

RESEARCH ARTICLE



Nano-diammonium phosphate enhances grain yield by modulating gas exchange traits and nutrient use efficiency in rice

R Roshil Ben Sweety¹, P Boominathan^{1*}, A Senthil¹, D Jegadeeswari², G Prabu Kumar³

¹Department of Crop Physiology, Tamil Nadu Agricultural University, Coimbatore, 641 003, Tamil Nadu, India ²Department of Soil Science and Agricultural Chemistry, Tamil Nadu Agricultural University, Coimbatore 641 003, Tamil Nadu, India ³Department of Agronomy, Tamil Nadu Agricultural University, Coimbatore, 641 003, Tamil Nadu, India

* Email: boominathan.p@tnau.ac.in

ARTICLE HISTORY

Received: 03 October 2024 Accepted: 02 November 2024 Available online Version 1.0: 28 December 2024

Check for updates

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/

journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/ index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an openaccess article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (https://creativecommons.org/licenses/ by/4.0/)

CITE THIS ARTICLE

Sweety RRB, Boominathan P, Senthil A, Jegadeeswari D, Kumar GP. Nanodiammonium phosphate enhances grain yield by modulating gas exchange traits and nutrient use efficiency in rice. Plant Science Today.2024;11(sp4):01-10. https://doi.org/10.14719/pst.5508

Abstract

Nutrients are primarily applied to crops through the soil to enhance productivity while maintaining crop and soil health. However, high doses of soil -applied nutrients often reduce nutrient use efficiency, increase cultivation costs, and contribute to environmental pollution. With their high surface areato-volume ratio, Nano-formulated nutrients have emerged as effective alternatives, requiring lower doses and demonstrating superior nutrient use efficiency (NUE) compared to conventional fertilizers. This experiment was conducted to study the influence of Nano-Diammonium Phosphate (Nano-DAP) on physiological and yield-associated traits in rice and to assess the extent of replacement of soil application of nitrogen(N) and phosphorous fertilizers with foliar application of nano-form of DAP. Field trials included nine treatments, comprising 50% and 75% of the recommended dose of nitrogen and phosphorus (RDNP) and 100% of the recommended dose of fertilizers (RDF), and foliar application of conventional and nano-DAP. Foliar treatments were applied during the active tillering and panicle initiation stages. Among the treatments, 75% RDF and two foliar sprays of Nano-DAP performed better in terms of morpho-physiological parameters, particularly leaf area, dry matter production, photosynthetic rate, stomatal conductance, and number of productive tillers compared to 100% RDF. Furthermore, the NUE of Nano-DAP treatments was significantly higher, reflecting an improved ability of the rice plants to utilize the nanoform nutrients, resulting in enhanced yield effectively.

Keywords

nano-DAP; nanofertilizer; nutrient use efficiency; phosphorus use efficiency

Introduction

Nutrient requirements of crops are primarily met through the soil application of high doses of conventional fertilizers. However, this practice reduces soil fertility, damages the flora, and causes soil erosion, drastically reducing NUE. Among the essential nutrients, N and phosphorus (P) are significant elements required for metabolic processes that regulate the growth and development of plants. These two nutrients are crucial for protein synthesis, energy transfer, and root development. While N promotes healthy foliage and overall plant vigor, P is vital for root establishment, flowering, and grain formation, ultimately enhancing crop yield (1). A N deficiency can cause stunted growth, reduced chlorophyll content, and poor yields due to its role in protein synthesis and chlorophyll formation (2). Although N is abundant in the atmosphere, only

a small portion is accessible to plants in nitrate or ammonium, necessitating careful management to ensure adequate nutrient supply (3). Insufficient P can result in plant membrane breakdown and hinder energy transfer within the plant (4). Although soils may hold P reserves greater than what plants need, only a small fraction of P is soluble and accessible for plant uptake (5).

Applying N to soil presents multiple challenges, including inefficient plant uptake and substantial losses through leaching, volatilization, and denitrification. These losses diminish N-fertilizers' effectiveness and exacerbate environmental problems such as water pollution and greenhouse gas emissions (6). P application in soil frequently faces challenges, including limited availability as P binds with soil particles, which impedes plant uptake. Moreover, P losses through runoff and erosion can lead to waterway eutrophication and environmental degradation (7). Hence, applying N and P gradually through foliar application allows plants to absorb and utilize them effectively for metabolic processes.

Improving N and P use efficiency is crucial for maximizing crop productivity and minimizing environmental impact by reducing fertilizer loss and improving plant uptake, leading to better crop performance (8). This issue is addressed through techniques such as split N application, N localization, precision farming, and foliar sprays designed to optimize NUE and reduce environmental concerns (8). Similarly, improving Phosphorus use efficiency (PUE) is essential for addressing environmental issues and decreasing reliance on nonrenewable resources. Methods to enhance PUE, such as using P-solubilizing microorganisms or adopting precision farming techniques, can significantly improve nutrient uptake and crop performance (9).

Most rice cultivation, mainly under wetland conditions, is characterized by applying high N and P, which leads to significant nutrient losses. Nutrient loss in wetland rice cultivation occurs due to leaching, runoff, and volatilization, where N and P are washed away or lost as gases. Additionally, anaerobic conditions promote denitrification and P fixation, reducing plant nutrient availability. In nitrogenous fertilizers, ammonia (NH3) fluxes represent a major N loss pathway, with higher emissions linked to increased N fertilization rates in wetland rice systems (10). In India, rice accounts for approximately 37% of total fertilizer N consumption, with significant losses (60-70%) occurring through various processes (11). Therefore, nano-form fertilizers as a foliar application have been given a major thrust to increase their efficiency. Using nano fertilizers can lower cultivation costs by decreasing the amount of fertilizer needed for cultivation (12) and provide a controlled release of nutrients, reducing losses associated with conventional fertilizers (13). Nano-fertilizers provide controlled nutrient release by encapsulating nutrients in nanomaterials, reducing losses through leaching, volatilization, and denitrification. For example, nano-urea releases N gradually, improving NUE and minimizing environmental impact. This controlled release enhances crop yields while reducing the need for frequent fertilizer applications (14).

mineral fertilizers are required to maintain a continuous nutritional balance in the different growth stages of rice. This approach reduces reliance on chemical fertilizers, fostering a more favorable environment for nutrient absorption. Additionally, foliar application allows for faster nutrient uptake by leaves than root applications, as nutrients are directly available in solution form rather than depending on soil uptake (15). This method also improves nutrient use efficiency, reduces environmental impact, and may offer enhanced health benefits for consumers (16). Moreover, it can be applied even under unfavorable soil and weather conditions. In addition, it allows plants to directly uptake nutrients, reducing fertilizer wastage. Consequently, the foliar application of nanofertilizers is becoming viable for enhancing nutrient use efficiency and boosting crop growth.

Newly developed nano-fertilizers are smaller, possess a larger surface area, and exhibit increased absorption capacity with controlled-release kinetics at specific sites (17). They are highly reactive and can penetrate the plant cuticle, ensuring controlled release and targeted nutrient delivery (18). Indian Farmers Fertiliser Cooperative Limited (IFFCO) is a leading Indian cooperative that manufactures and supplies nanofertilizers to enhance agricultural productivity and significantly supports farmers by providing high-quality and cost-effective fertilizers (19). One recent product of IFFCO is Nano-DAP fertilizer, which offers a cost-effective alternative to conventional DAP aimed to enhance crop yield and provide significant long-term benefits.

Further, the use of this Nano-DAP requires crop-specific dosage optimization compared with the RDF and analyzing rice's response regarding physiological and biochemical traits. Therefore, this experiment was designed to study the response of rice to various amounts of DAP nanofertilizer that alter physiological and biochemical traits, resulting in improved yields.

Materials and Methods

Study area and experimental design

A field experiment was conducted from December 2023 to April 2024 at wetland farms affiliated with Tamil Nadu Agricultural University, Coimbatore, India. The location is at 1° N latitude and 77° E longitude, with an altitude of approximately 426.72 meters above mean sea level. The experimental design employed was a Randomized Block Design (RBD), consisting of nine treatments, each replicated three times. The experimental field covered a total area of 675 m^2 , with dimensions of 53 meters in length and 25 meters in width. The field was divided into 27 plots, each measuring 25 m^2 with a spacing of 20 cm × 20 cm.

Treatment details

We imposed nine treatments with soil application of recommended N and P fertilizers (100%, 75%, and 50%) in which N as four splits and P as basal combined with foliar sprays of 2% DAP and 0.7% Nano-DAP. A uniform application of 100% potassium fertilizer was made in four splits across all the treatments. The stages of application of foliar sprays are active tillering and panicle initiation stage. Observations were

The simultaneous application of traditional and nanoparticle

recorded after the 10th day of the spray. The growth and physiological parameters were recorded 10 days after the first and second sprays.

The rice variety ADT 53 was used for this experiment, and regular cultivation practices were followed except for the fertilizer application. A fertile land with optimal irrigation infrastructure was selected for nursery cultivation, and the main field was properly leveled and well-puddled for transplanting. The soil texture is clay loam with pH of 7.7, electrical conductivity of 0.74 dS m-1, N of 231 kg ha-1, P of 13.9 kg ha-1, and potassium of 295 kg ha-1. Fertilizers were applied as per the treatment schedule, and no inputs were added to the control while the RDF was applied in T1. Control represents 0% N, 0% P and 100% K without DAP and Nano-DAP sprays. The growth and physiological parameters were recorded on the 10th day after the first and second sprays.

Measurement of morpho-physiological and biochemical traits

The plant height was recorded from the stem's base to the longest leaf's tip. Measurement of leaf area was done using a leaf area meter (LICOR, Model LI 3000, Lincoln, NE, USA). Leaf samples were collected from each replication for every treatment and placed into the leaf area meter, and the value was expressed as cm2 plant-1. The reading was taken during different stages of the crop, namely tillering and panicle initiation.

The chlorophyll index (SPAD value) was measured using a chlorophyll meter from Minolta (Model 502 Minolta Japan). Measurements were taken from the five physiologically active leaves for all replications, and then the average was calculated. The gas exchange parameters were recorded using a portable photosynthesis system (PPS; LI-6400 XT, Licor Inc., Lincoln, NE, USA). Three values were recorded for each replication, and an average was calculated. The photosynthetic rate, transpiration rate, and stomatal conductance were expressed as μ mol of CO2 m-2 s-1, mmol H2O m-2 s-1.

Total chlorophyll content was estimated by a standardized protocol (20). About 25 g of fresh sample with 10 ml of 80% acetone was separated using centrifugation for 3000rpm for 10 minutes. The supernatant was taken, and the volume was 25 ml using 80% acetone. The OD values were taken at 652 nm in a spectrophotometer (Shimadzu; UV-1800) and expressed in mg/g.

Soluble protein was estimated using the a standardized protocol (21). The procedure includes maceration of 0.25 g of leaf sample using 10 ml of phosphate buffer. Centrifugation is done at 3000 rpm for 10 minutes and made the volume to 25 ml. 1 ml of the supernatant was taken, 5 ml of Alkaline copper tartrate (ACT), and 0.5 ml of folin–Ciocalteau reagent was added, and the color developed was measured after 30 minutes at 660 nm in a spectrophotometer (Shimadzu; UV-1800) and expressed in mg/g. The standard was prepared using 50 mg of Bovine Serum Albumin (BSA), dissolved in 100 ml of distilled water with a stock of 500 ppm. Different concentrations of BSA stock solutions were prepared by diluting the stock solution (100, 200, 300, 400, and 500 ppm).

Quantification of nutrient content and its use efficiency

Total N was measured by the Kjeldahl method using a 4:1 sulfuric acid to perchloric acid mixture (22). Total P was determined via a triple acid extraction with nitric, sulfuric, and perchloric acids in a 9:2:1 ratio using a spectrophotometer (23).

NUE was calculated using the formula (Eqn. 1) (24)

Nitrogen use efficiency (kg kg⁻¹) = Dry yield Quantity of Nutrient applied + Nutrient in soil before transplanting

...(Eqn.1)

Measurement of yield components

The yield and yield components were observed during crop harvest. The number of productive tillers per plant, grain yield, straw yield, panicle length, panicle weight, number of total grains per panicle, number of filled grains per panicle, number of unfilled grains per panicle, 1000 seed weight all were recorded from randomly selected five plants average in each replication.

Statistical Analysis

The experiment was conducted using a randomized block design, and the data collected was analyzed using SPSS software. A one-way analysis of variance (ANOVA) was performed on nine treatments and three replications, and significance was determined at the 5% level.

Results and Discussion

Impact of nano-DAP on morpho-physiological parameters

Nano-diammonium phosphate is a nanomaterial-based fertilizer that provides plants with a controlled release of nutrients, specifically N and P. It contains 8% N and 16% P. Its advantages over conventional fertilizers include improved nutrient use efficiency, reduced nutrient losses, and enhanced plant uptake due to its smaller particle size. Foliar application of Nano-DAP significantly alters morphological traits, including plant height in rice variety ADT 53. The tallest plants (81.2 and 99.6 cm) were recorded in 75% RDNP with two Nano-DAP sprays, while the shortest (66.8 and 82.8 cm) were observed in the control and application of 100% RDF, resulting in heights of 80.1 and 97.6 cm. Nano-DAP spray significantly ($P \le 0.05$) increased plant height by 20.2% over the control and 2% over 100% RDF (Table 2). Therefore, Nano -DAP spray showed a marginal increase of 2% in plant height compared to RDF, suggesting that nanoform of N has a minimal impact on plant height, which may not hinder the yield enhancement crops such as rice. These results were similar to the findings of another study, which stated that DAP spray increased plant height by 2% (25). In cereal such as wheat, a reduction in plant height through peduncle length changes increases grain yield and grain number. Therefore, higher yields are not necessarily associated with increased plant height (26). Although few studies showed the increased plant height with nanofertilizer, our study result revealed that it was on par with an RDF.

Foliar application of nano-DAP significantly enhances leaf area and dry matter production

The treatment with 75% RDNP combined with two sprays of Nano-DAP resulted in the highest leaf area, with 1553.2 cm² plant-1 and 2053.9 cm² plant-1 after the first and second sprays, respectively. This represents an 11 to 12% increase compared to 100% RDF. The response of plants from first to second spray for leaf area in case of 75% RDNP + 2 sprays of Nano-DAP is 32.2% suggesting that two sprays of Nano-DAP are essential for significant ($P \le 0.05$) incremental increase in leaf area than single spray (Fig. 1). Reduced fertilizer doses (50% and 75% RDNP) without foliar sprays resulted in the lowest leaf area, indicating the importance of supplementing soil fertilization with foliar application. These results suggest that nano-DAP positively influences leaf area by supplying N and P in nano form, which enhances leaf production when applied at optimal concentrations. This reduces nutrient competition among plants, resulting in a larger leaf area. Besides, the efficiency of DAP application is predominantly due to N's role in increasing leaf area, as increasing the leaf area is important for light capture, which increases carbon assimilation in the leaves, leading to greater productivity. In addition, the nutrients stored in leaf tissues are also transported during seed formation, contributing to fuller seeds and greater weight gain (27).

In this study, the application of nanoform of N triggers chlorophyll production and, therefore, the rate of photosynthesis. N in nanoform is more effective than conventional forms because its smaller particle size allows for a controlled and gradual release, ensuring N remains available to plants over an extended period by minimizing N losses. The increased surface area of nano-N enhances its absorption by plant roots and leaves, improving NUE. This leads to better crop yields with fewer applications and a reduced environmental impact. This enhancement improves the translocation of assimilates and photosynthates to various plant parts, resulting in greater dry matter (DM) accumulation (28). Our study result revealed that 75% RDNP and two foliar sprays of Nano-DAP showed significant (P \leq 0.05) higher dry matter production of 45.8 g/plant after the first spray and 98 g/plant after the second spray, which was 15.3% higher than the 100% RDF treatment (Table 1). It is understood from the data that foliar application of nutrients is highly effective when applied in nanoform, as evidenced by the high accumulation of dry matter due to various growth and physiological traits. Besides, the foliar application of major nutrients improves the accumulation and translocation of photosynthates, leading to greater biomass production (29). In addition, N and P, which are vital for photosynthesis and carbohydrate production, also significantly increase dry matter accumulation due to nano-DAP when applied through foliar spray (30).

The role of N in chlorophyll synthesis is implicated in many studies as it is a key component of the chlorophyll molecule. Nitrogen is essential for chlorophyll synthesis as it is a key component of the chlorophyll molecule. It aids in forming amino acids crucial for producing chlorophyll and related proteins. Adequate N levels enhance chlorophyll production, improving a plant's light absorption for photosynthesis and ultimately boosting growth and productivity. Adequate N increases chlorophyll content and enhances photosynthetic efficiency, while deficiency leads to chlorosis and reduced plant growth. Recent research highlights the importance of optimized N management to maintain optimal chlorophyll levels and improve crop performance (31). Optimising N dose is critical in crops such as rice where excessive application can lead to nutrient imbalances, increased susceptibility to pests and diseases, and environmental issues such as water pollution and greenhouse gas emissions. In addition, high N levels often result in excessive vegetative growth at the expense of yield and quality (31). In the study, the chlorophyll index showed significant differences among treatments. The result indicated that the first and second sprays of Nano-DAP and DAP increased the chlorophyll index, with Nano-DAP showing a higher increase than DAP. Among the treatments, 75% RDNP + 2 sprays of Nano-DAP exhibited the highest values (41.7 and 46.2 SPAD units) after the first and second sprays, respectively, compared to 100% RDF.

From these results, the rate of chlorophyll content synthesis between the tillering and panicle initiation stage is significantly ($P \le 0.05$) higher (10.5%) in 75% RDNP + two sprays of Nano-DAP compared to 100% RDF (8.5%) (Table 3). This can be attributed to a slower release of N and P in Nano-DAP in active tillering and panicle initiation stages. Besides, the increased surface area of nanoparticles enhances absorption efficiency and promotes enzyme activities that inhibit ethylene production, reducing chlorophyll degradation (32). Additionally, nanoparticles activate photosynthesis enzymes, improving chloroplasts' ability to absorb sunlight and convert it into stored energy, increasing chlorophyll content.

Nano-DAP elevates the soluble protein content and gas exchange parameters:

Nitrogen and P significantly affect the enzymes involved in the application 75% RDNP and two applications of Nano-DAP during critical growth stages, as indicated by the highest amount of total soluble protein, which was 17.74 mg/g -3.2% higher ($P \le 0.05$) compared to 100% RDF (Table 1). Generally, soluble protein serves as a surrogate, which indicates the performance of the Rubisco enzyme involved in carbon fixation due to enhanced photosynthetic activity. The soluble protein content is a crucial regulator of cell osmotic potential, with higher levels helping to maintain a low osmotic potential and strengthening the plant's resistance to stress-induced damage (33). In this study, soluble protein content increased with N levels, suggesting that an optimal amount of N can boost protein synthesis and soluble protein content in rice. These results corroborate the findings of a previous experiment (34).

In the case of gas exchange parameters, the application of 75% RDNP + 2 Nano-DAP sprays exhibited the highest photosynthetic rates (24.8 and 28.4 μ mol CO2 m-2-s-1) (Fig. 2), transpiration rates (5.5 and 6.1 mmol H2O2 m-2s-1) (Fig. 3), and stomatal conductance (0.320 and 0.369 mol H220 m-2s-1) (Fig. 4) after each spray. Conversely, the 2% DAP with reduced RDNP levels showed lower rates across all parameters, implying the beneficial effects of Nano-DAP on key processes such as photosynthesis traits essential for

growth and productivity. The percent increase in two Nano-DAP sprays with 75% RDNP showed a significant ($P \le 0.05$) increase in photosynthetic rate, transpiration rate, and stomatal conductance to the level of 12.2%, 7%, and 12.2%, respectively, compared to RDF signifying that nano-DAP has a major impact on photosynthesis and stomatal conductance compared to transpiration rate. From active tillering to panicle initiation, the percent increase in photosynthetic rate (14.5%) and stomatal conductance (15.3%) is higher with 75% RDNP and 2 sprays of Nano-DAP compared to 100% RDF. P is

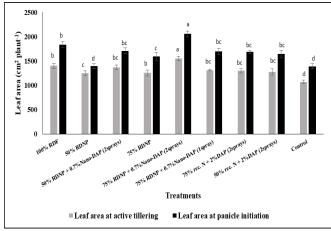
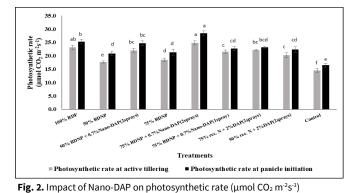
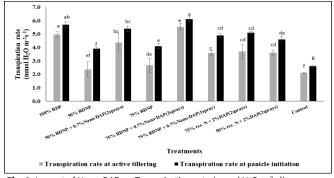


Fig.1. Impact of Nano-DAP on leaf area (cm² plant⁻¹)







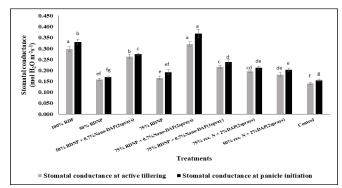


Fig. 4. Impact of Nano-DAP on Stomatal conductance (mol H₂O m⁻²s⁻¹)

Plants treated with nano-KH₂PO₄ in Yoshida solution exhibit improved PUE compared to conventional fertilizers, as nanofertilizers possess smaller particle sizes and larger surface areas that enhance metabolic processes (36). Nano N and P functions include the synthesis of chlorophyll and thylakoids, as well as the development of chloroplasts and also essential for energy transfer within the plant and support numerous enzymatic processes, including photosynthesis for promoting plant growth (37). Further, many studies suggest that applying nanoforms of N and phosphorous enhances nutrient absorption, promoting the optimal growth of plant parts and increasing metabolic processes, such as photosynthesis. This leads to increased accumulation of photosynthates and their efficient translocation to the plant's economic parts, thereby improving crop growth, development, and yield (38).

Nano-DAP increases grain yield primarily due to the number of productive tillers

Nano-DAP sprays enhanced yield primarily by enhancing the number of productive tillers, resulting in reduced soil application of fertilizers. Among the treatments, 75% RDNP + 2 Nano-DAP sprays recorded the highest number of productive tillers, compared to 100% RDF and 50% RDNP + 2 DAP sprays (Table 4). This amounts to a 13.3% percent increase ($P \le 0.05$) in 75% RDNP + 2 Nano-DAP treated crops compared to all other parameters, which can be attributed to N and P role in reproductive development as deficiencies can restrict tiller initiation and development, disrupting normal growth processes and significantly impacting grain yield in paddy fields (30).

Treatment with 75% RDNP + 2 Nano-DAP sprays recorded the highest 1000 seed weight of 14.98 g, although slight increase of 1.1% compared to 100% RDF. It is clear from the data that nano-DAP treatment has little influence; therefore, the 75% RDNP + 2 Nano-DAP treatment is on par with the recommended dose of fertilizer (Table 4). These results corroborate with a previous study (39). Among the treatments, the highest grain yield was recorded in 75% RDNP + 2 Nano-DAP sprays (6378 kg/ha), a 5.8% increase (P ≤ 0.05) over 100% RDF (6028 kg/ha), and also the highest straw yield at 8414 kg/ha, marking a 10.3% ($P \le 0.05$) improvement compared to 100% RDF (8164 kg/ha). In comparison, 50% RDNP + 2 Nano-DAP sprays produced a grain yield of 5573 kg/ ha, 8.2% decrease, and a straw yield of 7629 kg/ha, a 1.6% reduction relative to 100% RDF (Table 4). The higher grain yield in 75% RDNP + 2 Nano-DAP sprays compared to 100% RDF is due to the enhanced absorption, interception, and utilization of P from nanoform. This slow P release provides a steady supply throughout the growth period, increasing greater photosynthesis and resulting in biomass accumulation (40).

To optimize grain yield in paddy fields, providing sufficient P is crucial for promoting healthy and abundant grain production (41). The foliar application of nano fertilizer enhances nutrient absorption and translocation more rapidly, increasing photosynthesis and dry matter

Table 1. Details of Treatment

Treatments	Details of treatments
T1	Recommended dose of fertilizer (RDF) + all package of practices
T ₂	50% RDNP
T ₃	50% RDNP + 2 sprays of Nano-DAP at 0.7% during active tillering and panicle initiation
T ₄	75% RDNP
T ₅	75% RDNP + 2 sprays of Nano-DAP at 0.7% during active tillering and panicle initiation
T ₆	75% RDNP + 1 spray of Nano-DAP at 0.7% during tillering (for cost effectiveness)
T ₇	75% rec. N through fertilizer + 2 sprays of DAP at 2% during active tillering and panicle initiation
T ₈	50% rec. N through fertilizer + 2 sprays of DAP at 2% during active tillering and panicle initiation
T۹	Control (No nitrogen and phosphorus)
	The recommended dose of K were applied in the all treat- ments

accumulation, eventually leading to a higher straw yield. These findings align with the results of previous studies where it was reported that spraying nano di-ammonium phosphate on leaves improves plant metabolism and photosynthesis, which increases the number of flower clusters and grain development, leading to a substantial increase in crop yield (42, 43). Applying nano di-ammonium phosphate 30 days after planting and one week before

flowering activates metalloprotease and enzyme functions essential for enhancing grain and straw yield. This approach ensures efficient nutrient delivery throughout the crop's growth cycle (44).

The highest total grains per panicle was recorded at 75% RDNP + 2 Nano-DAP sprays (278), compared to 100% RDF (Table 4). The increase in the number of grains per panicle may be due to timely N delivery, which stimulates grain production and increases the overall count. Additionally, foliar spraying of nano urea improves photosynthate assimilation and translocation from source to sink, enhancing plant growth and development (45). Similarly, the number of filled grains per panicle was also higher in 75% RDNP + 2 Nano-DAP sprays (264) compared to 100% RDF (Table 4). The higher number of filled grains per panicle in 75% RDNP + 2 sprays of Nano-DAP is due to initial N from soil-applied urea, which supports biomass accumulation and a strong sourcesink relationship. Foliar application of nano fertilizer during active tillering and panicle initiation further enhances N availability, boosting photosynthate production and translocation. This increases the number of filled grains per panicle (46). The increased number of filled grains is due to improved enzymatic activity, which enhances the formation and transport of photosynthates. This process likely contributes to more grains per panicle (47). 75% RDNP + 2 Nano-DAP sprays had the fewest unfilled grains per panicle (14) (Table 4). A sufficient amount of N and P supply to the crop through nano fertilizer reduces the unfilled grains, which may be due to increased P content that helps in grain formation.

Among the treatments, 75% RDNP + 2 Nano-DAP sprays had

 Table 2. Effect of foliar application of Nano-DAP on the growth and biochemical traits of rice grown during Navarai, 2024

	Plant height (cm plant ⁻¹)		Dry matt	er produc-	Total ch	lorophyll	Soluble protein	
			tion (g plant ¹)		(mg g ⁻¹)		(mg g ⁻¹)	
Treatments	Active tillering	Panicle initia- tion	Active tillering	Panicle initiation	Active tillering	Panicle initiation	Active tillering	Panicle initiation
Recommended dose of fertilizer (RDF) + all package of practices	80.1 ^{ab}	97.6 ^{ab}	41 ^b	85 ^b	1.31ª	1.36 ^{ab}	16.09ª	17.12ª
50% RDNP	69.4 ^{cd}	86.2 ^{cd}	27 ^f	57 ^d	0.99 ^c	0.97 ^{de}	12.39 ^{de}	11.52 ^d
50% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation stage	75.5 ^{abc}	94 ^{abc}	40 ^b	83 ^b	1.28 ^b	1.32 ^{ab}	15.97ª	16.71 ^{ab}
75% RDNP	70.4 ^{cd}	88.8 ^{bcd}	31 ^e	67 ^c	1.06 ^c	1.02 ^c	12.83 ^{cde}	12.26 ^d
75% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation stage	81.23ª	99.6ª	46ª	98ª	1.36ª	1.43ª	16.83ª	17.74ª
75% RDNP + 1 spray of Nano-DAP at the rate of 0.7% during tillering stage	74.97 ^{abcd}	93.3 ^{abc}	38 ^{bc}	79 ^b	1.26 ^{bc}	1.30 ^{ab}	15.44 ^{ab}	16.37 ^{ab}
75% recommended dose of N through fertilizer + 2 sprays of DAP at the rate of2% during active tillering and panicle initiation stage	72.13 ^{bcd}	91.8 ^{abcd}	36 ^{cd}	80 ^b	1.19°	1.22 ^b	14.23 ^{bc}	15.11 ^{bc}
50% recommended dose of N through fertilizer + 2 sprays of DAP at the rate of 2% during active tillering and panicle initiation stage	70 ^{cd}	90.3 ^{abcd}	34 ^{de}	68 ^c	1.18 ^c	1.19 ^{bc}	13.46 ^{cd}	14.15°
Control (No nitrogen and phosphorus)	66.8 ^d	82.8 ^d	17 ^g	33 ^e	0.91 ^{bc}	0.85 ^e	11.72 ^e	10.93 ^d
CD (P ≤ 0.05)	8.47	9.64	3.34	8.40	0.09	0.14	1.65	1.68

	Chlorophyll Index (SPAD units)							
Treatments	Before spray	10 days after 1 st spray	15 days after 1 st spray	10 days after 2 nd spray	15 days after 2 nd spray			
Recommended dose of fertilizer (RDF) + all package of practices	34.6ª	38.6 ^{ab}	40 ^{ab}	43.2 ^{ab}	43.4 ^{ab}			
50% RDNP	35ª	33.2 ^c	33.0 ^c	31.9 ^d	31.3 ^c			
50% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation stage	36.8ª	38 ^{ab}	39.7 ^{ab}	41.2 ^{ab}	42.0 ^{ab}			
75% RDNP	35.8ª	35.4 ^{bc}	35.9 ^{bc}	34.6 ^{cd}	34.8 ^c			
75% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation stage	35.5ª	41.1ª	41.8ª	45.3ª	46.2ª			
75% RDNP + 1 spray of Nano-DAP at the rate of 0.7% during	36.6ª	38.9 ^{ab}	39.1 ^{abc}	41.2 ^{ab}	41.8 ^{ab}			
75% recommended dose of N through fertilizer + 2 sprays of DAP at the rate of 2% during active tillering and panicle initiation stage	36.5ª	38.1 ^{ab}	38.7 ^{ab}	41.1 ^{ab}	41.7 ^b			
50% recommended dose of N through fertilizer + 2 sprays of DAP at the rate of 2% during active tillering and panicle initiation stage	36.8ª	38 ^{ab}	37.1 ^{bc}	39 ^{bc}	39.5 ^b			
Control (No nitrogen and phosphorus)	37.9ª	35.6°	35.91°	30.8°	30.47°			
CD (P ≤ 0.05)	NS	4.31	4.35	4.43	4.48			

the highest weight of 3.3g, compared to 100% RDF and treatment with 50% recommended N with two Nano-DAP sprays at 2% during critical growth stages (Table 4). The increased panicle weight is primarily due to enhanced photosynthetic activity, dry matter production, and efficient translocation of photosynthates facilitated by the foliar application of nano fertilizers. These fertilizers improve nutrient delivery by penetrating stomata and providing a timely nutrient supply (48).

Nitrogen and phosphorus use efficiency significantly improved by Nano-DAP

The study showed nano-DAP treatments recorded higher N and P content in leaves than 100% RDF. 75% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation recorded higher N (1.19%) and phosphorous content (0.34%) content compared to 50% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation and 75% RDNP + 1 spray of Nano-DAP at the rate of 0.7% during tillering (Fig. 5). This could be because nano fertilizers have a large surface area and small particle size that enhances their ability to penetrate plants from the applied surface, improving both nutrient uptake and use efficiency. This increased contact increases penetration and nutrient uptake, resulting in higher nutrient content and improved absorption (40). A 75% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% recorded increased N and PUE of 53.3 and 54.4 kg/kg, respectively, compared to 50% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% (52.5 and 53.4 kg/kg) and 100% RDF application (Fig. 6).

The higher NUE and PUE in rice may be due to increased biomass production associated with higher uptake of N and P. Rice generally has higher NUE for biomass (NUEb) and grain (NUEg) compared to other C3 crops like soybean and wheat, due to its lower N content in straw and grain. While rice's efficiency in N remobilization is lower than maize due to its C3 nature, improving grain yield and N concentration through physiological strategies can enhance NUE in rice (49). Phosphorus use efficiency in crop plants, measured as dry matter production per unit of P uptake, is typically higher than that of N and potassium (K). Nano-DAP increases PUE by enhancing P availability and uptake with its larger surface area and controlled release features, which leads to improved crop yields and minimizes P losses compared to traditional fertilizers.

Conclusion

Nano-DAP application in rice cultivation has shown promising results in optimizing physiological and biochemical traits, resulting in enhanced crop performance, as indicated by

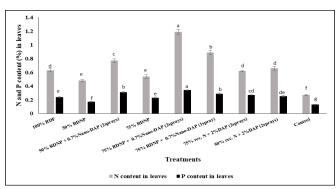


Fig. 5. Impact of Nano-DAP on N and P content (%) in leaves of rice at harvest

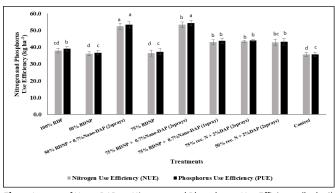


Fig. 6. Impact of Nano-DAP on Nitrogen and Phosphorus Use Efficiency (kg kg $^{\rm 1})$ in rice at harvest

	Yield attributes						Yield		
Treatments	Number of produc- tive tillers per hill	1000 seed weigh t (g)	Filled grains panicle ⁻	Unfilled grains panicle ⁻ 1	Total grains panicle ⁻¹	Pani- cle weight (g)	Grain Yield (kg ha ⁻¹)	Straw Yield (kg ha ⁻¹)	
Recommended dose of fertilizer (RDF) + all package of practices	15 ^b	14.8ª	244ª	18 ^d	262ª	3.15 ^{ab}	6028 ^{ab}	8164ª	
50% RDNP	12 ^c	12.7 ^{cd}	166 ^c	26 ^b	192 ^{cd}	2.34 ^c	4336 ^e	6294 ^{cd}	
50% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation stage	15 ^b	14.5 ^{ab}	242ª	20 ^c	260ª	3.13 ^{ab}	5573 ^{bc}	7629 ^{ab}	
75% RDNP	13 ^{bc}	13.0 ^{bcd}	176 ^c	24 ^b	200 ^c	2.36 ^c	4782 ^{de}	6821 ^{bc}	
75% RDNP + 2 sprays of Nano-DAP at the rate of 0.7% during active tillering and panicle initiation stage	17ª	15.0ª	264ª	14 ^d	278ª	3.3ª	6378ª	8414ª	
75% RDNP + 1 spray of Nano-DAP at the rate of 0.7% during tillering	14 ^b	14.2 ^{abc}	208 ^b	23 ^d	231 ^b	2.85 ^b	5528 ^{bc}	7589 ^{ab}	
75% recommended dose of N through fertilizer + 2 sprays of DAP at the rate of 2% during active tillering and panicle initia- tion stage	14 ^b	13.9 ^{abc}	182°	26 ^b	208°	2.53°	5236 ^{cd}	7206 ^b	
50% recommended dose of N through fertilizer + 2 sprays of DAP at the rate of 2% during active tillering and panicle initiation stage	13 ^{bc}	13.4 ^{abc}	184°	21 ^c	205°	2.45°	5048 ^{cd}	7180 ^b	
Control (No nitrogen and phosphorus)	9 ^d	12.4 ^d	141 ^d	30ª	171 ^d	1.96 ^d	3348 ^f	5614 ^d	
CD (P ≤ 0.05)	1.73	1.59	23.46	2.93	25.57	0.31	596.45	834.23	

higher yield compared to conventional soil application of fertilizers. This study demonstrates that nanotechnologybased foliar application, combined with a reduced recommended fertilizer dose, improved NUE and provided added benefits such as enhanced crop yield and sustainability compared to 100% soil application. .Among the nine treatment combinations tested, 75% of the recommended dose of fertilizer with two sprays of nano-DAP during the active tillering and panicle initiation stages proved the most effective in terms of morpho-physiological traits among them, leaf area (12.1%), dry matter production photosynthetic rate (12.3%), and stomatal (15.3%) conductance (12.2%) number of productive tillers (13.3%) were significantly impacted positively. The findings conclude that 25 % of soil applications of conventional fertilizers can be saved through two foliar applications of nano-DAP, in addition to achieving a higher yield in rice. Therefore, Nano-DAP fertilizers can be effectively used at the field level to improve rice productivity by altering plant physiological parameters and making it a valuable asset for sustainable agriculture. However, further research is needed to understand the process at the molecular level and make this technology more viable and sustainable. Investigating the signal transduction pathways activated by Nano-DAP can elucidate its effects on plant growth and photosynthesis. Additionally, studying the gene expression associated with nutrient transport will enhance our understanding of its impact on crop resilience and productivity.

Acknowledgments

We sincerely thank the Indian Farmers Fertiliser Cooperative Limited (IFFCO) for their generous financial support, which was crucial for this research project's progress and successful completion. We also appreciate the valuable guidance and resources provided by the IFFCO team, which greatly enhanced the quality of our work.

Authors' contributions

RRBS and PB participated in data collection and analysis. AS, DJ and GPK reviewed and composed the manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interest to declare.

Ethical issues: None

Declaration of generative AI and AI-assisted technologies in the writing process

While preparing this work, the authors used ChatGPT for language assisting service. After using this tool/service, the authors reviewed and edited the content as needed and took full responsibility for the publication's content.

References

- Sinha D, Tandon PK. An overview of nitrogen, phosphorus and potassium: key players of nutrition process in plants. In: Mishra K, Tandon PK, Srivastava S, editors. Sustainable solutions for elemental deficiency and excess in crop plants. Singapore: Springer; 2020. p. 85-117. https://doi.org/10.1007/978-981-15-8636-1_5
- Smith J, Nayak D, Yeluripati J. The potential use of biochar to reduce nitrogen waste from farming systems in India. Curr Res Environ Sustain. 2023;5:100224. https://doi.org/10.1016/ j.crsust.2023.100224
- 3. Johnson H, Simpson EM, Troldborg M, Ofterdinger U, Cassidy R,

Soulsby C, Comte JC. Evaluating groundwater nitrate status across the river than catchment (Scotland) following two decades of nitrate vulnerable zone designation. Environments. 2023;10(4):67. https://doi.org/10.3390/environments10040067

- Mahachandramuki E, Thirukumaran K, Karthikeyan R, Sivakumar R, Sellamuthu KM, Prabukumar G. Influence of super nano urea and nano-DAP on growth parameters of rice fallow cotton under high density planting system. Int J Plant Soil Sci. 2023;35(19):711-16. https://doi.org/10.9734/ijpss/2023/ v35i193602.
- Sohrt J, Lang F, Weiler M. Quantifying components of the phosphorus cycle in temperate forests. Wiley Interdiscip Rev Water. 2017;4(6):e1243. https://doi.org/10.1002/wat2.1243
- Martre P, Dueri S, Guarin JR, Ewert F, Webber H, et al. Global needs for nitrogen fertilizer to improve wheat yield under climate change. Nat Plants. 2024;10:1081-90. https:// doi.org/10.1038/s41477-024-01739-3
- Kokulan V, Schneider K, Macrae ML, Wilson H. Struvite application to field corn decreases the risk of environmental phosphorus loss while maintaining crop yield. Agr Ecosyst Environ. 2024;366:108936. https://doi.org/10.1016/ j.agee.2024.108936.
- Egan G, McKenzie P, Crawley M, Fornara DA. Effects of grassland management on plant nitrogen use efficiency (NUE): Evidence from a long-term experiment. Basic Appl Ecol. 2019;41:33-43. https://doi.org/10.1016/j.baae.2019.10.001.
- Khan N, Siddiqui MH, Ahmad S, Ahmad MM, Siddiqui S. New insights in enhancing the phosphorus use efficiency using phosphate-solubilizing microorganisms and their role in cropping system. Geomicrobiol J. 2024;41(5):485-95. https:// doi.org/10.1080/01490451.2024.2331111.
- Uddin S, Nitu TT, Milu UM, Nasreen SS, Hossenuzzaman M, Haque ME, Hossain B, et al. Ammonia fluxes and emission factors under an intensively managed wetland rice ecosystem. Environ Sci Process Impacts. 2021;23(1):132-43. https:// doi.org/10.1039/D0EM00374C
- Panda D, Nayak AK, Mohanty S. Nitrogen management in rice. Oryza. 2019;56(5):125-35. https://doi.org/10.35709/ ory.2019.56.s.5
- Acharya SM, Bhakare BD, Durgude AG, Thakare R. Foliar application of nano fertilizer in agricultural crops: A review. Bhartiya Krishi Anusandhan Patrika. 2023;38(4):339-48.https:// doi.org/10.18805/BKAP643
- Ding Y, Zhao W, Zhu G, Wang Q, Zhang P, Rui Y. Recent trends in foliar nanofertilizers: A review. Nanomaterials. 2023;13 (21):2906. https://doi.org/10.3390/nano13212906
- 14. Yadav A, Upadhyay A, Kumar R, Prajapati J, Pal S. Nanotechnology based nano urea to increase agricultural sustainability. Just Agric. 2023;3(12):266-74.
- Gul H, Said A, Saeed B, Mohammad F, Ahmad I. Effect of foliar application of nitrogen, potassium and zinc on wheat growth. ARPN J Agric Biol Sci. 2011;6(4):56-59.
- Otálora G, Piñero MC, López-Marín J, Varó P, del Amor FM. Effects of foliar nitrogen fertilization on the phenolic, mineral and amino acid composition of escarole (*Cichorium endivia* L. var. *latifolium*). Sci Hortic. 2018;239:87-92. https:// doi.org/10.1016/j.scienta.2018.05.031.
- Rameshaiah GN, Pallavi J, Shabnam S. Nano fertilizers and nano sensors-an attempt for developing smart agriculture. Int J Eng Res Gen Sci. 2015;3(1):314-20.
- 18. Mahil El, Kumar BA. Foliar application of nanofertilizers in agricultural crops–A review. J Farm Sci. 2019;32(3):239-49.
- Kumar P, Naib SD, Pandey R, Sharma G. Efficacy of emerging plant nutrition sources as alternative/supplement to the conventional nutrient sources. Indian J Fertil. 2022;18(10):974-

88.

- Yoshida S, McLean GW, Shafi M, Mueller KE. Effects of different methods of zinc application on growth and yields of rice in a calcareous soil, West Pakistan. Soil Sci Plant Nutr. 1970;16 (4):147-49. https://doi.org/10.1080/00380768.1970.10432832
- Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. J Biol Chem. 1951;193(1):265-75. https://doi.org/10.1016/S0021-9258(19) 52451-6
- Humphries EC. The absorption of ions by excised root systems: III. Observations on roots of pea plants grown in solutions deficient in phosphorus, nitrogen or potassium. J Exp Bot. 1952;3(3):291-309. https://doi.org/10.1093/jxb/3.3.291
- Sommers LE, Nelson DW. Determination of total phosphorus in soils: a rapid perchloric acid digestion procedure. Soil Sci Soc Am J. 1972;36(6):902-04. https://doi.org/10.2136/ sssaj1972.03615995003600060020x
- Karam F, Mounzer O, Sarkis F, Lahoud R. Yield and nitrogen recovery of lettuce under different irrigation regimes. J Appl Hort. 2002;4(2):70-76. https://doi.org/10.37855/ jah.2002.v04i02.21
- Al-Khuzai AH, Al-Juthery HW. Effect of DAP fertilizer source and nano fertilizers (silicon and complete) spray on some growth and yield indicators of rice (*Oryza sativa* L. cv. Anber 33). In: Proceedings of the 1st Scientific International Virtual Agricultural Conference; 2020 May 31–Jun 1. Al-Qadisiyah, Iraq. IOP Conference Series: Earth and Environmental Science; 2020. 553:012008. https://doi.org/10.1088/1755-1315/553/1/012008
- Du Y, Chen L, Wang Y, Yang Z, Saeed I, Daoura BG, Hu YG. The combination of dwarfing genes Rht4 and Rht8 reduced plant height, improved yield traits of rainfed bread wheat (*Triticum aestivum* L.). Field Crops Res. 2018;215:149-55. https:// doi.org/10.1016/j.fcr.2017.10.015.
- Sabah NU. Phosphorus utilization from rock phosphate by maize (*Zea mays* L.) through integrated organic manures [Doctoral thesis]. Sargodha (Pakistan): University of Sargodha; 2018.
- Singh P, Khangarot AK. Nano-fertilizers a technology to increase crop production. Agriculture and Food: e-newsletter. 2021;3 (11):379-81.
- Mishra B, Sahu GS, Mohanty LK, Swain BC, Hati S. Effect of nano fertilizers on growth, yield and economics of tomato variety Arka Rakshak. Indian J Pure Appl Biosci. 2020;8:200-04. https:// doi.org/10.18782/2582-2845.8420
- Maloth A, Thatikunta R, Parida BK, Naik DS, Varma N. Evaluation of nano-DAP on plant growth, enzymatic activity and yield in paddy (*Oryza sativa* L.). Int J Environ Climate Change. 2024;14 (1):890-97. https://doi.org/10.9734/ijecc/2024/v14i13907
- Mao L, Song Q, Li X, Zheng H, Zhu XG. Would reducing chlorophyll content result in a higher photosynthesis nitrogen use efficiency in crops?. Food Energy Secur. 2024;13(4):e576. https://doi.org/10.1002/fes3.576
- Liu C, Zhou H, Zhou J. The applications of nanotechnology in crop production. Molecules. 2021;26(23):7070. https:// doi.org/10.3390/molecules26237070
- Nathawat NS, Kuhad MS, Goswami CL, Patel AL, Kumar R. Interactive effects of nitrogen source and salinity on growth indices and ion content of Indian mustard. J Plant Nutr. 2007;30 (4):569-98. https://doi.org/10.1080/01904160701209329
- Liao S, Qin X, Luo L, Han Y, Wang X, Usman B, et al. CRISPR/Cas9
 -induced mutagenesis of semi-rolled leaf 1, 2 confers curled leaf
 phenotype and drought tolerance by influencing protein
 expression patterns and ROS scavenging in rice (*Oryza sativa* L.).
 Agronomy. 2019;9(11):728. https://doi.org/10.3390/
 agronomy9110728

- Gonzalez-Vega JC, Walk CL, Murphy MR, Stein HH. Requirement for digestible calcium by 25 to 50 kg pigs at different dietary concentrations of phosphorus as indicated by growth performance, bone ash concentration and calcium and phosphorus balances. J Anim Sci. 2016;94(12):5272-85. https:// doi.org/10.2527/jas.2016-0751
- Mahanta N, Kurmi K, Das JC, Basumatary A. Nutrient content, uptake and oil content of rapeseed as influenced by reduced tillage, mulching and INM practices. J Pharmacogn Phytochem. 2019;8(3):3291-95.
- Ali A, Khaliq T, Ahmad A, Ahmad S, Malik AU, Rasul F. How wheat responses to nitrogen in the field. A review. Crop Environ. 2012;3 (1-2):71-76.
- Benzon HR, Rubenecia MR, Ultra Jr VU, Lee SC. Nano-fertilizer affects the growth, development and chemical properties of rice. Int J Agron Agric Res. 2015;7(1):105-17.
- Manikandan A, Subramanian K. Evaluation of zeolite-based nitrogen nano-fertilizers on maize growth, yield and quality on inceptisols and alfisols. Int J Plant Soil Sci. 2016;9(4):1-9. https://doi.org/10.9734/IJPSS/2016/22103
- Adhikari T, Kundu S, Meena V, Rao AS. Utilization of nano rock phosphate by maize (*Zea mays* L.) crop in a vertisol of Central India. J Agric Sci Technol A. 2014;4(5):384-94.
- Deo HR, Chandrakar T, Srivastava LK, Nag NK, Singh DP, Thakur A. Effect of nano-DAP on yield, nutrient uptake and nutrient use efficiency by rice under Bastar plateau. *Pharma Innov J.* 2022;11 (9):1463-65.
- 42. Kumar Y, Tiwari KN, Nayak RK, Rai A, Singh SP, Singh AN, et al. Nanofertilizers for increasing nutrient use efficiency, yield and

economic returns in important winter season crops of Uttar Pradesh. Indian J Fertil. 2020;16(8):772-86.

- Abdel-Aziz HM, Hasaneen MN, Omer AM. Foliar application of nano chitosan NPK fertilizer improves the yield of wheat plants grown on two different soils. Egypt J Exp Biol. 2018;14(1):63-72. https://doi.org/10.5455/egyjebb.20180106032701
- Poudel A, Singh SK, Jiménez-Ballesta R, Jatav SS, Patra A, Pandey A. Effect of nano-phosphorus formulation on growth, yield and nutritional quality of wheat under semi-arid climate. Agronomy. 2023;13(3):768. https://doi.org/10.3390/ agronomy13030768
- 45. Raheem AHJ, Kadhem NH, Ghanim BN. Impact of levels and time of foliar application of nano fertilizer (Super Micro Plus) on some components of growth and yield of rice (*Oryza sativa* L.). Plant Arch. 2019;19(1):1279-83.
- Gewaily EE, Mohammed AT, Abd El-Rahem WT. Effect of different irrigation regimes on productivity and cooking quality of some rice varieties. World J Agric Sci. 2019;15(5):341-54.
- Midde SK, Perumal MS, Murugan G, Sudhagar R, Mattepally VS, Bada MR. Evaluation of nano urea on growth and yield attributes of rice (*Oryza sativa* L.). Chem Sci Rev lett. 2022;11 (42):211-14.
- Dhawne MN, Shamkuwar GR, Shamkuwar SG, Landge NH, Sable AD, Tinpayle AD. Influence of foliar applications of nano urea on morpho-physiological and yield parameters in rice (*Oryza sativa* L.). Biol Forum. 2023;15(11):548-53.
- Vijayalakshmi P, Kiran TV, Rao YV, Srikanth B, Rao IS, Sailaja B, et al. Physiological approaches for increasing nitrogen use efficiency in rice. Indian J Plant Physiol. 2013;18:208-22. https:// doi.org/10.1007/s40502-013-0042-y