



REVIEW ARTICLE

Eco-renaissance in floriculture: unlocking the power of plant growth-promoting bacteria

Mohamed Asik M¹, C Subesh Ranjith Kumar^{1*}, P Irene Vethamoni², Sivakumar Uthandi³, R Jagadeeswaran⁴ & M Djanaguiraman⁵

¹Department of Floriculture and Landscape Architecture, Horticultural College and Research Institute, TNAU, Coimbatore, Tamil Nadu - 641 003, India

²Horticultural College and Research Institute, TNAU, Coimbatore, Tamil Nadu - 641 003, India

³Department of Agricultural Microbiology, TNAU, Coimbatore, Tamil Nadu - 641 003, India

⁴Department of RS & GIS, TNAU, Coimbatore, Tamil Nadu - 641 003, India

⁵Department of Crop Physiology, TNAU, Coimbatore, Tamil Nadu - 641 003, India

*Email: subesh@tnau.ac.in



ARTICLE HISTORY

Received: 13 August 2024

Accepted: 30 August 2024

Available online

Version 1.0 : 25 September 2024

Version 2.0 : 01 October 2024



Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>)

CITE THIS ARTICLE

Asik MM, Kumar CSR, Vethamoni PI, Uthandi S, Jagadeeswaran R, Djanaguiraman M. Eco-renaissance in floriculture: unlocking the power of plant growth-promoting bacteria. *Plant Science Today*. 2024; 11(4): 98-105. <https://doi.org/10.14719/pst.4639>

Abstract

The conventional floriculture industry heavily relies on chemical fertilizers and pesticides to meet global demand, which can have detrimental environmental impacts. To promote sustainable flower cultivation, Plant Growth-Promoting Bacteria (PGPB) has emerged as eco-friendly tools to enhance the productivity of major commercial flower crops. This review summarizes the current knowledge on diverse PGPB genera associated with ornamentals like rose, gerbera, carnation, chrysanthemum and orchids. It examines the abilities of PGPB to improve yield, quality and stress tolerance of flowers through mechanisms such as biological nitrogen fixation, nutrient solubilization, phytohormone synthesis, induced systemic resistance and antagonism against pathogens. The potential of developing microbial consortia and bioformulations as PGPB-based biofertilizers, bio stimulants and biopesticides for sustainable floriculture is discussed. By identifying research gaps and prospects, this review highlights the role of PGPB in driving sustainable intensification of commercial flower production with reduced environmental footprint.

Keywords

Plant growth promoting bacteria; floriculture; sustainable flower production; phytohormones

Introduction

Floriculture, defined as the cultivation of flowering and ornamental plants for gardens and floristry, is an economically important sector of commercial agriculture. According to a report from Floriculture magazine, the global market for flowers and ornamental plants is projected to increase by 6.3 % in the next 5 years, reaching \$57.4 billion in 2024, up from \$42.4 billion in 2019 (1). Flower cultivation and trade play a vital role in the agriculture of many developing countries like Kenya, Colombia and Ethiopia. Production and export of cut flowers, potted flowering plants, cut foliage, flower seeds, bulbs and tubers provide employment and generate foreign exchange revenue. India ranks second in flower production after China but hold only the 14th position in exports (2). The Indian floriculture sector has experienced substantial growth, with loose and cut flower production increasing at CAGRs of 9.92 % and 26.66 %, respectively (3). However, India's share in global floriculture exports remains low at 0.6 % (4).

The major flower crops grown for commercial purposes include Rose, Chrysanthemum, Gerbera, Carnation, Lily, Tulip, Orchids, Anthurium, Gladiolus, Iris, Marigold, Petunia, Pansy, Impatiens, Poinsettia, Zinnia and Sunflower. Flowers are cultivated for their aesthetic appeal as decorative items or as raw materials for extractives used in perfumes, cosmetics and food products. The key objectives in floriculture are to maximize flower productivity in terms of earliness, number and size while maintaining quality. However, several challenges are encountered in intensive flower farming which hamper growth and productivity. Nutrient deficiencies, salinity stress, drought stress, high temperatures, light stress and fungal diseases affect the flowering behaviour and vase life of flowers leading to economic losses. Excessive reliance on agrochemicals for enhancing production can cause environmental hazards. Sustainable solutions are needed to increase the productivity and quality of flower crops.

In recent years, scientists are exploring the use of bacteria and other microorganisms as a sustainable way to enhance flower production and quality. Plant Growth Promoting Bacteria (PGPB) are beneficial soil bacteria that can enhance growth, flowering and postharvest attributes in diverse flower crops through mechanisms like phytohormone production, nitrogen fixation, phosphorus solubilization, induced systemic tolerance against stresses and disease control (5). PGPB-based bio stimulants and bio-inoculants provide a biological alternative to agrochemicals for enhancing productivity in floriculture. This review overviews major studies where plant growth-promoting bacteria (PGPB) sustainably enhanced growth, flowering, yield and vase life of major ornamental crops. It examines the eco-friendly mechanisms by which PGPB improves flower productivity and quality, such as biological nitrogen fixation and induced systemic resistance, offering sustainable alternatives to agrochemicals. The potential of developing PGPB-based bioproducts like biofertilizers and biopesticides for sustainable floriculture is discussed. Limitations in transitioning PGPB technologies from lab to field for reducing the environmental impacts of commercial flower farming are also identified.

2. Plant Growth Promoting Bacteria (PGPB)

Plant growth-promoting bacteria (PGPB) can help plants grow by either direct method such as providing nutrients (like nitrogen) or adjusting hormone levels in plants or indirectly by protecting plants from harmful bacteria and pests (acting as biocontrols). These PGPB can exist in different forms: free-living, symbiotic (like Rhizobia and Frankia), or within plant tissues (endophytes). Despite their diversity, they all use the same methods to promote plant growth (6) (Fig. 1).

2.1. Role of PGPB in improving Flower Crops

PGPB augment plant growth through diverse mechanisms such as atmospheric nitrogen fixation, phosphorus solubilization, iron chelation, phytohormone synthesis, 1-aminocyclopropane-1-carboxylate (ACC) deaminase activity, volatile organic compound (VOC) emission,

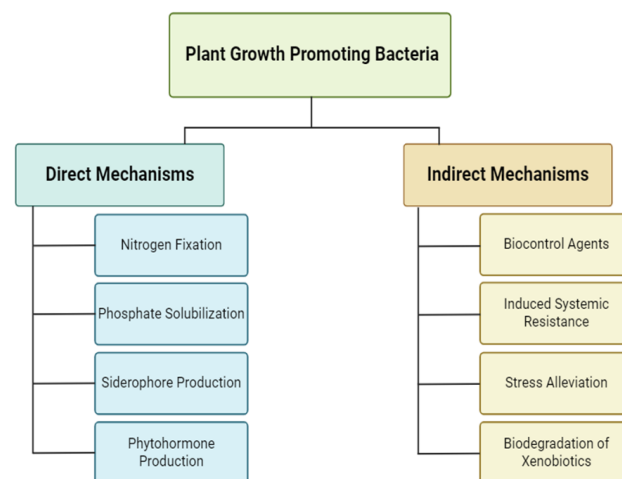


Fig. 1. PGPB classification based on its role in plant growth. (Figure created with Biorender.com)

improved nutrient uptake and disease suppression (7-9). Several of these traits are relevant for improving flowering behaviour, flower productivity and quality in ornamental plants. Recent studies have revealed intricate molecular interactions between PGPB and ornamental plants. For instance, research on *Chrysanthemum morifolium* showed that *Bacillus subtilis* CBR05 inoculation upregulates genes involved in jasmonic acid and ethylene signaling pathways, enhancing both growth and stress tolerance (10). Additionally, bacterial volatile organic compounds like 2,3-butanediol and acetoin produced by *Bacillus subtilis* GB03 have been found to promote growth and flower production in *Pelargonium peltatum* by modulating auxin transport and gibberellin biosynthesis (11). These advancements in understanding PGPB mechanisms offer new possibilities for sustainable ornamental plant production and stress management (Fig. 2).

2.1.1. Phytohormone Production

Many PGPB belonging to genera *Azospirillum*, *Azotobacter*, *Bacillus*, *Burkholderia*, *Enterobacter*, *Klebsiella*, *Pantoea*, *Pseudomonas*, *Rhizobium*, *Serratia* and *Stenotrophomonas* have the ability to synthesize phytohormones including auxin, cytokinin, gibberellin and ethylene (12-14). Of these, auxin (primarily indole-3-acetic acid) and cytokinin are well known for their role in plant growth and organ development. Auxin stimulates root elongation while cytokinin promote shoot proliferation. The relative levels of these 2 hormone types determine

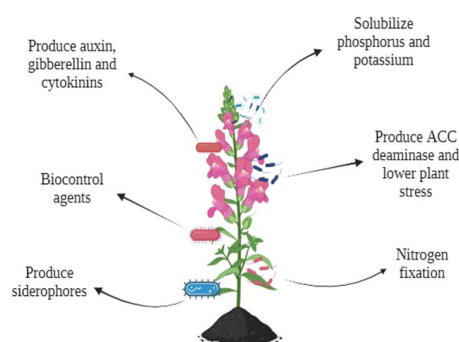


Fig. 2. Mechanisms which PGPB use to promote the growth of horticultural plants. (Figure created with Biorender.com)

plant morphology.

PGPB-synthesized phytohormones, particularly auxins and cytokinins, play crucial roles in modulating various aspects of plant reproductive development. These bacterial-derived hormones influence multiple stages of floral ontogeny, including the initiation of flowering, the emergence and differentiation of floral primordia and the subsequent development and maturation of floral organs (15, 16). Since floral transition and blooming involve complex interactions between different phytohormones, strategic modulation of hormone levels by PGPB inoculations can stimulate early flowering and enhance flower productivity in ornamentals (17, 18). Specific PGPB strains can finetune the auxin: cytokinin balance to achieve desired effects on flowering time and yield in target flower crops.

PGPB are also sources of gibberellin which stimulate internodal and stem elongation, flowering and flower development. For instance, significant increase in endogenous gibberellin content leading to improved flowering was noted in rose inoculated with *Bacillus pumilus* (19). Ethylene is involved in initiation of corolla wilting and senescence. Many PGPB induce ACC deaminase which degrades the immediate precursor of ethylene biosynthesis thereby delaying senescence of flowers (20). Thus, PGPB have potential for prolonging the display life of cut flowers by lowering ethylene production.

2.1.2. Atmospheric Nitrogen Fixation

Biological Nitrogen Fixation (BNF) by soil and rhizosphere microorganisms converts atmospheric nitrogen into plant-available forms, enhancing nitrogen nutrition and reducing reliance on synthetic fertilizers. This process, primarily performed by diazotrophic bacteria and cyanobacteria, promotes growth and flowering in plants (21). Studies have demonstrated that nitrogen-fixing PGPB isolated from flower crop rhizospheres, when applied as inoculants, significantly increase plant nitrogen content and improve growth and flowering parameters. Beneficial effects have been observed in various ornamental plants, including orchids (*Herbaspirillum frisingense* and *Burkholderia* sp.) (22), roses (*Azospirillum amazonense*) (23), petunias and chrysanthemums (*Rhodopseudomonas* sp.) (24, 25) and gerberas and tuberose (*Rhizobium* and *Azotobacter* spp.) (26, 27). The supplemental nitrogen provided by these PGPB enhances overall plant performance, particularly in terms of flowering quality and quantity.

2.1.3. Phosphorus Solubilization

Phosphorus is an essential macronutrient influencing many growth and metabolic processes in plants. Most soil phosphorus exists as insoluble phosphates, which plants cannot directly absorb (28, 29). Phosphate-solubilizing bacteria convert inorganic phosphates into plant-usable forms by secreting organic acids, which chelate cations and lower pH, making phosphates soluble.

Phosphate-solubilizing PGPB have been identified in the rhizosphere of various ornamental plants, including species from genera such as *Bacillus* and *Pseudomonas* (30-33). By enhancing phosphorus nutrition, these PGPB

improve growth and flowering attributes in ornamentals, leading to benefits like early blooming and increased flower production. These positive effects are directly linked to the increase in soil available phosphorus facilitated by the bacterial activity.

2.1.4. 1, 3-Propanediol and Volatile Organic Compounds

Some PGPB like *Bacillus subtilis* and *B. amyloliquefaciens* produce 1, 3-propanediol (1, 3-PD) which elicits induced systemic tolerance against abiotic stresses in plants (34). 1, 3-PD can penetrate plant cell walls and activate stress defence responses. This helps protect flowers against environmental stresses like high temperature and drought.

Several *Bacillus* and *Pseudomonas* spp. also emit Volatile Organic Compounds (VOCs) such as acetoin, 2, 3-butanediol, 2-methyl-n-1-tridecene and lipopeptides which trigger growth promoting effects in plants (11, 35). The VOCs may activate phytohormone signalling pathways leading to enhanced flowering and flower development in commercial flower crops as observed in crops treated with VOC-producing strains.

2.1.5. Induction of Systemic Tolerance

PGPB are known to induce systemic tolerance against biotic and abiotic stresses in plants through priming of various cellular defence responses (36). This mechanism of induced systemic resistance (ISR) operates through pathways mediated by growth regulators such as jasmonic acid, ethylene and salicylic acid. Exposure to beneficial PGPB confers stress resilience in plants.

In flower crops, PGPB treatments have been shown to elicit key protective responses, including the activation of antioxidant enzymes, upregulation of stress-responsive genes and accumulation of compatible solutes (37). These metabolic changes enhance the ability of flowers to withstand environmental fluctuations, thereby extending their longevity and display life. By inducing systemic tolerance, PGPB can thus improve floral traits related to shelf life and postharvest quality.

2.1.6. Antagonism against Pathogens

Fungal diseases represent a significant threat in flower cultivation, leading to substantial economic losses. Pathogens such as *Fusarium*, *Rhizoctonia*, *Phytophthora*, *Botrytis* and powdery mildews adversely impact the productivity and vase life of flower crops. PGPB suppress these pathogenic microbes through various mechanisms viz. production of antibiotics, cell wall degrading enzymes, siderophores and induction of systemic resistance in plants (38, 39).

Biocontrol PGPB with antifungal properties have been isolated from rose (*Bacillus amyloliquefaciens*) (40), gerbera (*Bacillus subtilis*) (41), carnation (*Pseudomonas aeruginosa*, *P. fluorescens*) (42) and orchids (*Bacillus* sp., *Burkholderia* sp., *Pseudomonas* sp.) (43). Inoculations with these PGPB controlled common fungal diseases and improved flower quality in the respective crops. Exploiting such biocontrol traits of PGPB is a sustainable strategy for mitigating the biotic stresses in commercial flower

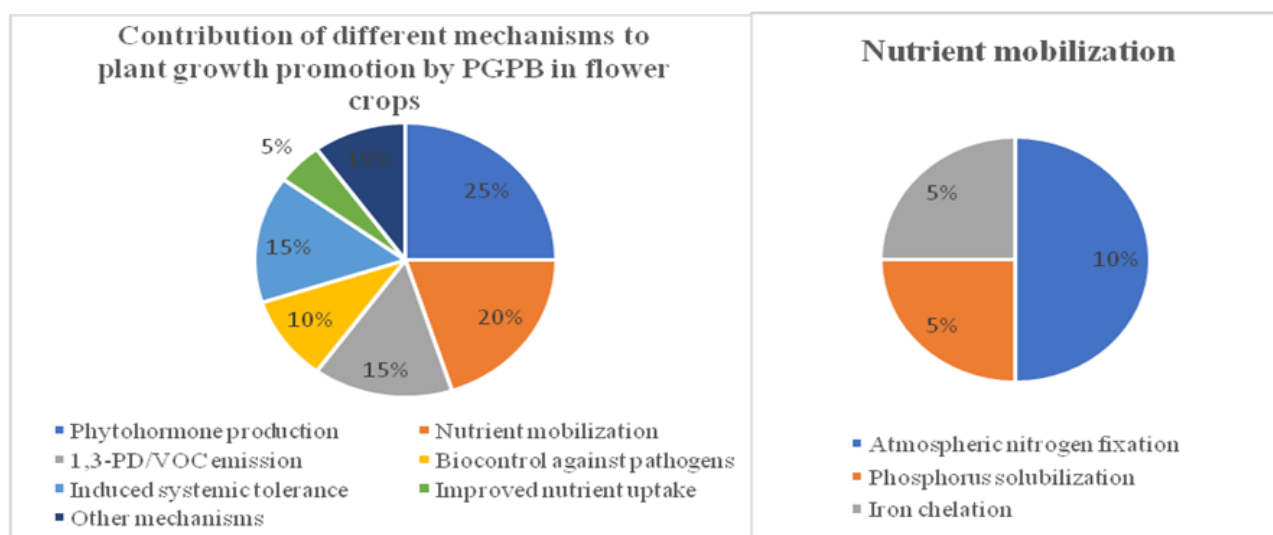


Fig. 3. Contribution of different mechanisms to plant growth promotion by PGPB in flower crops.

cultivation (Fig. 3)

3. PGPB in Major Flower Crops

Plant Growth-Promoting Bacteria (PGPB) have demonstrated significant benefits across various ornamental flower crops, including roses, chrysanthemums, gerberas, orchids and gladiolus. These beneficial bacteria enhance multiple aspects of flower production and quality through various mechanisms.

3.1. Roses

PGPB strains such as *Bacillus pumilus* INR7 (44), *Pseudomonas putida* PW15 (45), *Burkholderia phytofirmans* PsJN (46) and *Azospirillum amazonense* (23) have shown remarkable effects on rose cultivation. These bacteria improved flower yield, increased the number of flowers per plant, extended flowering duration and enhanced postharvest longevity. They also boosted nutrient uptake, chlorophyll content and induced systemic tolerance against stresses (23, 44-47).

3.2. Chrysanthemum

For chrysanthemums, PGPB strains including *Pseudomonas chlororaphis* HT66 (48), *Rhodopseudomonas* sp. CQ001 (25) and *Bacillus subtilis* AU195 (46) have proven effective. These bacteria increased flower diameter, prolonged vase life, enhanced flower numbers and yield and augmented plant biomass. They achieved these improvements through mechanisms such as induced systemic tolerance, improved nutrient fixation and solubilization and production of growth-promoting compounds (25, 48-50).

3.3. Gerbera

Gerbera cultivation benefited from PGPB strains like *Bacillus subtilis* RZ19 (31), *Azotobacter chroococcum* Az3 (26) and *Pseudomonas aeruginosa* EF121 (51). These bacteria increased flower numbers and size, reduced disease incidence and extended shelf life. They functioned through various mechanisms, including pathogen antagonism, nitrogen fixation and induction of systemic resistance (26, 31, 51, 52).

3.4. Orchids

In orchid farming, PGPB strains such as *Bacillus subtilis* OH 131.1 (22), *Burkholderia* sp. F-390 (53) and *Pseudomonas oryzihabitans* (54) showed promising results. These bacteria increased spike length, boosted flower numbers and reduced disease incidence. They achieved these improvements through pathogen antagonism, enhanced nutrient uptake and production of growth-promoting compounds (22, 53-55).

3.5. Gladiolus

Gladiolus cultivation was enhanced by PGPB strains including *Pseudomonas fluorescens* PF-08 (56), *Bacillus* sp. EU-1 (57) and *Azotobacter chroococcum* (58). These bacteria increased spike length, number of florets, corm weight and cut flower longevity. Their beneficial effects were attributed to various plant growth-promoting traits such as nutrient solubilization, nitrogen fixation and phytohormone production (56-58).

3.6. Other Flower crops

Bacillus megaterium augmented growth, early flowering, number of flowers and phosphorus uptake in marigold (30). *Azospirillum lipoferum* and *B. coagulans* increased the growth and flower yield in tuberose (27). *Pseudomonas* sp. strain P45 and *Bacillus* sp. strain EPB10 enhanced growth, flower size and postharvest life of lilies (59, 60). *Pseudomonas putida* strain PW12 induced early flowering, increased flower size and shelf life in carnation (61). *Bacillus* sp. strains IN937b and WB800N stimulated growth and flowering in *Petunia* and *Gaillardia* (24, 62). *Klebsiella variicola* improved flowering behavior and flower quality traits in pansy (63). Thus, PGPB confer beneficial effects on productivity and quality across the diverse ornamentals.

In conclusion, across these diverse ornamental crops, PGPB consistently demonstrated the ability to enhance flower yield, quality and longevity while also improving plant health and stress tolerance. These findings highlight the significant potential of PGPB as a sustainable approach to improving ornamental flower

Table 1. Effect of PGPB inoculations on growth and flowering parameters in major ornamental plants.

Flower Crop	PGPB Strain	Key Effects	References
Rose	<i>Bacillus pumilus</i> INR7	↑ Flower yield by 29 % ↑ NPK uptake	(44)
Chrysanthemum	<i>Pseudomonas chlororaphis</i> HT66	↑ Flower diameter ↑ Vase life by 2.7 days	(48)
Gerbera	<i>Bacillus subtilis</i> RZ19	↑ Number of flowers by 37.5 % ↓ Disease incidence by 42 %	(34)
Orchids	<i>Bacillus subtilis</i> OH 131.1	↑ Spike length by 28 % ↑ Number of flowers by 42 %	(22)
Gladiolus	<i>Pseudomonas fluorescens</i> PF-08	↑ Spike length by 19 % ↑ Vase life by 2 days	(56)

production (Table 1).

4. Commercial products of PGPB in Floriculture

Research on PGPB has paved the way for developing microbial bio stimulant products and bioinoculants for application in floriculture. Some examples of commercially available PGPB-based formulations for flower crops are presented (Table 2 and 3).

5. Limitations in PGPB application

A critical analysis of current research on Plant Growth-Promoting Bacteria (PGPB) in floriculture reveals several limitations that must be addressed to successfully

translate laboratory findings to field applications. Studies on various ornamental crops, including roses (44-46), chrysanthemums (48, 49), gerberas (31, 51), orchids (22, 53, 54) and gladiolus (56-58), often occur under controlled conditions, potentially misrepresenting field performance. Additionally, most experiments lack long-term efficacy assessments and fail to adequately address PGPB interactions with indigenous soil microorganisms. Optimization of formulations and concentrations for different flower crops remains a challenge, despite promising results with specific strains (23, 25). The lack of standardized protocols and quality regulations further

Table 2. Some of the commercial PGPB products used in floriculture sector (64, 65).

Product	Manufacturer	Composition	Application
Rhizoflow FP	Rizobacter, Argentina	<i>Bacillus amyloliquefaciens</i>	Improving growth and flowering in ornamentals
Blünger	Sourcon Padena, Germany	<i>Pseudomonas</i> sp., <i>Bacillus</i> sp.	Shelf-life enhancer for cut flowers
Quantum Vita	Anthurium Biotech, Mauritius	<i>Bacillus megaterium</i>	Biofertilizer for anthurium
Radix WP	New Bio-Products, South Africa	<i>Bacillus subtilis</i>	Biofungicide and growth promoter for ornamentals
NutriLife PB	Terramera, Canada	<i>Bacillus</i> sp.	Stimulate flowering and yield in flowers
EcoVita	Mapleton Agri Biotec, Australia	<i>Azospirillum brasilense</i>	Biofertilizer to improve growth and nitrogen uptake in ornamentals
PhosNfix	Novozymes, Denmark	<i>Penicillium bilaiae</i>	Enhance nutrient availability in flowering plants
Zhongkemairui	Guangdong VTR Bio-Tech Co., China	<i>Bacillus subtilis</i> , yeast extracts	Increase yield in chrysanthemum
IronMaxx	BioWorks, USA	<i>Pseudomonas putida</i>	Prevent iron deficiency in potted flowering plants
Serifel	BASF, Germany	<i>Serratia plymuthica</i>	Stimulate flowering in ornamentals and vegetables
Florgib	Valagro, Italy	Gibberellic acid producing bacteria	Increase size and number of flowers

Table 3. India based PGPB products (66-69).

Product	Manufacturer	In India	
		Composition	Application
Biotor Plus	Mapri Bioculture Pvt. Ltd.	<i>Azotobacter</i> , <i>Azospirillum</i> , <i>Pseudomonas</i> , <i>Bacillus</i>	Ornamental crops for plant growth, nutrient uptake, disease resistance
Bio-Vert	Bharat Biotech International Ltd.	<i>Bacillus subtilis</i> , <i>Bacillus licheniformis</i> , <i>Pseudomonas fluorescens</i>	Ornamental crops like roses, carnations, orchids for plant growth and biocontrol
BioPro	Growrich Biotechnologies Pvt. Ltd.	<i>Bacillus subtilis</i> , <i>Bacillus amyloliquefaciens</i> , <i>Pseudomonas putida</i>	Ornamental plants for nutrient uptake, root development, plant vigor
Biomax	Multiplex Biotech Pvt. Ltd.	<i>Azotobacter</i> , <i>Azospirillum</i> , <i>Pseudomonas</i>	Ornamental crops for plant growth, nutrient availability, stress tolerance
Biogreen Plus	Novozymes South Asia Pvt. Ltd.	<i>Bacillus amyloliquefaciens</i> , <i>Trichoderma</i>	Ornamental plants like roses, chrysanthemums, gerberas for root growth, nutrient uptake, plant vigor

hinders result comparisons and product quality assurance.

To address these limitations, future research should prioritize long-term field trials under diverse conditions, PGPB-native microbiome interaction studies, development of PGPB consortia and exploration of site-specific strains. Standardization of PGPB formulation, application methods and quality control measures is crucial, as are economic viability studies in commercial settings. Advancing PGPB from laboratory to field applications in floriculture will require interdisciplinary collaboration, public-private partnerships and supportive policies. By addressing these challenges, the full potential of PGPB in enhancing flower crop production and quality can be realized, paving the way for more sustainable and efficient floriculture practices.

6. Future perspective

Advancing plant growth-promoting bacteria (PGPB) applications in floriculture requires focused research in key areas. These include identifying flower crop-specific PGPB strains, elucidating PGPB-ornamental plant molecular interactions and developing synergistic PGPB consortia. Integration with precision agriculture and microbiome engineering, assessment of long-term ecological impacts and optimization of field application and delivery systems are also crucial. Successful implementation will necessitate collaboration between researchers, industry stakeholders and policymakers. By concentrating on these critical aspects, the floriculture sector can leverage PGPB to enhance sustainability and efficiency in global commercial flower production.

Conclusion

Commercial flower cultivation is an economically important yet resource-intensive agricultural sector. Excessive reliance on agrochemical inputs in intensive floriculture can cause environmental degradation and pollution. Plant growth-promoting bacteria (PGPB) offer an eco-friendly and sustainable solution to enhance productivity, quality, and stress resilience of diverse flower crops. This review documented significant benefits of PGPB inoculations across major ornamentals like rose, chrysanthemum, gerbera, orchids and gladiolus. Key mechanisms employed by PGPB include nutrient mobilization, modulation of phytohormones, induction of systemic tolerance and biocontrol of diseases. Several PGPB-based commercial products are available as biofertilizers, bio stimulants and biopesticides for the floriculture industry. Future research should prioritize long-term field trials, PGPB-native microbiome interactions, development of crop-specific consortia, optimization of formulations and application methods, economic and environmental impact assessments and integration with precision agriculture techniques. Addressing these areas will facilitate the transition towards sustainable, environmentally responsible commercial flower production systems with minimized ecological impacts.

Acknowledgements

We would like to express gratitude to department of floriculture and landscaping (Botanical Garden) and department of agricultural microbiology. Special thanks are extended to the Chairman, Dean (HC&RI) and advisory members for their valuable feedback and constructive suggestions on the manuscript.

Authors' contributions

MAM conducted literature search and data extraction, analysed and interpreted the compiled information. SK conceptualized the review topic, provided guidance on the review process and approved the final manuscript. PIV and SU developed key ideas for the review, critically reviewed the manuscript and approved the final manuscript. RJ assisted in summarizing and revising the manuscript. MD contributed to manuscript editing, summarization and revision.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interests to declare.

Ethical issues: None.

Declaration of generative AI and AI-assisted technologies in the writing process

During the preparation of this work the author(s) not used AI tools and the author(s) reviewed and edited the content as needed and take(s) full responsibility for the content of the publication.

References

1. Van der Star G, A. van Klink, Florensis: A company willing to take responsibility. 2022;1-22. <https://doi.org/10.21427/w2jm-k018>
2. Anumala, Nikhila Vaagdevi, Ranjit Kumar. Floriculture sector in India: current status and export potential. *The Journal of Horticultural Science and Biotechnology*. 2021;96:673-80. <https://doi.org/10.1080/14620316.2021.1902863>
3. Misra, Debajit, Sudip Ghosh. Growth and export status of Indian floriculture: A review. *Agricultural Reviews*. 2016;37:77-80. <https://doi.org/10.18805/ar.v37i1.9269>
4. Malviya, Aditi, et al. Recent floriculture in India. *International Association of Biologicals and Computational Digest*. 2022; <https://doi.org/10.56588/iabcd.v1i1.8>
5. Khabbaz SE, et al. Plant growth promoting bacteria (PGPB)- A versatile tool for plant health management. *Can J Pestic Pest Manag.* 2019;1(1):1-25. <https://doi.org/10.34195/can.j.ppm.2019.05.001>
6. Glick BR. The enhancement of plant growth by free-living bacteria. *Canadian Journal of Microbiology*. 1995;41(2):109-17. <https://doi.org/10.1139/m95-015>
7. Ojuederie OB, OS Olanrewaju, OO Babalola. Plant growth promoting rhizobacterial mitigation of drought stress in crop plants: Implications for sustainable agriculture. *Agronomy*. 2019;9(11):712. <https://doi.org/10.3390/agronomy9110712>
8. Bhattacharyya PN, DK Jha. Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. *World Journal of Microbiology*

- and Biotechnology. 2011;28:1327-350. <https://doi.org/10.1007/s11274-011-0979-9>
9. Kaymak HÇ. Potential of PGPR in agricultural innovations. 2010. https://doi.org/10.1007/978-3-642-13612-2_3
 10. Wang Yuean, et al. Transcriptome analysis of growth and quality response of chrysanthemum to co-inoculation with *Bacillus velezensis* and *Pseudomonas aeruginosa*. Scientia Horticulturae. 2024; <https://doi.org/10.1016/j.scienta.2023.112722>
 11. Fincheira Paola, Andrés Quiroz. Microbial volatiles as plant growth inducers. Microbiological Research. 2018;208:63-75. <https://doi.org/10.1016/j.micres.2018.01.002>
 12. Cassán F, J Vanderleyden, S Spaepen. Physiological and agronomical aspects of phytohormone production by model plant-growth-promoting rhizobacteria (PGPR) belonging to the genus *Azospirillum*. Journal of Plant Growth Regulation. 2014;33:440-59. <https://doi.org/10.1007/s00344-013-9362-4>
 13. Orozco-Mosqueda M.d.C, G Santoyo, BR Glick. Recent advances in the bacterial phytohormone modulation of plant growth. Plants. 2023;12(3):606. <https://doi.org/10.3390/plants12030606>
 14. Spaepen S. Plant hormones produced by microbes, in principles of plant-microbe interactions: microbes for sustainable agriculture. Springer. 2014;247-56. https://doi.org/10.1007/978-3-319-08575-3_26
 15. Yamaguchi N, et al. A molecular framework for auxin-mediated initiation of flower primordia. Developmental Cell. 2013;24(3):271-82. <https://doi.org/10.1016/j.devcel.2012.12.017>
 16. Heisler MG, et al. Patterns of auxin transport and gene expression during primordium development revealed by live imaging of the *Arabidopsis* inflorescence meristem. Current Biology. 2005;15(21):1899-911. <https://doi.org/10.1016/j.cub.2005.09.052>
 17. Meijón M, et al. Epigenetic and physiological effects of gibberellin inhibitors and chemical pruners on the floral transition of azalea. Physiologia Plantarum. 2011;141(3):276-88. <https://doi.org/10.1111/j.1399-3054.2010.01430.x>
 18. Kurepin LV, et al. Burkholderia phytofirmans-induced shoot and root growth promotion is associated with endogenous changes in plant growth hormone levels. Plant Growth Regulation. 2014;75:199-07. <https://doi.org/10.1007/s10725-014-9944-6>
 19. Joo GJ, et al. Gibberellins-producing rhizobacteria increase endogenous gibberellins content and promote growth of red peppers. The Journal of Microbiology. 2005;43(6):510-15.
 20. Nadeau JA, et al. Temporal and spatial regulation of 1-aminocyclopropane-1-carboxylate oxidase in the pollination-induced senescence of orchid flowers. Plant Physiology. 1993;103(1):31-39. <https://doi.org/10.1104/pp.103.1.31>
 21. Aasfar A, et al. Nitrogen fixing *Azotobacter* species as potential soil biological enhancers for crop nutrition and yield stability. Frontiers in Microbiology. 2021;12:628379. <https://doi.org/10.3389/fmicb.2021.628379>
 22. Gontijo JB, et al. Bioprospecting and selection of growth-promoting bacteria for *Cymbidium* sp. orchids. Scientia Agricola. 2018;75:368-74. <https://doi.org/10.1590/1678-992x-2017-0117>
 23. Aguirre-Medina JF, et al. Tabebuia donnell-smithii Rose growth inoculated with mycorrhizal fungi and *Azospirillum brasilense*. Agrociencia. 2014;48(3). <https://www.scielo.org.mx/pdf/agro/v48n3/v48n3a8.pdf>
 24. Dixon R. Genetic strategies for improved nitrogen fixation and utilization by crops. Biochemical Aspects of Crop Improvement. 1990;359. <https://www.cabidigitallibrary.org/doi/full/10.5555/19931697262>
 25. Chandran H, M Meena, P Swapnil. Plant growth-promoting rhizobacteria as a green alternative for sustainable agriculture. Sustainability. 2021;13(19):10986. <https://doi.org/10.3390/sul131910986>
 26. Anop Kumari AK, MC Mahesh Choudhary, SB Satpal Baloda. Biofertilizers and their application in flower crops- a review. 2013. <https://www.cabidigitallibrary.org/doi/full/10.5555/20133247803>
 27. Chaudhary S. Biofertilizers and their application in floriculture- A review. Annals of Horticulture. 2010;3(1):29-33. <https://www.indianjournals.com/ijor.aspx?target=ijor:ah&volume=3&issue=1&article=004>
 28. Pradhan N, L Sukla. Solubilization of inorganic phosphates by fungi isolated from agriculture soil. African Journal of Biotechnology. 2006;5(10). <https://www.ajol.info/index.php/ajb/article/view/42884>
 29. Kafle A, et al. Harnessing soil microbes to improve plant phosphate efficiency in cropping systems. Agronomy. 2019. <https://doi.org/10.3390/agronomy9030127>
 30. Zhao Y, et al. Phosphate-solubilising bacteria promote horticultural plant growth through phosphate solubilisation and phytohormone regulation. New Zealand Journal of Crop and Horticultural Science. 2022;1-16. <https://doi.org/10.1080/01140671.2022.2103156>
 31. Devindrappa M, et al. Nematicidal rhizobacteria with plant growth-promoting traits associated with tomato in root-knot infested polyhouses. Egyptian Journal of Biological Pest Control. 2022;32(1):51. <https://doi.org/10.1186/s41938-022-00539-1>
 32. Wang T, MQ Liu, HX Li. Inoculation of phosphate-solubilizing bacteria *Bacillus thuringiensis* B1 increases available phosphorus and growth of peanut in acidic soil. Acta Agriculturae Scandinavica. Section B-Soil and Plant Science. 2014;64(3):252-59. <https://doi.org/10.1080/09064710.2014.905624>
 33. Garcia-Ochoa E, et al. *In vitro* biological activity of metabolic extracts of wild and cultivated species of the genus *Polygonum*. In: IX International Symposium on New Ornamental Crops; 2019. 1288. <https://doi.org/10.17660/ActaHortic.2020.1288.22>
 34. Choudhary DK, et al. Bacterial-mediated tolerance and resistance to plants under abiotic and biotic stresses. Journal of Plant Growth Regulation. 2016;35:276-300. <https://doi.org/10.1007/s00344-015-9521-x>
 35. Park YS, et al. Promotion of plant growth by *Pseudomonas fluorescens* strain SS101 via novel volatile organic compounds. Biochemical and Biophysical Research Communications. 2015;461(2):361-65. <https://doi.org/10.1016/j.bbrc.2015.04.039>
 36. Maksimov I, et al. Plant growth-promoting bacteria in regulation of plant resistance to stress factors. Russian Journal of Plant Physiology. 2015;62:715-26. <https://doi.org/10.1134/S1021443715060114>
 37. Kohli SK, et al. Interaction of 24-epibrassinolide and salicylic acid regulates pigment contents, antioxidative defense responses and gene expression in *Brassica juncea* L. seedlings under Pb stress. Environmental Science and Pollution Research. 2018;25:15159-173. <https://doi.org/10.1007/s11356-018-1742-7>
 38. Gamalero E, BR Glick. Mechanisms used by plant growth-promoting bacteria. Bacteria in Agrobiology: Plant Nutrient Management. 2011;17-46. https://doi.org/10.1007/978-3-642-21061-7_2
 39. Meena M, et al. PGPR-mediated induction of systemic resistance and physiochemical alterations in plants against the pathogens: Current perspectives. Journal of Basic Microbiology. 2020;60:828-61. <https://doi.org/10.1002/jobm.202000370>
 40. Nakkeeran S, T Surya, S Vinodkumar. Antifungal potential of plant growth promoting *Bacillus* species against blossom blight of rose. Journal of Plant Growth Regulation. 2020;39:99-111. <https://doi.org/10.1007/s00344-019-09966-1>
 41. Suneeta P, KE Arutkani Aiyathan, S. Nakkeeran. Investigations on antifungal activity of *Bacillus* spp. against *Fusarium oxysporum* f. sp. gerberae (FOG) causing wilt of gerbera under protected cultivation. Journal of Pure and Applied Microbiology.

- 2016;10(3). <https://www.cabidigitallibrary.org/doi/full/10.5555/20173288469>
42. Kaur M, et al. Plant growth promoting properties of *Pseudomonas* species from carnation and medicinal plants. *Progressive Horticulture*. 2010;42(2):148-56. <https://www.indianjournals.com/ijor.aspx?target=ijor:pho&volume=42&issue=2&article=005>
 43. Herrera H, et al. Orchid-associated bacteria and their plant growth promotion capabilities. In: *Orchids Phytochemistry, Biology and Horticulture: Fundamentals and Applications*. Springer;2022:175-200. https://doi.org/10.1007/978-3-030-38392-3_35
 44. Lenin G, M Jayanthi. Efficiency of plant growth promoting rhizobacteria (PGPR) on enhancement of growth, yield and nutrient content of *Catharanthus roseus*. *Int J Res Pure Appl Microbiol*. 2012;2(4):37-42.
 45. Yagi M, M Eldeen, N Elgemaby. Effect of bactericides and sucrose pulsing on longevity and vase life of rose cut flowers. *International Journal of Sciences: Basic and Applied Research*. 2014;14:117-29.
 46. Wang B, C Mei, JR Seiler. Early growth promotion and leaf level physiology changes in *Burkholderia phytofirmans* strain PsJN inoculated switchgrass. *Plant Physiology and Biochemistry*. 2015;86:16-23. <https://doi.org/10.1016/j.plaphy.2014.11.008>
 47. Canli F, S Kazaz. Biotechnology of roses: progress and future prospects. *Turkish Journal of Forestry*. 2009;10(1):167-83. <https://dergipark.org.tr/en/pub/tjf/issue/20892/224325>
 48. Tanwar A, AA Karishma. Growth and flower enhancement of *Chrysanthemum indicum* on inoculation with *arbuscular mycorrhizae* and other bioinoculants. 2013. <https://agris.fao.org/search/en/providers/122648/records/64747342bf943c8c7982fe51>
 49. Adesemoye A, M Obini, E Ugoji. Comparison of plant growth-promotion with *Pseudomonas aeruginosa* and *Bacillus subtilis* in three vegetables. *Brazilian Journal of Microbiology*. 2008;39:423-26. <https://doi.org/10.1590/S1517-83822008000300003>
 50. Hebbal M, et al. Genotypic and phenotypic path analysis for flower yield in chrysanthemum (*Dendranthema grandiflora* Tzvelve). *Int J Curr Microbiol App Sci*. 2018;7(8):4515-21. <https://doi.org/10.20546/ijcmas.2018.708.478>
 51. Prashanth P, RC Sekhar, KCS Reddy. Influence of floral preservatives on scape bending, biochemical changes and post-harvest vase life of cut gerbera (*Gerbera jamesonii* Bolus ex. Hook.). *Asian Journal of Horticulture*. 2010;5(1):1-6. <https://www.cabidigitallibrary.org/doi/full/10.5555/20103272018>
 52. Chauhan R, et al. Effect of different media on growth, flowering and cut flower yield of gerbera under protected condition. *Asian Journal of Horticulture*. 2014;9(1): 228-31. <https://doi.org/10.15740/HAS/TAJH/9.2/404-407>
 53. Te-Chato S, P Nujeen, S Muangsorn. Paclobutrazol enhance bud break and flowering of Friederick's Dendrobium orchid *in vitro*. *Journal of Agricultural Technology*. 2009;5(1):157-65. <https://www.thaiscience.info/Journals/Article/IJAT/10842918.pdf>
 54. Kerbauy GB. *In vitro* flowering of *Oncidium varicosum* mericlones (Orchidaceae). *Plant Science Letters*. 1984;35(1):73-75. [https://doi.org/10.1016/0304-4211\(84\)90160-3](https://doi.org/10.1016/0304-4211(84)90160-3)
 55. Jain A, et al. Changes in global Orchidaceae disease geographical research trends: recent incidences, distributions, treatment and challenges. *Bioengineered*. 2021;12(1):13-29. <https://doi.org/10.1080/21655979.2020.1853447>
 56. Hassan F, M Fetouh. Does moringa leaf extract have preservative effect improving the longevity and postharvest quality of gladiolus cut spikes? *Scientia Horticulturae*. 2019;250:287-93. <https://doi.org/10.1016/j.scienta.2019.02.059>
 57. Damodaran T, et al. Rhizosphere and endophytic bacteria for induction of salt tolerance in gladiolus grown in sodic soils. *Journal of Plant Interactions*. 2014;9(1):577-84. <https://doi.org/10.1080/17429145.2013.873958>
 58. Shanmugam V, et al. Biocontrol of vascular wilt and corm rot of gladiolus caused by *Fusarium oxysporum* f. sp. gladioli using plant growth promoting rhizobacterial mixture. *Crop Protection*. 2011;30(7):807-13. <https://doi.org/10.1016/j.cropro.2011.02.033>
 59. Upadhyay A, S Srivastava. Evaluation of multiple plant growth promoting traits of an isolate of *Pseudomonas fluorescens* strain Psd. 2010. <https://nopr.niscpr.res.in/handle/123456789/9077>
 60. Li B, et al. Genomics assisted functional characterization of *Bacillus velezensis* E as a biocontrol and growth promoting bacterium for lily. *Frontiers in Microbiology*. 2022;13:976918. <https://doi.org/10.3389/fmicb.2022.976918>
 61. Cipriano MA, SS Freitas. Effect of *Pseudomonas putida* on chrysanthemum growth under greenhouse and field conditions. *African Journal of Agricultural Research*. 2018;13(6):302-10. <https://doi.org/10.5897/AJAR2017.12839>
 62. Kulkova I, et al. Plant growth promotion using *Bacillus cereus*. *International Journal of Molecular Sciences*. 2023;24(11):9759. <https://doi.org/10.3390/ijms24119759>
 63. Kusale SP, et al. Inoculation of *Klebsiella variicola* alleviated salt stress and improved growth and nutrients in wheat and maize. *Agronomy*. 2021;11(5):927. <https://doi.org/10.3390/agronomy11050927>
 64. Ipek M, et al. Plant growth-promoting rhizobacteria (PGPR) increase yield, growth and nutrition of strawberry under high-calcareous soil conditions. *Journal of Plant Nutrition*. 2014;37(7):990-01. <https://doi.org/10.1080/01904167.2014.881857>
 65. Menéndez E, et al. Rhizobial biofertilizers for ornamental plants. In: *Biological Nitrogen Fixation and Beneficial Plant-Microbe Interaction*. Springer; 2016. https://doi.org/10.1007/978-3-319-32528-6_2
 66. Yadav SK, et al. Co-inoculated biopriming with *Trichoderma*, *Pseudomonas* and *Rhizobium* improves crop growth in *Cicer arietinum* and *Phaseolus vulgaris*. *International Journal of Agriculture, Environment and Biotechnology*. 2013;6:255-59. <https://www.indianjournals.com/ijor.aspx?target=ijor:ijaeb&volume=6&issue=2&article=010>
 67. Lyubenova A, et al. Plant extracts and *Trichoderma* spp.: possibilities for implementation in agriculture as biopesticides. *Biotechnology and Biotechnological Equipment*. 2023;37:159-66. <https://doi.org/10.1080/13102818.2023.2166869>
 68. Sumbul A, et al. Azotobacter: A potential bio-fertilizer for soil and plant health management. *Saudi Journal of Biological Sciences*. 2020;27:3634-40. <https://doi.org/10.1016/j.sjbs.2020.08.004>
 69. Maheshwari DK, et al. Carrier based formulations of biocontrol consortia of disease suppressive *Pseudomonas aeruginosa* KRP1 and *Bacillus licheniformis* KRB1. *Ecological Engineering*. 2015;81:272-77. <https://doi.org/10.1016/j.ecoleng.2015.04.066>