaraca J, Mendoza J, Ccanto R, Meza K, et al. Traditional soil fertility management ameliorates climate change impacts on traditional Andean crops within smallholder farming systems. *Science of the Total Environment*. 2024 Feb 20;912. <https://doi.org/10.1016/j.scitotenv.2023.168725>
6. Carlsen M. Los Andes. In: *FundacionEmpresasPolar*, editor. GEOVENEZUELA. Caracas; 2012. p. 1-1.
7. Greenpeace. El rol de la Cordillera de Los Andes frente a las olas de calor: sus montañas reducen la temperatura [Internet]. En línea: 9 de Febrero del 2024 [Consultado el 23 de Mayo de 2024]. Disponible en: <https://www.greenpeace.org/argentina/blog/problemas/climayenergia/el-rol-de-la-cordillera-de-los-andes-frente-a-las-olas-de-calor-sus-montanas-reducen-la-temperatura/>
8. Jimenez ME, Rossi A, Sammán N. Health properties of Oca (*Oxalis tuberosa*) and Yacón (*Smallanthus sonchifolius*). *Food Funct*. 2015;6(10):3266-74. <https://doi.org/10.1039/c5fo00174a>
9. Apaza TL, Tena Pérez V, Șerban AM, Alonso Navarro MJ, Rumbero A. Alkamides from *Tropaeolum tuberosum* inhibit inflammatory response induced by TNF- α and NF- κ B. *J Ethnopharmacol*. 2019;235:199-205. <https://doi.org/10.1016/j.jep.2019.02.015>
10. Malpartida Yapias RJ, Adama Astete JM, Cajachagua Uscuchagua YY, Rosales Sánchez MC. Características fisicoquímicas, composición nutricional y compuestos bioactivos en tres variedades de Mashua (*Tropaeolum tuberosum* Ruiz y Pavón): Una revisión. *Revista Tecnológica - ESPOL*. 2022;34(2):40-50. <https://doi.org/10.37815/rte.v34n2.891>
11. Morillo AC, Morillo Y, Leguizamo MF. Diversidad genética de ibias (*Oxalis tuberosa* Molina) y cubios (*Tropaeolum tuberosum* Ruiz y Pavón) en Boyacá. *Temas Agrarios*. 2016;21(1):44-53. <https://doi.org/10.21897/rta.v21i1.86>
12. Paniagua-Zambrana NY, Bussmann RW, Romero C. *Oxalis tuberosa* Molina Oxalidaceae. In: *Etnobotánica de los Andes, Etnobotánica de las regiones montañosas*; 2020. pág. 1-7. https://doi.org/10.1007/978-3-319-77093-2_216-1
13. Campos D, Chirinos R, Gálvez Ranilla L, Pedreschi R. Bioactive potential of Andean fruits, seeds and tubers. *Adv Food Nutr Res*. 2018;84(1):287-343. <https://doi.org/10.1016/bs.afnr.2017.12.005>
14. Zhu F, Cui R. Comparison of physicochemical properties of oca (*Oxalis tuberosa*), potato and maize starches. *Int J Biol Macromol*. 2020;148:601-07. <https://doi.org/10.1016/j.ijbiomac.2020.01.028>
15. Chirinos R, Betalleluz-Pallardel I, Huamán A, Arbizu C, Pedreschi R, Campos D. HPLC-DAD characterization of phenolic compounds from Andean oca (*Oxalis tuberosa* Mol.) tubers and their contribution to the antioxidant capacity. *Food Chem*. 2009;113(4):1243-51. <https://doi.org/10.1016/j.foodchem.2008.08.015>
16. Bussmann RW, Paniagua Zambrana NY, Romero C, Hart RE. Astonishing the medicinal plant markets of Bogotá, Colombia. *J Ethnobiol Ethnomed*. 2018;14(43). <https://doi.org/10.1186/s13002-018-0241-8>
17. Morillo CAC, Morillo CY, Leguizamo MMF. Caracterización morfológica y molecular de *Oxalis tuberosa* Mol. en el departamento de Boyacá. *Rev Colomb Biotecnol*. 2019;21(1):18-28. <https://doi.org/10.15446/rev.colomb.biote.v21n1.57356>
18. Chirinos R, Campos D, Warnier M, Pedreschi R, Rees JF, Larondelle Y. Antioxidant properties of Mashua (*Tropaeolum tuberosum*) phenolic extracts against oxidative damage using biological *in vitro* assays. *Food Chem*. 2008;111(1):98-105. <https://doi.org/10.1016/j.foodchem.2008.03.038>
19. Chirinos R, Pedreschi R, Rogez H, Larondelle Y, Campos D. Phenolic compound contents and antioxidant activity in plants with nutritional and/or medicinal properties from the Peruvian Andean region. *Ind Crops Prod*. 2013;47:145-52. <https://doi.org/10.1016/j.indcrop.2013.02.025>
20. Aguilar-Galvez A, Pedreschi R, Carpentier S, Chirinos R, García-Ríos D, Campos D. Proteomic analysis of Mashua (*Tropaeolum tuberosum*) tubers subjected to postharvest treatments. *Food Chem*. 2020;305. <https://doi.org/10.1016/j.foodchem.2019.125485>
21. Cejudo-Bastante MJ, Hurtado N, Mosquera N, Heredia FJ. Potential use of new Colombian sources of betalains. Color stability of Ulluco (*Ullucus tuberosus*) extracts under different pH and thermal conditions. *Food Res Intern*. 2014;64:465-71. <https://doi.org/10.1016/j.foodres.2014.07.036>
22. Heil N, Bravo K, Montoya A, Robledo S, Osorio E. Wound healing activity of *Ullucus tuberosus*, an Andean tuber crop. *Asian Pac J Trop Biomed*. 2017;7(6):538-43. <https://doi.org/10.1016/j.apjtb.2017.05.007>
23. Pacheco MT, Moreno FJ, Moreno R, Villamiel M, Hernandez-Hernandez O. Morphological, technological and nutritional properties of flours and starches from Mashua (*Tropaeolum tuberosum*) and melloco (*Ullucus tuberosus*) cultivated in Ecuador. *Food Chem*. 2019;301. <https://doi.org/10.1016/j.foodchem.2019.125268>
24. Parra-Quijano M, Panda S, Rodríguez N, Torres E. Diversity of *Ullucus tuberosus* (Basellaceae) in the Colombian Andes and notes on Ulluco domestication based on morphological and molecular data. *Genet Resour Crop Evol*. 2012;59(1):49-66. <https://doi.org/10.1007/s10722-011-9667-8>
25. Londoño-Restrepo SM, Rincón-Londoño N, Contreras-Padilla M, Millan-Malo BM, Rodríguez-García ME. Morphological, structural, thermal, compositional, vibrational and pasting characterization of white, yellow and purple Arracacha Lego-like starches and flours (*Arracacia xanthorrhiza*). *Int J Biol Macromol*. 2018;113:1188-97. <https://doi.org/10.1016/j.ijbiomac.2018.03.021>
26. Quijano-Célis CE, Piedrahita D, Pino JA. Essential oil of *Arracacia xanthorrhiza* Bancr. leaves from Colombia. *Journal of Essential Oil-Bearing Plants*. 2016;19(5):1296-99. <https://doi.org/10.1080/0972060X.2016.1195710>
27. Albano KM, Franco CML, Telis VRN. Rheological behavior of Peruvian carrot starch gels as affected by temperature and concentration. *Food Hydrocoll*. 2014;40:30-43. <https://doi.org/10.1016/j.foodhyd.2014.02.003>
28. Campos D, Betalleluz-Pallardel I, Chirinos R, Aguilar-Galvez A, Noratto G, Pedreschi R. Prebiotic effects of Yacon (*Smallanthus sonchifolius* Poepp. & Endl), a source of fructooligosaccharides and phenolic compounds with antioxidant activity. *Food Chem*. 2012;135(3):1592-99. <https://doi.org/10.1016/j.foodchem.2012.05.088>
29. Peková L, Žiarovská J, Fernández-Cusimamani E. Medicinal plants with antidiabetic activity used in the traditional medicine

- in Bolivia: A review. *Bol Latinoam Caribe Plantas Med Aromat.* 2023;22(4):417-30. <https://doi.org/10.37360/blacpma.23.22.4.31>
30. Lachman J, Fernández EC, Orsák M. Yacon [*Smallanthus sonchifolia* (Poepp. et Endl.) H. Robinson] chemical composition and use - A review. *Plant Soil Environ.* 2003;49(6):283-90. <https://doi.org/10.17221/4126-PSE>
 31. Yin J, Wang Z, Ma G, Liu W. Complete chloroplast genome and phylogenetic analysis of *Smallanthus sonchifolius* (Asteraceae). *Mitochondrial DNA B Resour.* 2023;8(8):916-20. <https://doi.org/10.1080/23802359.2023.2248683>
 32. Silva ET de V, Queiroz AJM de, Figueirêdo RMF de, Moura HV, Santos FS dos, Silva AP de F, et al. Dynamic modelling of degradation kinetics of phenolic compounds, phenolic profiles, mineral content and overall antioxidant capacity of germinated peanut flours. *LWT.* 2023;183. <https://doi.org/10.1016/j.lwt.2023.114927>
 33. D'Costa A, Chen A, Hamann E, El Iraki R, Venugopal K, Bordenave N. Impact of potato starch on the inhibition of pancreatic lipase by potato phenolic acids. *Food Bios.* 2024;57. <https://doi.org/10.1016/j.fbio.2023.103414>
 34. Shen L, Li C, Wang W, Wang X, Tang D, Xiao F, Xia T. Buckwheat extracts rich in flavonoid aglycones and flavonoid glycosides significantly reduced blood glucose in diabetes mice. *Journal of Functional Foods.* 2024;113(48):106029. <https://doi.org/10.1016/j.jff.2024.106029>
 35. He D, Pei X, Liu B, Li J, Dong J, Efferth T, Ma P. Lignan contents of *Schisandra chinensis* (Turcz.) Baill. from different origins - A new model for evaluating the content of prominent components of Chinese herbs. *Phytomed.* 2024;128:155361. <https://doi.org/10.1016/j.phymed.2024.155361>
 36. Wang L, Zhang M, Qin L, Qian Y, Huang O, Zhang R, Duan D. VqNAC44 enhances stilbene synthesis and disease resistance in Chinese wild grape by interacting with VqMYB15. *Plant Science.* 2024;341. <https://doi.org/10.1016/j.plantsci.2024.111994>
 37. Hatami T, Jarles E, Araújo S, Baião A, Innocentini L, Martínez J. Mechanism of multicyclic β -carotene impregnation into corn starch aerogels via supercritical CO₂ with mathematical modeling. *Food Res Int.* 2024;178. <https://doi.org/10.1016/j.foodres.2024.114002>
 38. Ueno H, Sato T, Higurashi S, Tazaki H, Toba Y. Xanthophylls in human milk and maternal diet: A cross-sectional analysis of data from the Japanese human milk study cohort. *Curr Dev Nutr.* 2022;6(6). <https://doi.org/10.1093/cdn/nzac093>
 39. Abdelshafeek K, El-Shamy A. Review on glucosinolates: Unveiling their potential applications as drug discovery leads in extraction, isolation, biosynthesis, biological activity and corrosion protection. *Food Biosci.* 2023;56. <https://doi.org/10.1016/j.fbio.2023.103071>
 40. Okuda A, Kintara Y, Tanabe K, Nakayama T, Shimouchi A, Oku T, Nakamura S. Fructooligosaccharide feeding during gestation to pregnant mice provided excessive folic acid decreases maternal and female fetal oxidative stress by increasing intestinal microbe-derived hydrogen gas. *Nutr Res.* 2023;120:72-87. <https://doi.org/10.1016/j.nutres.2023.09.008>
 41. Chelliah CK, Murugan M, Rajivgandhi G, Gnanasekaran C, Govindan R, Maruthupandy M, et al. Phytochemical derivatives and secondary metabolites rich *Rhizophora mucronata* as an active anti-oxidant and anti-bacterial agent against multi drug resistant bacteria. *J King Saud Univ Sci.* 2023 Nov 1;35(8). <https://doi.org/10.1016/j.jksus.2023.102912>
 42. Wang P, Shan N, Ali A, Sun J, Luo S, Xiao Y, et al. Comprehensive evaluation of functional components, biological activities and minerals of yam species (*Dioscorea polystachya* and *D. alata*) from China. *LWT.* 2022 Oct 1;168. <https://doi.org/10.1016/j.lwt.2022.113964>
 43. Odeh AA, Al-Jaber HI, Barhoumi LM, Al-Fawares O, Shakya AK, Al-Qudah MA, et al. Phytochemical and bioactivity evaluation of secondary metabolites and essential oils of *Sedum rubens* growing wild in Jordan. *Arabian Journal of Chemistry.* 2023 Jun 1;16(6). <https://doi.org/10.1016/j.arabjc.2023.104712>
 44. Morales Bozo I, Ortega Pinto A, Rojas Alcayaga G, Aitken Saavedra JP, Salinas Flores JO, Lefimil Puente C, et al. Reporte preliminar sobre el efecto de un sustituto salival a base de manzanilla (*Matricaria chamomilla*) y linaza (*Linum usitatissimum*) en el alivio de la xerostomía en adultos mayores. *Revista Clínica de Periodoncia, Implantología y Rehabilitación Oral.* 2015 Aug;8(2):144-49. <https://doi.org/10.1016/j.piro.2015.01.001>
 45. Ivane NMA, Wang W, Ma Q, Wang J, Sun J. Harnessing the health benefits of purple and yellow-fleshed sweet potatoes: Phytochemical composition, stabilization methods and industrial utilization- A review. *Food Chem X [Internet].* 2024 May;101462. Available from: <https://linkinghub.elsevier.com/retrieve/pii/S2590157524003493>
 46. Carrera EJ, Cejudo-Bastante MJ, Hurtado N, Heredia FJ, González-Miret ML. Revalorization of Colombian purple corn *Zea mays* L. by-products using two-step column chromatography. *Food Research International.* 2023 Jul 1;169. <https://doi.org/10.1016/j.foodres.2023.112931>
 47. Pacheco MT, Hernandez-Hernandez O, Moreno FJ, Villamiel M. Andean tubers grown in Ecuador: New sources of functional ingredients. *Food Biosci.* 2020;35. <https://doi.org/10.1016/j.fbio.2020.100601>
 48. Jiang Q, Zhao S, Zhao W, Wang P, Qin P, Wang J, et al. The role of water distribution, cell wall polysaccharides and microstructure on radish (*Raphanus sativus* L.) textural properties during dry-salting process. *Food Chem X.* 2024 Jun 30;22. <https://doi.org/10.1016/j.fochx.2024.101407>
 49. Mardani N, Jahadi M, Sadeghian M, Keighobadi K, Khosravi-Darani K. Antimicrobial activities, phenolic and flavonoid contents, antioxidant and DNA protection of the internal and outer layers of *Allium cepa* L. from Iran. *NFS Journal.* 2023 Jun 1;31:93-101. <https://doi.org/10.1016/j.nfs.2023.03.003>
 50. Yang D, Chen H, Wei H, Liu A, Wei DX, Chen J. Hydrogel wound dressings containing bioactive compounds originated from traditional Chinese herbs: A review. *Smart Mater Med.* 2024;5(1):153-65. <https://doi.org/10.1016/j.smaim.2023.10.004>
 51. Bradbury AJ, Emshwiller E. The role of organic acids in the domestication of *Oxalis tuberosa*: A new model for studying domestication resulting in opposing crop phenotypes. *Econ Bot.* 2011;65(1):76-84. <https://doi.org/10.1007/s12231-010-9141-0>
 52. Chuquilín Goicochea RC, Martínez Laurente MC, Rodrigo-Chumbes JT. Propiedades funcionales de productos tradicionales congelados y secados al sol de Oca (*Oxalis tuberosa* Molina) y olluco (*Ullucus tuberosus* Caldas): Una revisión. *Puriq.* 2020;2(3):363-87. <https://doi.org/10.37073/puriq.2.3.100>
 53. Dimas-López D de J, Soto-Simental S, Güemes-Vera N, Ojeda-Ramírez D, Quintero-Lira A, Piloni-Martini J. Optimization of anthocyanin extraction from *Oxalis tuberosa* peel by ultrasound, enzymatic treatment and their combination. *Food Measurement and Characterization.* 2023;17(2):1775-82. <https://doi.org/10.1007/s11694-022-01721-7>
 54. Valle-Parra M, Lalaleo L, Pomboza-Tamaquiza P, Ramírez-Estrada K, Becerra-Martínez E, Hidalgo D. From morphological traits to the food fingerprint of *Tropaeolum tuberosum* through metabolomics by NMR. *LWT.* 2020;119. <https://doi.org/10.1016/j.lwt.2019.108869>
 55. Daza LD, Umaña M, Simal S, Váquiro HA, Eim VS. Non-conventional starch from cubio tuber (*Tropaeolum tuberosum*): Physicochemical, structural, morphological, thermal characterization and the evaluation of its potential as a packaging materi-

- al. *Int J Biol Macromol.* 2022;221(1):954-64. <https://doi.org/10.1016/j.ijbiomac.2022.09.092>
56. Aguilar A, García D, Ramírez D, Lindo J, Chirinos R, Pedreschi R, et al. *In vitro* and *in vivo* biotransformation of glucosinolates from mashua (*Tropaeolum tuberosum*) by lactic acid bacteria. *Food Chem.* 2023;404(1):134631. <https://doi.org/10.1016/j.foodchem.2022.134631>
 57. Mosquera N, Cejudo-Bastante MJ, Heredia FJ, Hurtado N. Identification of new betalains in separated betacyanin and betaxanthin fractions from Ulluco (*Ullucus tuberosus* Caldas) by HPLC-DAD-ESI-MS. *Plant Foods Hum Nutr.* 2020;75(3):434-40. <https://doi.org/10.1007/s11130-020-00837-9>
 58. Svenson J, Smallfield BM, Joyce NI, Sansom CE, Perry NB. Beta-lains in red and yellow varieties of the Andean tuber crop Ulluco (*Ullucus tuberosus*). *J Agric Food Chem.* 2008;56(17):7730-37. <https://doi.org/10.1021/jf8012053>
 59. Galindez A, Daza LD, Homez-Jara A, Eim VS, Váquiro HA. Characterization of Ulluco starch and its potential for use in edible films prepared at low drying temperature. *Carbohydr Polym.* 2019;215:143-50. <https://doi.org/10.1016/j.carbpol.2019.03.074>
 60. Gangta R, Thakur NS, Hamid, Gautam S, Thakur A. Optimization of pre-drying treatment and drying mode for reducing browning to produce shelf stable fructooligosaccharide rich yacon (Ground Apple) powder. *S Afr J Bot.* 2023;157(1):96-105. <https://doi.org/10.1016/j.sajb.2023.03.051>
 61. Martino HSD, Kolba N, Tako E. Yacon (*Smalanthus sonchifolius*) flour soluble extract improve intestinal bacterial populations, brush border membrane functionality and morphology *in vivo* (*Gallus gallus*). *Food Res Intern.* 2020 Nov 1;137. <https://doi.org/10.1016/j.foodres.2020.109705>
 62. Acurio L, Salazar D, García-Segovia P, Martínez-Monzó J, Igual M. Third-generation snacks manufactured from Andean tubers and tuberous root flours: Microwave expansion kinetics and characterization. *Foods.* 2023;12(11). <https://doi.org/10.3390/foods12112168>
 63. Ore-Areche F, Flores DDC, Pacovilca-Alejo OV, Montesinos CCZ, Carrasco SM, Chirre ETC, et al. Effect of different concentrations of stem juice of *Oxalis tuberosa* Mol. and juice of the fruit of *Gaultheria glomerata* (Cav.) sleumer on the antioxidant activity of the heat-treated functional beverage. *Braz J Biol.* 2023;83:1-7. <https://doi.org/10.1590/1519-6984.274986>
 64. Campos-Montiel RG, Vicente-Flores M, Chanona-Pérez JJ, Espino-Manzano SO, De Los Montero-Sierra MJG. Physicochemical, rheological and sensory characterization of a gluten-free English bread added with *Oxalis tuberosa* flour. *Rev Mex Ing Quim.* 2021;20(3). <https://doi.org/10.24275/rmiq/Alim2572>
 65. González Victoriano L, Guemes Vera N, Chel Guerrero LA, Bernardino Nicanor A, Soto Simental S, Chanona Pérez JJ, et al. Physical-chemical characterization and antioxidant properties of extruded products made from mixtures composed of corn grits and red Potato flour (*Oxalis tuberosa*). *CYTA - J Food.* 2019;17(1):69-77. <https://doi.org/10.1080/19476337.2018.1554703>
 66. Castro-Mendoza MP, Palma-Rodriguez HM, Heredia-Olea E, Hernández-Urbe JP, López-Villegas EO, Serna-Saldivar SO, et al. Characterization of a mixture of Oca (*Oxalis tuberosa*) and oat extrudate flours: Antioxidant and physicochemical attributes. *J Food Qual.* 2019;2019:1-10. <https://doi.org/10.1155/2019/1238562>
 67. Mosso AL, Lobo MO, Sammán NC. Development of a potentially probiotic food through fermentation of Andean Tubers. *LWT.* 2016;71:184-89. <https://doi.org/10.1016/j.lwt.2016.03.008>
 68. Fuel M, Mesas C, Martínez R, Ortiz R, Quiñonero F, Prados J, et al. Antioxidant and antiproliferative potential of ethanolic extracts from *Moringa oleifera*, *Tropaeolum tuberosum* and *Annona cherimola* in colorectal cancer cells. *Biomed Pharmacother.* 2021;143. <https://doi.org/10.1016/j.biopha.2021.112248>
 69. Apaza Ticona L, Arnanz Sebastián J, Serban AM, Rumbero Sánchez Á. Alkaloids isolated from *Tropaeolum tuberosum* with cytotoxic activity and apoptotic capacity in tumor cell lines. *Phytochemistry.* 2020;177. <https://doi.org/10.1016/j.phytochem.2020.112435>
 70. Ticona LA, Peña G, Andía V, Durán B, Sánchez R. Anti-glycative and anti-inflammatory effects of macamides isolated from *Tropaeolum tuberosum* in skin cells. *Nat Prod Res.* 2022;36(22):5803-07. <https://doi.org/10.1080/14786419.2021.2016751>
 71. Montes-Lora S, Hurtado N, Mosquera N, Heredia FJ, Cejudo-Bastante MJ. Effect of technological practices on individual betalains and antioxidant activity of Columbian betalain-rich raw materials. *Int J Food Sci Technol.* 2016;51(4):1041-47. <https://doi.org/10.1111/ijfs.13056>
 72. Pérez LV, Sánchez HJ, Cando VM, Sánchez E, Salazar DM. Fortification of low-fat yogurt with mellico flour (*Ullucus tuberosus*): physicochemical and rheological effects. *Afr J Food Agric Nutr Dev.* 2022;22(10):22041-58. <https://doi.org/10.18697/ajfand.115.20870>
 73. dos Santos TPR, Franco CML, Demiate IM, Li XH, Garcia EL, Jane J lin, et al. Spray-drying and extrusion processes: Effects on morphology and physicochemical characteristics of starches isolated from Peruvian carrot and cassava. *Int J Biol Macromol.* 2018;118:1346-53. <https://doi.org/10.1016/j.ijbiomac.2018.06.070>
 74. García A, Pacheco-Delahaye E. Evaluation of a milky instant drink with Arracacha flour (*Arracacia xanthorrhiza*) and addition of folic acid. *Rev Chil Nutr.* 2010;37(4):480-92. <https://doi.org/10.4067/S0717-75182010000400009>
 75. Zhang Y, Yu L, Jin W, Ao M. Effect of ethanolic extract of *Lepidium meyenii* Walp on serum hormone levels in ovariectomized rats. *Indian J Pharmacol.* 2014;46(4):416-19. <https://doi.org/10.4103/0253-7613.135955>
 76. Sales SS, Dionísio AP, Adriano LS, Cordeiro BR, Pinto FA, Carvalho HA, et al. Previous gut microbiota has an effect on postprandial insulin response after intervention with yacon syrup as a source of fructooligosaccharides: a randomized, crossover, double-blind clinical trial. *Nutr.* 2023;109(1):111948. <https://doi.org/10.1016/j.nut.2022.111948>
 77. Gomes AF, Viana ML, Vaz-Tostes M das G, Costa NMB. Yacon (*Smalanthus sonchifolius*) and kefir improved intestinal and bone health but without symbiotic benefits in rats. *Nutr Res.* 2023;118:85-93. <https://doi.org/10.1016/j.nutres.2023.07.009>
 78. Grancieri M, Viana ML, de Oliveira DF, Vaz Tostes M das G, Costa Ignacchiti MD, Costa AGV, et al. Yacon (*Smalanthus sonchifolius*) flour reduces inflammation and had no effects on oxidative stress and endotoxemia in Wistar rats with induced colorectal carcinogenesis. *Nutrients.* 2023;15(14). <https://doi.org/10.3390/nu15143281>
 79. Oh HY, Lee TH, Lee CH, Lee DY, Sohn MY, Kwon RW, et al. Effects of by-products from producing yacon (*Smalanthus sonchifolius*) juice as feed additive on growth performance, digestive enzyme activity, antioxidant status, related gene expression and disease resistance against *Streptococcus iniae* in juvenile black rockfish (*Sebastes schlegelii*). *Aquac.* 2023;569(1):739383. <https://doi.org/10.1016/j.aquaculture.2023.739383>
 80. Widowati W, Tjokropranoto R, Onggowidjaja P, Widya Kusuma H, Wijayanti C, Marthania M, et al. Protective effect of Yacon leaves extracts (*Smalanthus sonchifolius* (Poepp.) H. Rob) through antifibrosis, anti-inflammatory and antioxidant mechanisms toward diabetic nephropathy. *Res Pharm Sci.* 2023;18(3):336-45. <https://doi.org/10.4103/1735-5362.371589>

81. Unión de Organizaciones Campesinas de San Juan "UCASAJ". Elaboración de yogur de mashua [Internet]. EquatorInitiative; 2017 [Consultado el 23 de mayo del 2024]. Disponible en: <https://www.equatorinitiative.org/es/2017/07/10/elaboracion-de-yogur-de-mashua/>
82. ThePerúChef. OLLUCO ENTERO EN SALMUERA PERU CHEF [Internet]. Perúpati [Consultado el 23 de mayo del 2024]. Disponible en: <https://perupationline.com/product/olluco-entero-en-salmuera/>
83. HerbaZest. Harina de Arracacha [Internet]. Herbs&SmartConsumption [Consultado el 23 de mayo del 2024]. Disponible en: <https://shop.herbazest.com/es/products/arracacha-flour>
84. YACON BIOAMAZONIC. Dietary Supplement. [Internet]. Perúpati [Consultado el 23 de mayo del 2024]. Disponible en: <https://perupationline.com/product/yacon-bioamazonic/>
85. Salud Vida. Sirope de Yacón Eco. [Internet]. NATURITAS [Consultado 25 mayo del 2024]. Disponible en: <https://www.naturitas.mx/p/alimentacion/endulzantes-y-mieles/sirope/sirope-de-yacon-eco-345-g-salud-viva>
86. International Potato Center. YACON [Internet]. [Consultado 25 mayo del 2024]. Disponible en: <https://cipotato.org/es/raices-y-tuberculos/yacon/>
87. Manrique I, Arbizu C, Vivanco F, Ramirez C, Chavez O, Tay D, et al. *Tropaelum tuberosum* Ruíz & Pav. Catálogo de la colección de germoplasma de mashua (*Tropaelum tuberosum* Ruíz & Pav.) conservada en el Centro Internacional de la Papa (CIP). Primera. *Tropaelum tuberosum* Ruíz & Pav. Catálogo de la colección de germoplasma de mashua (*Tropaelum tuberosum* Ruíz & Pav.) conservada en el Centro Internacional de la Papa (CIP). Lima: Centro Internacional de la Papa; 2013. 1-123 p. <https://doi.org/10.4160/9789290604310>
88. AGROPIA Cooperativa. Olluco. [Internet]. [Consultado 25 mayo del 2024]. Disponible en: <https://www.agropiaperu.com/producto/olluco/>
89. AGROSAVIA. Cultivar de arracacha AGROSAVIA - La 22. Ficha Técnica; Material Reproductivo [Internet]. [Consultado 25 mayo del 2024]. Disponible en: <https://www.agrosavia.co/productos-y-servicios/oferta-tecnologica/ladnea-agrosavia-dcola/raices-y-tuberculos/material-reproductivo/549-cultivar-de-arracacha-agrosavia-la-22>
90. Secretaria de Agroindustria. YACÓN “Cultivo de raíces dulces”. [Internet]. Nutrición y Educación Alimentaria; Alimentos Argentinos 2013 [Consultado 25 de mayo del 2024]. Disponible en: https://alimentosargentinos.magyp.gob.ar/HomeAlimentos/seguridad-alimentaria-y-nutricion/fichaspdf/Ficha_21_Yacon.pdf