



RESEARCH ARTICLE

Assessment of the cytotoxic effect of aerial parts of *Gazania rigens* hexane extract on HRT-18 and MCF-7 cell line and chemical composition analysis using GC/MS and LC/MS

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ARTICLE HISTORY

Received: 23 June 2024

Accepted: 17 October 2024

Available online

Version 1.0: 05 December 2024



Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

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Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/index.php/PST/indexing_abstracting

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CITE THIS ARTICLE

Jawad HA, Kadhim EJ. Assessment of the cytotoxic effect of aerial parts of *Gazania rigens* hexane extract on HRT-18 and MCF-7 cell line and chemical composition analysis using GC/MS and LC/MS. Plant Science Today (Early Access). <https://doi.org/10.14719/pst.4164>

Abstract

Gazania rigens are cultivated for their vividly colored flowerheads. The herb, belonging to the Asteraceae family, displays various classes of secondary metabolites, including terpenes, phenols and fatty acid derivatives. Antioxidant, hepatoprotective, nephroprotective, and antimicrobial activities of the herb have been evaluated and have proven to benefit humans clinically specially in case of cancer affected people. Breast cancer is the second most frequent reason for female mortality, and the third most frequent reason in the world is colorectal cancer. A previous study stated the *in vitro* anticancer effect of the genus *Gazania rigens* on the MCF-7 breast cancer cell. N-hexane extract of *Gazania rigens* aerial parts was prepared by maceration, cold extraction methods and subjected to GC/MS and LC-MS analysis to characterize its constituents. In GC/MS, the total number of identified compounds were 2-pentadecanone, 6,10,14-trimethyl, hexadecanoic acid methyl ester, 11-octadecenoic acid methyl ester, methyl stearate hexanedioic acid, and bis(2-ethylhexyl) ester. LC/MS analysis revealed and confirmed the presence of lupeol. Based on the result, n-hexane extract was evaluated for its *in vitro* anticancer effect against the MCF-7 and HRT-18 cell lines using an MTT assay. Results from the MTT assay showed there is a significant cytotoxic effect for hexane extract against both cell line HRT-18 and MCF-7, the IC₅₀ value for HRT-18 was 102.2±10 µg/ml, while the IC₅₀ value for MCF-7 was 121.2±12 µg/ml.

Keywords

Gazania; HRT-18 cell line; MCF-7 cell line; cytotoxic

Introduction

Cancer is a complex illness that is typically incurable and remains as one of the leading causes of death globally (1). Breast cancer is the second most frequent reason for female mortality (3), and metastases from breast cancer primarily affect the central nervous system, bone, lungs, liver, and chest wall (4). There is now no known cure for cancer. Hence, the need for a reliable and secure therapy for the cancer is important and has been considered as a main goal to decrease mortality rate. The poor solubility, the low stability of the drugs, the low proportion that the tissues absorb from the drug, and the tumor treatment resistance often limits the effectiveness of carcinoma medications (5). Additionally, a high percentage of side effects are linked to antineoplastic medications, which increase the risk of toxicity (6). Conse-

quently, a great deal of the side effects connected to the chemotherapeutic drugs that are already available drive the search for new, extremely potent, and readily absorbable drugs. There has been a surge in pharmacological research using plant extracts in an effort to yield safer and more effective chemotherapeutic drugs that are derived from plants (7). There are more than 32,000 identified flowering plant species in more than 1,900 genera within the Asterales order, which make up the family Asteraceae. Genus *Gazania*, which belongs to the family Asteraceae of flowering plants, is native to South Africa. *Gazania* species are cultivated for their vividly coloured flowerheads, which arise in late spring and frequently bloom from summer to autumn. *Gazania rigens* is a plant belonging to Asteraceae family. *Gazania rigens* are indigenous to South Africa and Mozambique. In folk medicine, gazania has been used to treat toothaches and miscarriages. It has also been used in purgative medicines. The biological effects of *Gazania* have been part of a few reported investigations, which involve antioxidant and hepatoprotective properties of *G. nivea* (8) and the antibacterial activity of *G. rigens* (9). Reports also have shown that the *Gazania rigens* plant possesses anti-microbial, antioxidant, and hepatoprotective properties (9, 10). The isolated compounds from *G. rigens* include phenolic compounds (Gallic acid, 3,5-di-O-caffeoylquinic acid) and flavonoids (Rutin, Apigenin, Luteolin) (10, 11). Thus, study on HRT-18 and MCF-7 cell lines anticancer properties along with isolation and identification of some pharmacological active compound in n-hexane extract of aerial parts of *G. rigens* cultivated in Iraq, was conducted. *G. rigens* was selected as plant because there is no study for its anti-cancer effect but there is study for other species like *G. linearis*. We selected breast and colon cancer cell because they are most common cancers worldwide (2).

Materials and Methods

Plant materials

In November 2023, *G. rigens* aerial parts (stems, leaves, flowers) were gathered from Iraq, the Baghdad city at latitude 33°18'00.00" North, longitude 44°24'00.00" East. Prof. Dr. Sukyna Abass of the University of Baghdad's Department of Biology and College of Sciences confirmed and identified the plant. After cleaning, the parts were allowed to air dry before being processed using a machine grinder to a powder. The hexane extract was prepared using maceration (cold method). Firstly, the dried powder was weighed and then extraction was done by cold method maceration using n-hexane as a solvent for 24 hrs. Then the sample was filtered and rotary evaporator was used to dry the residue, then weighed again (12).

Initial testing of plant extracts

Chemical tests were used to check the presence of sterols, steroids, terpenoids, coumarin, flavonoid, phenolic compounds (13). The Salkowski test was for checking the presence of terpenoids. It was performed by taking 1 mL of sample in a test tube and dissolving it in 1 mL chloroform. An equal volume of concentrated sulphuric acid was

added to it and the presence of red color indicated the presence of terpenoids. Keller-Kiallian's test: The Keller-Kiallian's test was performed by adding 1ml of the extracts in 1ml of glacial acetic acid, 2-3 drops of ferric chloride and 2ml of concentrated sulphuric acid was added carefully along the walls of the test tube, a reddish-brown ring indicated the presence of cardiac glycoside.

Identification and characterization of isolated lupeol by Liquid chromatography-mass spectrometry (LC-MS)

The analytical LC/MS had been carried out in the Jordan University of Science and Technology at Irbid, Jordan. The following conditions for liquid chromatography were employed: column: GL-Science-C18- 250mm x 4.6 (5µm particle size) – Japan; oven temperature: 35 °C; injection volume: 10 µm; flow rate: 1 ml/min; run duration: 25 min. The mass parameter was as follows: software AB-Siecx-OS, ionization mode ESI Positive, scan range (50-800 m/z), ion source voltage 5500V, and LCMSMS-Q-TOF model X500 QTOF.

By Gas chromatography-mass spectrometry (GC/MS) analysis

The n-hexane extract of the *G. rigens* was analyzed using Gas Chromatograph. Agilent Technologies (7820A) Gas, GC Mass Spectrometer (5977E) USA having 30 m length, 250 µm inner diameter, and 0.25 µm film thickness were employed in the HP-5ms Ultra Inert column. The injection volume was 1µl, and the scan range was m/z 25–1000. The following parameters were used in the analysis: splitless injection type, carrier gas: helium 99.99%, pressure: 11.933 psi, temperatures of the injector are 250 °C, the inlet line is 250 °C, and the aux heaters are 300 °C.

Determination of the cytotoxicity

To assess the cytotoxic activity of n-hexane extract against Breast cancer and colon cancer, HRT-18 and MCF-7 cells were employed as models and work conducted in triplicate.

Maintenance of cell cultures

A humidified atmosphere was maintained at 37°C for the cell lines culture in MEM (US Biological, USA) supplemented with 10% (v/v) fetal bovine serum (FBS) (Capricorn-Scientific, Germany), 100 IU penicillin, and 100 µg streptomycin (Capricorn-Scientific, Germany). For the experiments, exponentially growing cells were employed (14, 15).

Cytotoxicity Assays

A 96-well microplate (NEST Biotech, China) was seeded with 10,000 cells per well, and the cells were then cultured at 37 °C for 72 hours or until monolayer confluence was reached. The 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide (MTT) test (Elabscience, China) was utilized to examine cytotoxicity. A range of concentrations (1000, 500, 250, 125, 62, and 31 µg) were applied to the cells. After 72h of infection, MTT dye solution 28 µL of (2 mg/ml) was added to each well. Three hours were spent on the incubation process. Each well received 100 µl of DMSO, which was then incubated for 15 minutes. The opti-

cal density was determined with a microplate reader at 492 nm (14, 16). Cytotoxicity % was measured by the following equation: Cytotoxicity % = (OD Control – OD sample)/OD Control × 100, where OD control is the mean optical density of untreated wells, and OD Sample is the optical density of treated wells (17).

Statistical analysis

Using GraphPad Prism 8 and Tukey's ANOVA multiple comparison test, the collected data were statistically examined. The values were shown as the triple measurements' mean ± standard deviation.

Results and Discussion

Initial testing of plant extracts

This screening was carried out to provide an overview of the terpenoids, sterols, cardiac glycoside contained in the *G. rigens* aerial part extract. The components contained in the extract were analyzed by their compounds by color test (qualitative) with several reagents like the Salkowski test is commonly used to detect the presence of terpenoids and sterols in plant extracts, and Keller-Kiallian's test to detect the presence of cardiac glycosides, both Salkowski test and Keller-Kiallian's test gave a positive result in n-hexane extract.

GC/MS analysis

Data in Fig. 1 shows identified 20 peaks GC/MS chromatogram. Identified through comparison of the mass spectrum fragmentation, peak retention duration, peak area (%), height (%), known compounds listed in the National Institute of Standards and Technology (NIST) collection.

According to the results, five chemicals were found in the n-hexane extraction.

The phytoconstituents in the n-hexane extract of *G. rigens* were found to be 2-pentadecanone, 6,10,14-trimethyl, hexadecanoic acid, methyl ester, 11-octadecenoic acid, methyl ester, methyl stearate, and hexanedioic acid, bis(2-ethylhexyl) ester. Table 1 displays the structure, molecular weight, peak area, and Chemical Abstracts Service of the five phytocompounds found in n-hexane extracts.

Identification of isolated compound by LCMS/MS spectroscopy

Recognition of peaks of lupeol isolated from *G. rigens* was done by examining mass spectra of standard data as shown in Fig. 2 and literature. From the full scan mass spectra of the isolated lupeol, and Fig. 3 show mass fragments of lupeol, some of the fragments were 218 m/z (C₁₆H₂₆), 203 m/z (C₁₅H₂₃) these fragments were closely similar to that reported in literature for lupeol (18, 19).

Cytotoxic effect

In this study, the cytotoxic effect of *G. rigens* n-hexane extract versus tumor cells was assessed using the MCF-7 and HRT-18 cell lines. The outcomes show that there is a highly significant cytotoxic action against the cell lines HRT-18 colon cancer and MCF-7 that represent breast cancer as shown in Fig. 4. The data below demonstrates the capacity of n-hexane extract to significantly suppress the proliferation of colon cancer HRT-18 and breast cancer MCF-7 cell lines in a concentration-dependent way as shown in Fig. 5. The IC₅₀ of n-hexane value was 102.2 µg/ml for HRT-18; for MCF-7, the IC₅₀=121.2±12µg/ml. The data demonstrated the capacity of n-hexane extract to significantly suppress

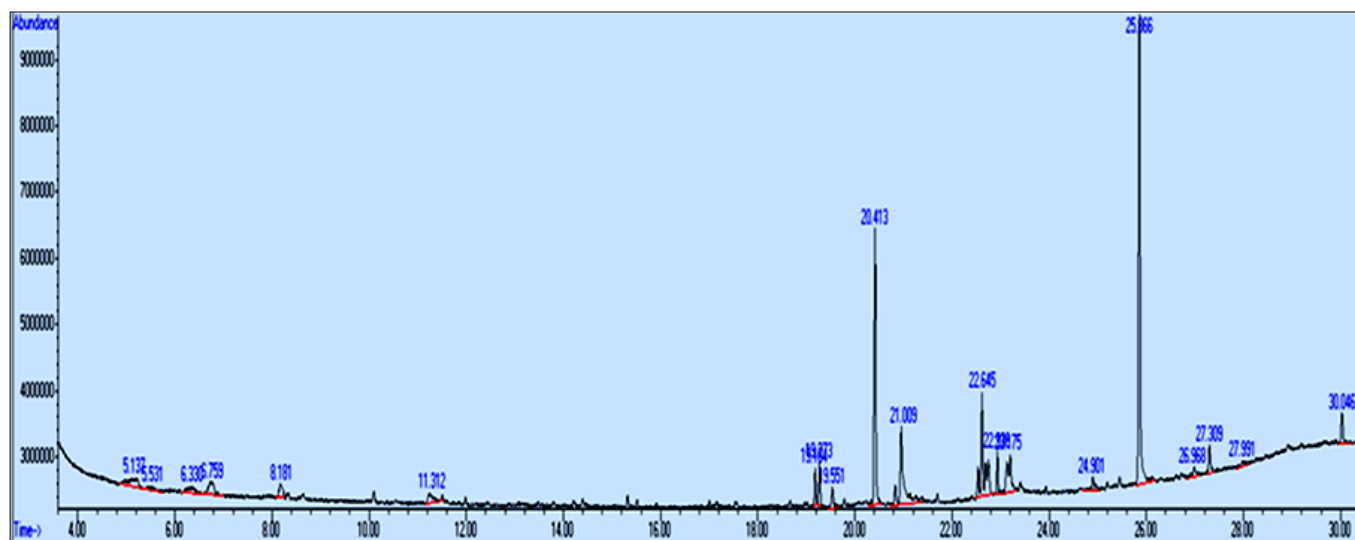


Fig. 1. GC/MS chromatogram of the *Gazania rigens* n-hexane extract.

Table 1. GC/MS analysis of hexane extract.

No	CAS	Name of the compound	Molecular formula	Molecular weight	Peak area (%)	Retention Time (min)
1	502-69-2	2-Pentadecanone, 6,10,14-trimethyl	C ₁₈ H ₃₆ O	268.4778	2.05	19.270
2	112-39-0	Hexadecanoic acid, methyl ester	C ₁₇ H ₃₄ O ₂	270.4507	15.10	20.413
3	52380-33-3	11-Octadecenoic acid, methyl ester	C ₁₉ H ₃₆ O ₂	296.4879	11.55	22.646
4	112-61-8	Methyl stearate	C ₁₉ H ₃₈ O ₂	298.5038	2.03	22.931
5	103-23-1	Hexanedioic acid, bis(2-ethylhexyl) ester	C ₂₂ H ₄₂ O ₄	370.5665	29.68	25.866

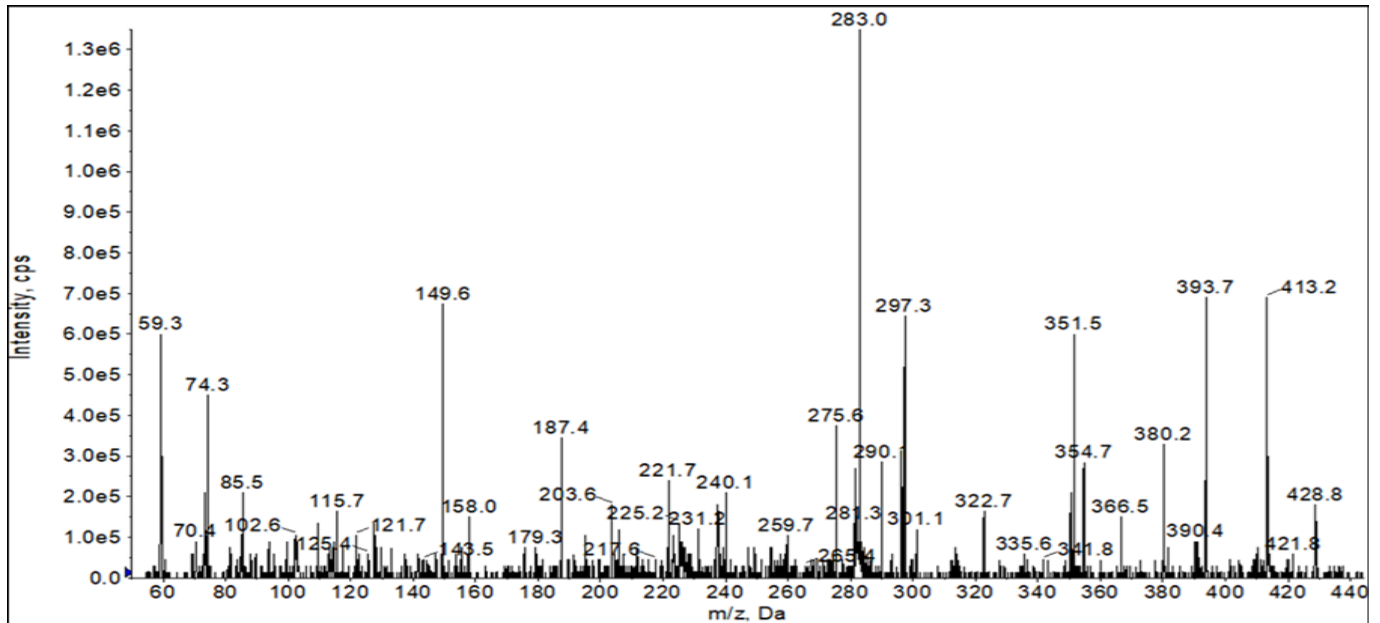


Fig. 2. LC-MS of lupeol.

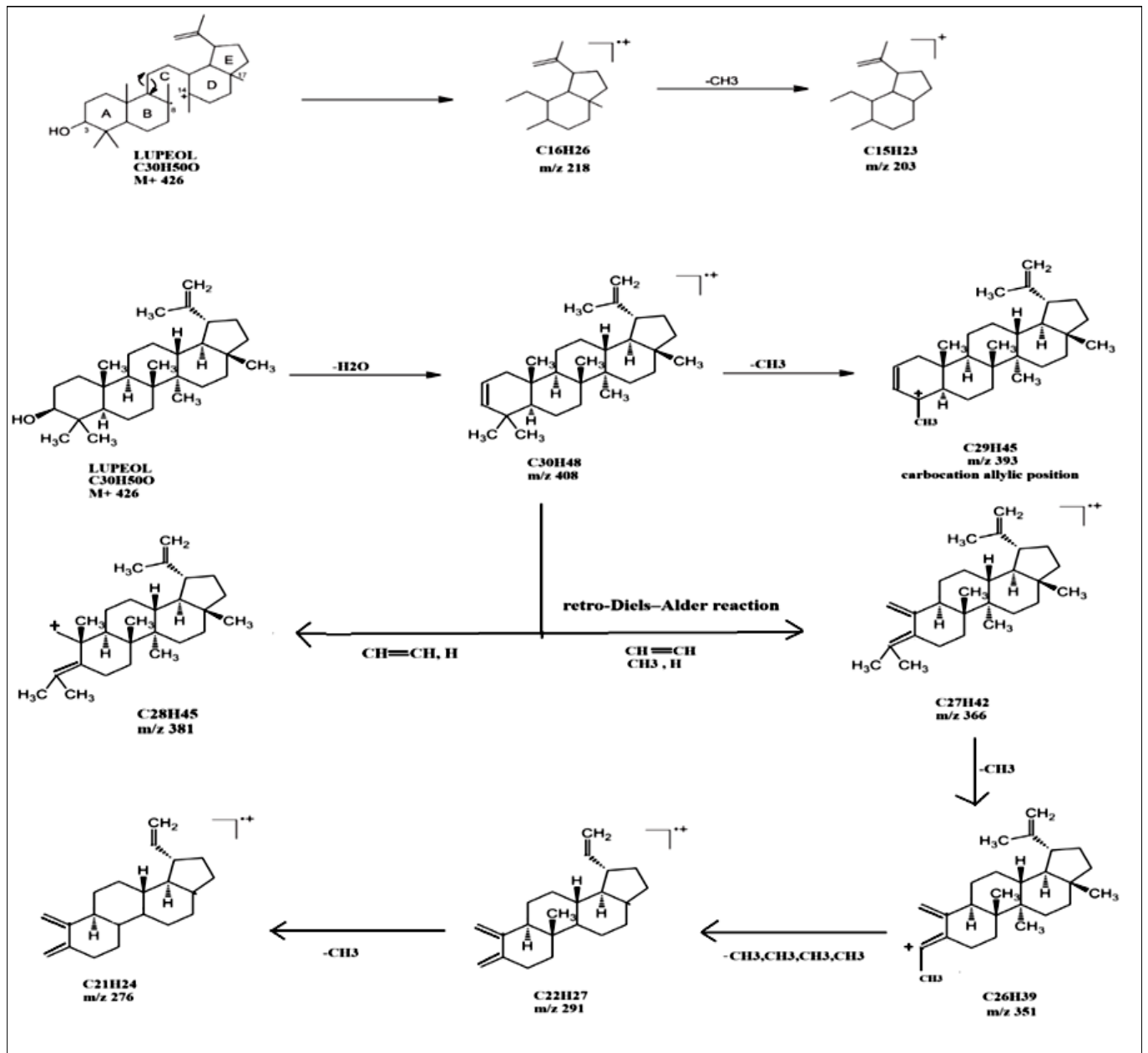


Fig. 3. Mass Fragment of lupeol.

the proliferation of colon cancer HRT-18 and breast cancer. The IC₅₀ of n-hexane value was 102.2 µg/ml for HRT-18;

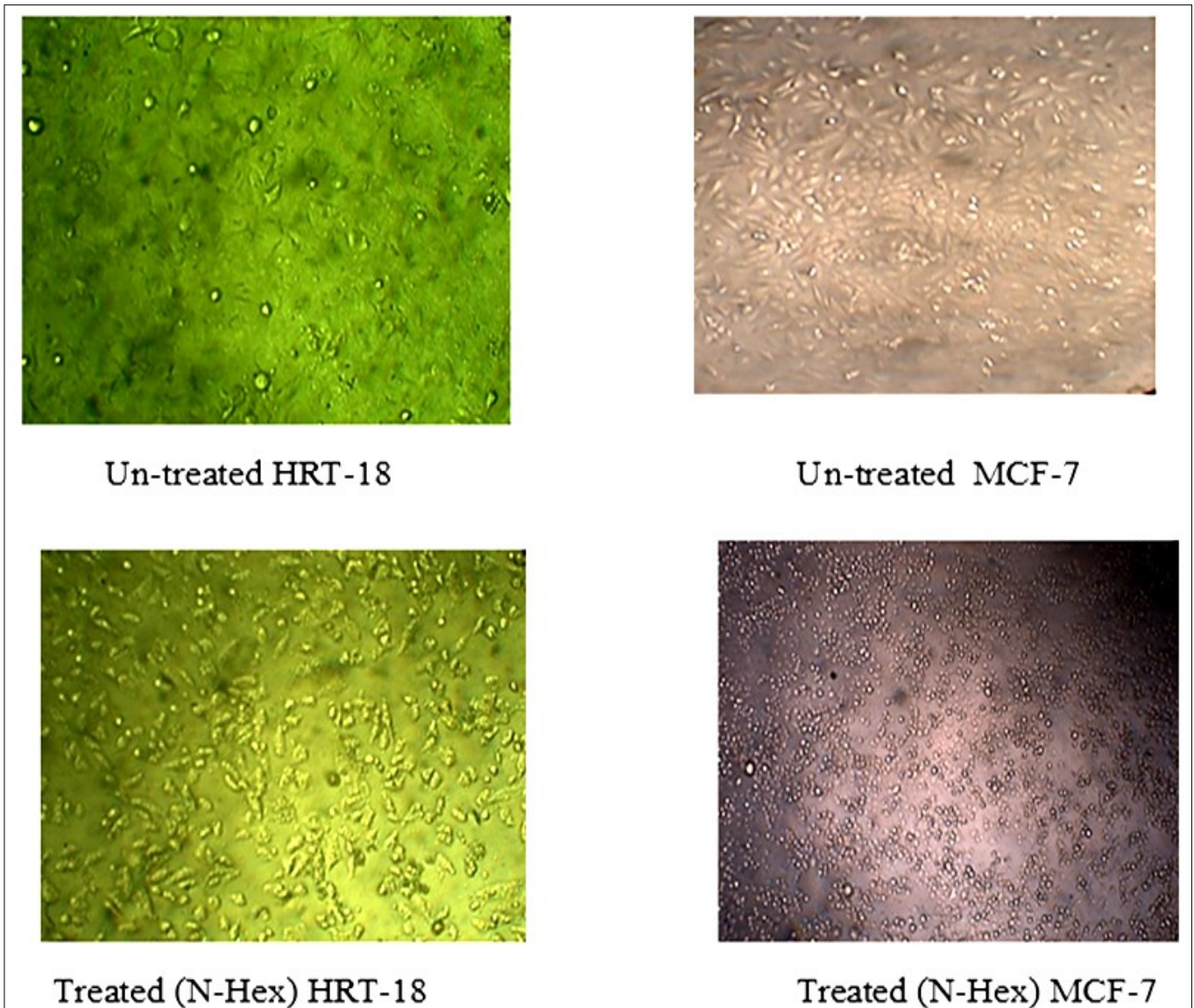


Fig. 4. Morphological changes in MCF-7 and HRT-18 cell lines before and after being treated with *Gazania rigens* n-hexane extract.

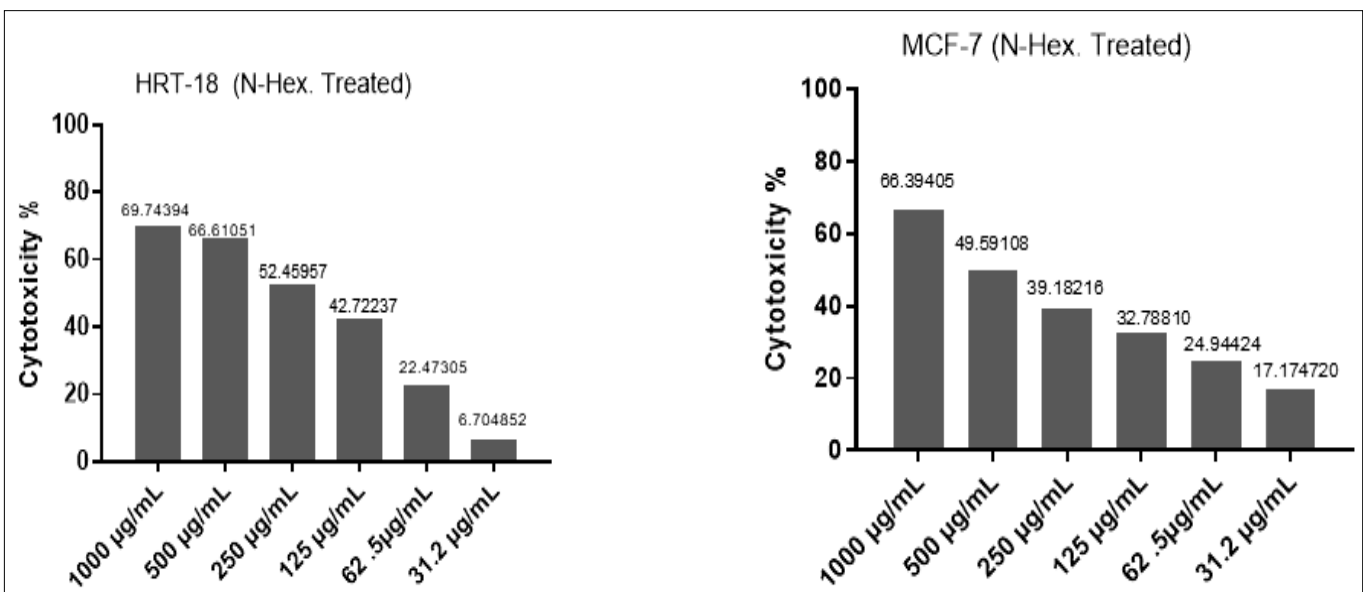


Fig. 5. Cytotoxic effect of *Gazania rigens* n-hexane extract on MCF-7 and HRT-18 in a concentration-dependent way.

for MCF-7, the $IC_{50}=121.2\pm 12\mu\text{g/ml}$. The presence of terpenoids, fatty acids in n-hexane extract, may be the cause of their antiproliferative properties. The terpenoids are able

to inhibit tumor cell growth by inhibiting multiple cancer-specific targets, including the proteasome, NF- κ B, and antiapoptotic protein Bcl-2 (20, 21). Lupeol suppresses EGFR/STAT3 activity, activates the mitochondrial-mediated apoptosis pathway, inhibits the Akt/PKB path-

way and promotes mitochondrial hyperfission which causes cancer cells to die (22). This mechanism of action of terpenoid and lupeol may play an important role in the folk medicine use of *G. rigens* to treat toothaches and miscarriages (9). In comparison of IC₅₀ of *G. rigens* n-hexane extract with a previous study done on the same extract using same MTT assay method but different plant species, IC₅₀ for *G. linearis* in MCF-7 cell line was 2.43×10⁷ ug/ml, which has higher concentration of IC₅₀ by about 200,495 times than *G. rigens* n-hexane fraction IC₅₀ concentration (23).

Conclusion

The plant is a rich source of terpenoids and fatty acids. The obtained data demonstrated the presence of terpenoids, fatty acids in cold method hexane extract. The present study shows that cold hexane extract *G. rigens* exhibits a significant cytotoxic effect against the colon cancer (HRT-18) cell line and breast cancer (MCF-7) cell line, this work highlights the importance of terpenoids in cancer treatment and throws a light on it as a natural rich source of terpenoid, which may be used as a supplement in the future as for cancer patient.

Acknowledgements

The authors appreciate the role of the pharmacognosy and medicinal plants Department / College of the Pharmacy / University of Baghdad for providing the opportunity to accomplish this study.

Authors' contributions

The authors conceived and planned the experiments and carried out the sample preparation, extraction process, identification, isolation and structure elucidation. The authors also wrote the manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interests to declare.

Ethical issues: None.

AI Declaration

None

References

- Rahib L, Smith BD, Aizenberg R, Rosenzweig AB, Fleshman JM, Matrisian LM. Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver and pancreas cancers in the United States. *Cancer Research*. 2014 Jun 1;74(11):2913-21. <https://doi.org/10.1158/0008-5472.CAN-14-0155>
- Lee SY, Haq F, Kim D, Jun C, Jo HJ, Ahn SM, Lee WS. Comparative genomic analysis of primary and synchronous metastatic colorectal cancers. *PLoS One*. 2014 Mar 5;9(3):e90459. <https://doi.org/10.1371/journal.pone.0090459>
- Nor Aini AS, Merrina A, Stanslas J, Sreeramanan S. Cytotoxic potential on breast cancer cells using selected forest species found in Malaysia. *International Journal of Cancer Research*. 2008 Jun 15;4(3):103-09. <https://doi.org/10.3923/ijcr.2008.103.109>
- Zia MK, Rmali KA, Mansel RE, Jiang WG. Level of expression of parathyroid hormone related protein and its receptor in human breast cancer and its correlation with the clinical outcome. *Int J Cancer Res*. 2007;3:92-102. <https://doi.org/10.3923/ijcr.2007.92.102>
- Akindede AJ, Wani ZA, Sharma S, Mahajan G, Satti NK, Adeyemi OO, et al. *In vitro* and *in vivo* anticancer activity of root extracts of *Sansevieria liberica* Gerome and Labroy (Agavaceae). *Evidence-Based Complementary and Alternative Medicine*. 2015;2015(1):560404. <https://doi.org/10.1155/2015/560404>
- Prakash OM, Kumar A, Kumar P. Anticancer potential of plants and natural products. *Am J Pharmacol Sci*. 2013;1(6):104-15. <https://doi.org/10.12691/ajps-1-6-1>
- Ismail NR, Kadhim EJ. Phytochemical screening and isolation of new compounds. *International Journal of Drug Delivery Technology*. 2021;11(3):1033-39.
- Taha SE, Mona HH, Shahira ME, Amany AS. Pharmacognostical and biological studies on the rhizomes of *Gazania nivea* DC cultivated in Egypt. *Bull Fac Pharm., Cairo Univ*. 2008;46(2):253-65.
- Hammoda H. Biologically guided phytochemical investigation of *Gazania rigens* L. Hala M. Hammoda. *Alex. J Pharm Sci*. 2009;23:96.
- Desoukey SY, El Kady WM, Salama AA, Hagag EG, El-Shenawy SM, El-Shanawany MA. Hepatoprotection and antioxidant activity of *Gazania longiscapa* and *G. rigens* with the isolation and quantitative analysis of bioactive metabolites. *International Journal of Pharmacognosy and Phytochemical Research*. 2016;8(7):1121-31.
- Gomaa AA, Samy MN, Attia EZ, Attia ME, Fawzy MA, Desoukey SY, Kamel MS. Antioxidant, hepatoprotective and nephroprotective activities of *Gazania rigens* against carbon tetrachloride-induced hepatotoxicity and nephrotoxicity in rats. *Tradit Med Res*. 2022 Aug 3;7(5):44. <https://doi.org/10.53388/TMR20220409001>
- Satar_AL_Baaj A, Abdul-Jalil TZ. Phytochemical screening of petroleum ether fractions by GC/MS and isolation of lupeol from two different parts of Iraqi *Leucaena leucocephala*. *Conference Paper. Iraqi Journal of Pharmaceutical Sciences (P-ISSN 1683-3597 E-ISSN 2521-3512)*. 2022;31(Suppl.):62-74. <https://doi.org/10.31351/vol31issSuppl.pp62-74>
- Abdlkareem SK. Isolation, identification and quantification of two compounds from *Cassia glauca* cultivated in Iraq. *Iraqi Journal of Pharmaceutical Sciences (P-ISSN 1683-3597 E-ISSN 2521-3512)*. 2023 Dec 30;32(3):95-104. <https://doi.org/10.31351/vol32iss3pp95-104>
- Freshney RI. *Culture of animal cells: a manual of basic technique and specialized applications*. John Wiley and Sons; 2015 Dec 23.
- Olewi MA, Zalzal MH. Synthesis, molecular docking study and cytotoxicity evaluation of some quinazolinone derivatives as nonclassical antifolates and potential cytotoxic agents. *Iraqi Journal of Pharmaceutical Sciences (P-ISSN 1683-3597 E-ISSN 2521-3512)*. 2022 Dec 25;31(2):283-96. <https://doi.org/10.31351/vol31iss2pp283-296>
- Zuhair Abdul-Lalil T. Ultrasound-assisted extraction of fennel leaves: Process optimization, thin layer chromatography and cytotoxic activity of ethanolic extract. *Iraqi Journal of Pharmaceutical Sciences (P-ISSN 1683-3597 E-ISSN 2521-3512)*. 2024

- Mar 26;33(1):94-103. <https://doi.org/10.31351/vol33iss1pp94-103>
17. Salman MI, Al-Shammari AM, Emran MA. 3-dimensional coculture of breast cancer cell lines with adipose tissue-derived stem cells reveals the efficiency of oncolytic newcastle disease virus infection via labeling technology. *Frontiers in Molecular Biosciences*. 2022 Sep 12;9:754100. <https://doi.org/10.3389/fmolb.2022.754100>
 18. Santos PF, Gomes LN, Mazzei JL, Fontão AP, Sampaio AL, Siani AC, Valente LM. Polyphenol and triterpenoid constituents of *Eugenia florida* DC. (Myrtaceae) leaves and their antioxidant and cytotoxic potential. *Química Nova*. 2018;41:1140-49. <https://doi.org/10.21577/0100-4042.20170284>
 19. Kadhim EJ. Determination and isolation of valuable bioactive compound (lupeol) from *Portulacaria afra* Jacq. *International Journal of Drug Delivery Technology*. 2023;13(1):199-204. <https://doi.org/10.25258/ijddt.13.1.30>
 20. Yang H, Ping Dou Q. Targeting apoptosis pathway with natural terpenoids: implications for treatment of breast and prostate cancer. *Current Drug Targets*. 2010 Jun 1;11(6):733-44. <https://doi.org/10.2174/138945010791170842>
 21. Mahmood MA, Abd AH, Kadhim EJ. Investigating the impact of phenolic and terpene fractions extracted from *Prunus arabica* on p53 protein expression in AMJ13 and SK-GT-4 human cancer cell lines. *Tropical Journal of Natural Product Research*. 2023 Nov 1;7(11). <https://doi.org/10.26538/tjnpr/v7i11.35>
 22. Liu K, Zhang X, Xie L, Deng M, Chen H, Song J, et al. Lupeol and its derivatives as anticancer and anti-inflammatory agents: Molecular mechanisms and therapeutic efficacy. *Pharmacological Research*. 2021 Feb 1;164:105373. <https://doi.org/10.1016/j.phrs.2020.105373>
 23. Elkhayat ES. Chemical constituents from *Gazania linearis* cultivated in Egypt. *Bulletin of Faculty of Pharmacy, Cairo University*. 2016 Dec 1;54(2):257-61. <https://doi.org/10.1016/j.bfopcu.2016.08.001>