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Mini Review

Diverse application and future prospects for commercial cultivation of microalgae species: A review

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Abstract

Industrial revolutions, advancements in health care, pharmaceuticals, transportation can be attributed to advancements made in the field of science and technology. Environment and natural resources has paid a heavy cost for most of industrial development. Rapid depletion of non-renewable sources of energy eventually leading towards the energy crisis, direct or indirect release of industrial effluents into soil and natural water bodies, global warming are among major consequences of industrialization. Ever since these environmental concerns have been recognized substantial studies have been conducted to minimize, control pollution and restore environment and natural resources. Among several measures cultivation of algae on large scale stands out to be a multipurpose solution. Inherent potential of microalgae species to accumulate lipids makes algae an efficient source of biofuel. Beside this ability of algae to detoxify polluted water and industrial effluent support utilization of algae for environment management and restoration. Efficient CO₂ fixation, ability to tolerate wide range of environmental conditions, minimal nutritional requirements further support commercial cultivation of algal species to achieve their widespread application. However, efforts are required to develop large scale cultivation protocols (beyond the range of photobioreactors) so as to achieve practical applicability of algae and their products. Alongwith, cultivation protocols there is simultaneous need of either selection of naturally occurring high yielding strains / species or genetic improvement. Standardization of optimum cultivation conditions along with harvesting procedure is equally important.

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Introduction

Microalgae are unicellular photosynthetic eukaryotes occupying diverse range of habitats including different environmental and geographical regions. Microalgae possess diverse applications including biofuel production,

bioremediation, utilization as biofertilizers and animal feed, bioplastic synthesis, waste water treatment, synthesis of compounds with industrial/pharmaceutical applications including pigments, anti-aging compounds (1, 2). Algal proteins and polysaccharides are of industrial and nutritional importance. The ability to convert

solar energy to lipid renders microalgae as the most appropriate and potential alternate source of energy. Besides this, their inherent ability to produce oxygen and bioremediation potential is crucial to maintain ecological balance for sustained environment (3, 4). The above mentioned applications become more pronounced owing to advantages associated with microalgae cultivation. Ability to tolerate and survive in wide range of environmental conditions, minimal nutritional requirement and rapid growth rate, efficient CO₂ fixation makes cultivation of algae convenient and efficient. However, there are also certain factors which check or restrict algae growth. Presence of toxins, contamination and competitive microbes are primary concern during cultivation of algae. Studies are required to optimize large scale commercial cultivation practices alongwith optimization of cultivation conditions, screening / production of potential / improved strains, convenient and effective harvesting methods, etc.

Microalgae species have inherent potential to convert atmospheric carbon dioxide (CO₂) into lipids. (5, 6). Biofuel production from algae species have been recognized as an effective alternate of energy source (7). However, biofuel production from algae species encounters economical challenges in terms of biofuel production on commercial scale. Several methods including hydrothermal liquefaction, pyrolysis, gasification and direct combustion have been employed for biofuel production from different algal species (8,9). Hydrothermal liquefaction (HTL) plants have been established by industries for the large-scale production of bio-crude oils (black viscous oils). Biocrude / bio-oil production focusing on the conversion of just a single microalgae through HTL technique is an effective procedure for the production of bio-oil or bio-crude production from algae feedstock (10). Dalmas Neto *et al* (11) reported fast pyrolysis of algae to produce bio oil with high yield and low cost. Du *et al* (12) reported microwave assisted pyrolysis of *Chlorella* species to produce bio-oil. Du *et al* (12) and Kapoor *et al* 2018 (13) in their microwave assisted pyrolysis (MAP) of *Chlorella* spp. using a microwave oven resulted in the production of bio-oil yield of 28.6%. GC-MS results of the bio-oil product showed the presence of aliphatic, aromatic hydrocarbons, phenols and other compounds which are the desirable compounds present in crude oils, petroleum and diesel. Production of biofuel from algae species generally employ utilization of thermo chemical processes, due to which a large proportion of proteins undergo decomposition. Kumar *et al* (14) depicted a process in which proteins present in algae were separated out (through hydrolysis) before algal biomass was subjected to biofuel production. The study also reported synthesis of polyurethane from algal proteins.

Studies conducted have revealed variation in optimum light is important for lipid synthesis in algal species (5, 15-17). Ever since the potential of algae species to produce biofuel has been recognized, there has been efforts to optimize methods to enhance lipid synthesis. Reduction in antenna size of algae species has been reported to enhance photosynthesis efficiency. Several methods including UV mutagenesis (18), Insertional mutagenesis (19), RNAi mediated gene silencing (20), Random mutagenesis (21), EMS induced mutagenesis (22, 23) have been reported to enhance / improve photosynthetic efficiency of algae species.

With ever increasing industrialization, industrial effluents are directly or indirectly released into the natural resources results in environmental pollution. Effluents released from various industries are enriched with potential toxic components such as heavy metals, enhanced phosphate, sulphate and nitrate content, ammonia, dissolved solids, phenolic compounds. Due to the presence of these potentially toxic components release of industrial effluent into water bodies have gained concern.

Minimizing or treatment of polluted soil or water has become utmost important for maintaining sustainability of environment. Bioremediation techniques are not only effective and efficient but environment-friendly approach for treatment of polluted soil. Besides plants (phytoremediation); bacteria and algae reportedly possess inherent potential for bioremediation (24).

Several studies have indicated application of algal species in the treatment of waste water from various sources (Table 1). Among different algal species, *Chlorella vulgaris* is the most commonly utilized algae for waste water treatment (25-30). Kumar *et al* (31) reported *Chlorella minutissima* and *Scenedesmus* spp. to effectively reduce COD, TDS and phosphate content of sewage water. Similarly, Lim *et al* (25), Kassas and Mohammed (32) and Ramirez *et al* (33) treated textile industry effluent with different algae. All three studies reported that *Chlorella vulgaris* effectively reduced COD contents of effluent. Lim *et al* (25) also reported 33% reduction in the phosphate content with algal treatment. Kassas and Mohammed (31) simultaneously reported *Scenedesmus* spp., *C. vulgaris* and *Muriellopsis* spp. found equally effective for the treating textile industry effluent. Salgueiro *et al* (26) reported significant reduction in COD and removal of phosphate in domestic waste water treated with *C. vulgaris*. Acid mine drainage treated with *Chlorella* and *Scenedesmus* spp. resulted in the reduction of COD (34). Kumar *et al* (14) found that *C. marina* to possess potential to reduce phosphate, nitrate and ammonia from industrial effluent. Hammud *et al* (35) also reported *Spirulina platensis* to absorb copper ion from industrial waste water. The study also

Table 1. Summary of potential of different algae species utilized for treatment of effluents

S. No.	Water sample from various sources	Algae species employed for treatment	Parameters analyzed	Response / outcome	Author
1.	Sewage water	<i>Chlorella minutissima</i> , <i>Scenedesmus</i> spp.	COD, TDS, Phosphate content	Reduction in COD and TDS, removal (87.3%) of Phosphate	(31)
2	Textile industrial effluent	<i>Chlorella vulgaris</i>	COD, Phosphate content	Reduction in COD, removal (33%) of Phosphate	(25)
3.	Textile industrial effluent	<i>Chlorella</i> sp., <i>Scenedesmus</i> sp., <i>Muriellopsis</i> sp.	COD	Reduction in COD	(33)
4.	Textile industrial effluent	<i>Chlorella vulgaris</i>	COD	Reduction up to 17.5% COD	(32)
5.	Domestic, industrial, agro waste water	<i>Chlorella vulgaris</i>	COD, Phosphate content	Reduced COD Removal of Phosphate (97%)	(26)
6.	Acid mine drainage	<i>Chlorella</i> , <i>Scenedesmus</i> spp.	COD	Reduction in COD	(34)
7.	Electroplating, tanning industrial waste	<i>Chlorella vulgaris</i>	Chromium	Adsorbed Chromium (23.6 mg/g) by algae	(27)
8.	Olive mill waste water	<i>Chlorella vulgaris</i> , <i>Spirulina platensis</i>	COD, Phenolic compounds	Reduction in COD and Phenolic compounds	(36)
9.	Industrial effluent with heavy metal	<i>Chlorella vulgaris</i> , <i>Spirulina maxima</i>	Zinc and Copper	Reduced conc. of Cu upto 81% and Zn up to 94.1%	(37)
10.	Industrial effluent	<i>Chlorella marina</i>	Phosphorus, Nitrate, Ammonia	Reduced conc. of PO ₄ (88%), NO ₃ (64%), NH ₃ (85%)	(14)
11.	Industrial waste water	<i>Spirulina platensis</i>	Copper	Adsorption of Cu ions increased with decrease in algae biomass concentration	(35)
12.	Industrial effluent	<i>Chlorella minutissima</i>	Phosphorus and Nitrate	Reduced conc. of P (4.47 ppm to 1.15ppm) and N (3.6ppm to 0.3ppm)	(39)
13.	Industrial effluent	<i>Chlorella vulgaris</i> , <i>Scenedesmus quadricauda</i>	Phosphorus, Nitrate, COD, BOD	Removal efficiency of P, N, BOD, COD increased	(29)
14.	Industrial effluent	<i>Scenedesmus obliquus</i> , <i>Chlorella vulgaris</i> , <i>Chlorella kessleri</i>	COD, Phosphate content	Reduction in COD, removal of phosphate 22%-62%	(40)
15.	Industrial effluent, sewage water	<i>Chlorella vulgaris</i> , <i>Scenedesmus dimorphus</i>	COD, TDS	Reduction in COD & TDS	(41)
16.	Industrial effluent	<i>Chlorella vulgaris</i>	COD, TDS	Reduction in COD & TDS	(28)
17.	Sewage and sea well samples	<i>Chlorella vulgaris</i> , <i>Chlorella salina</i>	COD, TDS, Phosphate content	Reduction in COD & TDS, removal of phosphate	(30)

reported increased absorption of copper ions with decrease in concentration of algal biomass.

C. vulgaris and *C. salina* have also been reported to reduce TDS, COD and phosphate content from sewage and sea water samples (30). Several other studies have also reported potential of *C. vulgaris*, *S. dimorphus*, *S. quadricauda*, *S. obliquus*, *C. kessleri* to reduce COD, TDS and phosphate content of industrial effluent (28-30, 40, 41). Waste water obtained from olive mills potentially possesses high COD along with phenolic compounds. Ismail *et al* (36) utilized *Chlorella vulgaris* and *Spirulina platensis* to treat waste water obtained from olive mill and achieved reduction in COD and phenolic compounds.

One of the major concern associated with release of industrial effluent into environment (soil, water) is presence of heavy metals. Jyoti and Awasthi (27) reported *Chlorella vulgaris* to absorb chromium from waste water obtained from

electroplating and tanning industry. Chan *et al* (37) also reported *C. vulgaris* and *Spirulina maxima* to reduce the concentration of zinc and copper in industrial effluent containing heavy metals. Algae have the potential for biosorption/neutralization of heavy metals present in the industrial effluents (38). Microalgae are useful in biomonitoring and restoring the aquatic ecosystems, which make it easier to extract and degrade the various organic and inorganic pollutants (24).

Algal species exhibit antimicrobial, antioxidant activity, anti-inflammatory, hypolipidemic, antihelminthic, anticancer, antiprotozoal (42) and pharmacological properties (Table 2). Hoseini *et al* (43) reported antibacterial and antiviral potential of *Spirulina platensis*. Nicoletti (44) reported antibacterial activity of *S. platensis* and *Chlorella*. Antioxidant activity by *C. closterium* is also on record (45). Priyadarshani and Ratu (46) demonstrated antimicrobial and

Table 2. Biological activities of algae species

Sl No.	Microalgae species	Medicinal uses	Author
1	<i>Spirulina platensis</i>	Antibacterial, antiviral	(43)
2	<i>Spirulina platensis</i> and <i>Chlorella</i>	Antibacterial, dietary supplement	(44)
3	<i>Chlorella closterium</i>	Antioxidant	(45)
4	<i>Isochrysis</i> , <i>Chaetoceros</i> , <i>Chlorella</i> , <i>Arthrospira</i> , <i>Dunaliella</i>	Antimicrobial, Antifungal	(46)
5	<i>Phaeocystis pouchetii</i>	Antibiotic production	(47)
6	<i>Scenedesmus almeriensis</i> , <i>Chlorella protothecoides</i>	Reduce risk of retina damage by blue-light	(50)
7	<i>Arthrospira</i>	Antioxidant	(51)
8	<i>Phaeocystis pouchetii</i>	Antibiotic production	(48)
9	<i>Spirulina platensis</i>	Antioxidant, dietary supplement	(52)
10	<i>Chlorella sorokiniana</i>	Lowers the risk of premature death	(49)

antifungal activity of *Isochrysis*, *Chaetoceros*, *Chlorella*, *Arthrospira* and *Drenaniella*. Mostafa (47) and Sigmani *et al* (48) stated that *Phaeocystis pouchetii* produces antibiotic with potential antimicrobial action. Sathashivam *et al* (49) observed that, *Chlorella sorokiniana* lowers the risk of premature death. *Scenedesmus almeriensis* and *Chlorella protothecoides* are capable to reduce risk of retina damage caused by blue light (50). *Arthrospira* (51) and *S. platensis* (55) possess antioxidant potential.

Synthesis of bioplastics from algal polymers successfully serves as a field of growing interest for researchers. Proteins obtained from different sources have been utilized for synthesis of plastics. Wheat gluten, soy protein, peanut protein are most commonly utilized plant proteins whereas gelatin, keratin, collagen are animal proteins utilized for synthesis of plastics (53-55). However, utilization of plant material for production of plastic appears to be practically unfavourable since the same plant material can be better utilized in food industry, as animal fodder, etc. Hence, utilization of algae for synthesis of bioplastic appears to be an efficient and appropriate alternative. Utilization of algae proteins for synthesis of bioplastics has several advantages including high protein content with high algal biomass yield; comparatively cost effective cultivation in natural habitat makes algae species most suitable alternate for production of plastics (55). Zeller *et al* (56) revealed bioplastics synthesized from *Spirulina* and *Chlorella* species to be compatible with plastics synthesized from protein of plant or animal origin. Substantial technological development is required to make production of bioplastics from algae a cost effective process. There can be no debate that plastics of algae are environment friendly and biodegradable as compared to conventional plastics. Besides this increasing viability of algal bioplastics can prove to be an effective

contribution to economic development. Commercialization of bioplastics will automatically aid in conservation of fossil resources.

Conclusion

Widespread distribution and application of algal species including production of biofuel, bioremediation potential, synthesis of phytocompounds with medicinal, pharmaceutical and other biological activities renders algae to be cultivated on commercial scale. Diverse application of algae becomes more pronounced owing to their minimal growth requirement. Studies are required to optimize cultivation protocol for mass propagation of algae on large scale. Along with development of efficient cultivation protocol efforts are required to genetically modify / improve existing strains / species followed by selection of high yielding varieties. Practical application is required while utilizing algae for various purposes. Separation of proteins from algae species before extraction of oil provides an alternate to synthesize bio polymers since algal proteins are degraded during thermal/chemical processes employed during extraction of biofuel. In the present scenario, algal species stands out to be most potential bioresource for the treatment of polluted water simultaneously being an extremely effective alternate source of energy along with medicinal importance of many algal species.

Authors' contribution

NS and AS contributed towards design and prepared the initial draft of the manuscript. FL, V and RS have worked on acquisition, collection, compilation of literature for the present review. NS and RS have revised the manuscript after review. All authors read and approved the final manuscript.

References

1. Sporaore P, Joannis CC, Duran E, Isambert A. Commercial applications of microalgae. *Journal of Bioscience and Bioengineering* 2006; 101:87-96. <https://doi.org/10.1263/jbb.101.87>
2. Del CA, García GM, Guerrero M G. Outdoor cultivation of microalgae for carotenoid production: current state and perspectives. *Applied Microbiol Biotechnol.* 2007; 74:1163-74. <https://doi.org/10.1007/s00253-007-0844-9>
3. Gordon and Polle. Ultrahigh bioproductivity from algae. *Applied Microbiol. Biotechnol.* 2007; 76: 969-75. <https://doi.org/10.1007/s00253-007-1102-x>
4. Carvalho AA., Silva SO, Jose MB, Malcat XF. Light requirements in microalgal photobioreactors: an overview of biophotonic aspects. *Appl. Microbio. Biotechnol.* 2011; (89):1275-88. <https://doi.org/10.1007/s00253-010-3047-8>
5. Ibrahim M, Salman M, Kamal S, Aneza R, Sajid R, Akash H. Production and Processing of Algal Biomass. *Algae Based Polymers, Blends, and Composites. Chemistry, Biotechnology and Materials Science.* 2017; 155-271. <https://doi.org/10.1016/B978-0-12-812360-7.00006-9>
6. Ghosh A. An approach for phycoremediation of different wastewaters and biodiesel production using microalgae. *Environ. Sci. Pollut. Int.* 2018; 25(19):18673-81. <https://doi.org/10.1007/s11356-018-1967-5>
7. Ghosh SK, Tale MP, Kapadnis BP, Isolation and characterization of microalgae for biodiesel production from Nisargruna biogas plant effluent. *Bioresour Technol.* 2014; 169:328-58. <https://doi.org/10.1016/j.biortech.2014.06.017>
8. Kumar G, Shobana S, Chen WH, Bach QV, Kim SH, Atabani AE, Chang JS. A review of thermochemical conversion of microalgal biomass for biofuels: Chemistry and process. *Green Chemistry.* 2014; 19(1):44-87. <https://doi.org/10.1039/C6GC01937D>
9. Sharma PK, Saharia M, Srivastava R, Kumar S, Sahoo L. Tailoring microalgae for efficient biofuel production. *Front. Mar Sci.* 2018; 5(382):1-18. <https://doi.org/10.3389/fmars.2018.00382>
10. Barreiro DL, Wolter P, Ronse F, Brilman W. Hydrothermal liquefaction (HTL) of microalgae for biofuel production: state of the art review and future prospects. *Biomass Bioener.* 2013; 53(16):113-27. <https://doi.org/10.1016/j.biombioe.2012.12.029>
11. Dalmas NCJ, Bittencourt S, Assmann R, Coraucci D, RicardoSoccol R, Chapter 4: Production of biofuels from algal biomass by fast pyrolysis. *Biofuel Algae.* 2014;143-53. <https://doi.org/10.1016/B978-0-444-59558-4.00007-3>
12. Du, Zhen-Yi & Li, Yecong & Wang, Xiaoquan & Wan, Yiqin & Chen, Qin & Wang, Chenguang & Lin, Xiangyang & Liu, Yuhuan & Chen, Paul & Ruan, Roger. Microwave-assisted pyrolysis of microalgae for biofuel production. *Bioresour Technol.* 2011; 102:4890-96. <https://doi.org/10.1016/j.biortech.2011.01.055>
13. Kapoore RV, Butler TO, Pandhal J, Vaidyanathan S. Microwave assisted extraction for microalgae: from biofuels to biorefinery. *Biotech. crossmark.* 2018;7(99):1-21. <https://doi.org/10.3390/biology7010018>
14. Kumar KS, Dahms HU, Won EJ, Lee JS, Shin KH, Microalgae-a promising tool for heavy metal remediation. *Exotoxicology Env Safety.* 2015; 113:329-52. <https://doi.org/10.1016/j.ecoenv.2014.12.019>
15. Pal D, Khozin GI, Cohen Z, Boussiba S. The effect of light, salinity, and nitrogen availability on lipid production by *Nannochloropsis* sp. *Appl Microbiol Biotechnol.* 2011; 90(4):1429-41. <https://doi.org/10.1007/s00253-011-3170-1>
16. Takeshita T, Ota S, Yamazaki T, Hirata A, Zachleder V, Kawano S. Starch and lipid accumulation in eight strains of six *Chlorella* species under comparatively high light intensity and aeration culture conditions. *Bioresour Technol.* 2014; 158:127-34. <https://doi.org/10.1016/j.biortech.2014.01.135>
17. Mandotra SK, Kumar P, Suseela MR, Nayaka, Ramteke PW. Evaluation of fatty acid profile and biodiesel properties of microalga *Scenedesmus abundans* under the influence of phosphorus, pH and light intensities. *Bioresour Technol.* 2016; 201:222-29. <https://doi.org/10.1016/j.biortech.2015.11.042>
18. Nakajima Y, Tsuzuki M, Ueda R. Improved productivity by reduction of the content of light-harvesting pigment in *Chlamydomonas perigranulata*. *J. Appl. Phycol.* 2001; 13:95-101. <https://doi.org/10.1023/A:1011192832502>
19. Polle JE, Kanakagiri SD, Melis A. tla, a DNA insertional transformant of the green alga *Chlamydomonas reinhardtii* with a truncated light-harvesting chlorophyll antenna size. *Planta.* 2003; 217: 49-59.
20. Perrine Z, Negi S, Sayre RT. Optimization of photosynthetic light energy utilization by microalgae. *Algal Research.* 2012; 1: 134-42. <https://doi.org/10.1016/j.algal.2012.07.002>
21. Cazzaniga S, Dall'Osto L, Szaub J, Scibilia L, Ballottari M, Purton S, Bassi R. Domestication of the green alga *Chlorella sorokiniana*: reduction of antenna size improves light-use efficiency in a photobioreactor. *Biotechnol Biofuels.* 2014; 7(1):157. <https://doi.org/10.1186/s13068-014-0157-z>
22. Perin G, Bellan A, Segalla A, Meneghesso A, Alboresi A, Morosinotto T. Generation of random mutants to improve light-use efficiency of *Nannochloropsis gaditana* cultures for biofuel production. *Biotechnology for Biofuels.* 2015; 8:161. <https://doi.org/10.1186/s13068-015-0337-5>
23. Shin WS, Lee B, Jeong B, Chang YK, Kwon JH. Truncated light-harvesting chlorophyll antenna size in *Chlorella vulgaris* improves biomass productivity. *J Appl Phycol.* 2016; 28:3193-3202. <https://doi.org/10.1007/s10811-016-0874-8>
24. Chekroun. The role of algae in bioremediation of organic pollutants. *Int. Res. J. Pub. Env. Health.* 2014; 1(2):19-32.
25. Lim SL, Wan LC, Siew MP. Use of *Chlorella vulgaris* for bioremediation of textile wastewater. *Biosource Technology.* 2010; 101: 7314-22. <https://doi.org/10.1016/j.biortech.2010.04.092>
26. Salgueiro JL, Perez L, Maceiras R, Sanchez A, Cancela A. Bioremediation of wastewater using *Chlorella vulgaris* microalgae: phosphorus and organic matter. *Int. J. Environ. Res.* 2016; 10:465-70.
27. Jyoti J, Awasthi M. Bioremediation of wastewater Chromium through microalgae: a review. *Int. J. Eng. Res & Tech.* 2014;3(6):1210-15.
28. Chalivendra S, Saikumar. Bioremediation of wastewater using microalgae (electronic thesis or dissertation). Retrieved from <https://etd.ohiolink.edu/2014>

29. Kshirsagar AD. Bioremediation of wastewater by using microalgae: an experimental study. *Int. J. Lifesc. Bt Pharm. res.* 2013; 2:339-46.
30. El-Sheekh MM, Farghl A A, Galal HR, Bayoumi HS. Bioremediation of different types of polluted water using microalgae. *Rend Fis Acc Lincei* 2016; 27:401. <https://doi.org/10.1007/s12210-015-0495-1>
31. Kumar GS, Khan SA. Bioremediation of sewage water using selective algae for manure production. *Int. J. Env. Eng. Man.* 2013; 4(6):573-80.
32. Kassas, Mohammed. Bioremediation of the textile waste effluent by *Chlorella vulgaris*. *The Egypt J Aqua Res* 2014; 40(3):301-08. <https://doi.org/10.1016/j.ejar.2014.08.003>
33. Ramirez ME, Velez L, Rendon, Alzate E. Potential of microalgae in the bioremediation of water with chloride content. *Brazilian J. Biology.* 2017; 78:1-5. <https://doi.org/10.1590/1519-6984.169372>
34. Bwapwa J, Jayeola A, Chetty R. Bioremediation of acid mine drainage using algae strains: A review. *S. African J. Chem. Engineering.* 2017; 24:62-70. <https://doi.org/10.1016/j.sajce.2017.06.005>
35. Hammud HH, Ali El-S, Essam K, El-Sayed M. Adsorption studies of Lead by Enteromorpha algae and its silicates bonded material. *Advances in chemistry.* 2014; Article ID 205459,1-11, <http://dx.doi.org/10.1155/2014/205459>
36. Ismail, Azza AM, Abd El-All, Han AM. Biological influence of some microorganisms on olive mill waste water. *Egypt J.Agric.Res.* 2013; 91:1-11.
37. Chan A, Hamidreza S, McBean. Heavy metal removal (copper and zinc) in secondary effluents from waste water treatment plants by microalgae. *ACS chemistry sus. Chem. Engg.* 2014; 2:130-37. <https://doi.org/10.1021/sc400289z>
38. Zeraatkar AK, Ahmadzadeh H, Talebi AF, Moheimani NR, McHenry MP. Potential use of algae for heavy metal bioremediation, a critical review. *J.Env.Man.* 2016; 181:817-37. <https://doi.org/10.1016/j.jenvman.2016.06.059>
39. Sharma, Khan. Bioremediation of sewage wastewater using selective algae for manure production. *Int. J. Env. Eng Manag.* 2013; 4:573-80.
40. Delrue F, Pablo DAD, Sing SF, Fleury G, Sassi.JF. The environmental biorefinery: using microalgae to remediate wastewater, a win-win paradigm. *Energies.* 2015;9 (132):1-19. <https://doi.org/10.3390/en9030132>
41. Prabha Y, Soni SK, Sharmita G, Sonal. Potential of Algae in bioremediation of Wastewater: Current Research. *Int. J. Curr. Microbial. App. Sci.* 2016; 5:693-700. <https://doi.org/10.20546/ijcmas.2016.502.076>
42. Shahid A, Knan AZ, Liu T, Malik S, Afzal I, Mehmood MA. Algae-based biologically active compounds. Algae based polymers, blends, and composites. *Chem biotech materials Sci.* 2017; 273-99. <https://doi.org/10.1016/B978-0-12-812360-7.00007-0>
43. Hosseini SM, Khosravi-Darani K, Mozafari MR. Nutritional and Medical Applications of Spirulina microalgae Mini-review. *Med. Chem.* 2013; 13:1231-37. <https://doi.org/10.2174/1389557511313080009>
44. Nicoletti M. Microalgae Nutraceuticals. *Foods.* 2016; 5(3):1-13. <https://doi.org/10.3390/foods5030054>
45. Sathasivam R, Radhakrishnan R, Hashem A, Allah E. Microalgae metabolites: A rich source for food and medicine. *Saudi J. Bio Sci.* 2017; 26:709-22. <https://doi.org/10.1016/j.sjbs.2017.11.003>
46. Priyadarshni, Rath B. Commercial and industrial applications of microalgae. A review. *J Algal Biomass Utln.* 2012; 3(4):89-100.
47. Mostafa SM Chapter 12: Microalgal Biotechnology. *Plant Sci.* 2012; 275-314.
48. Sigamani S, Natarajan H. A review on potential biotechnological applications of microalgae. *J. App. Pharma. Sci.* 2016; 6(8): 179-84. <https://doi.org/10.7324/JAPS.2016.60829>
49. Yamaguchi K. Recent advances in microalgal bioscience in Japan, with special reference to utilization of biomass and metabolites: a review. *J Appl Phycol* 1996; 8(6): 487-502. <https://doi.org/10.1007/BF02186327>
50. Bhalamurugan GL, Valerie O Mark L. Valuable bioproducts obtained from microalgal biomass and their commercial applications. A review: *Environ. Eng. Res.* 2018; 23(3):229-41. <https://doi.org/10.4491/eer.2017.220>
51. Wells ML, Potin P, Craigie JS, Raven JA, Merchant SS, Helliwell KE, Smith AG, Camire ME, Brawley SH. Algae as nutritional and functional food sources: revisiting. *J. Appl. Phycol.* 2016; 29(2):949-82. <https://doi.org/10.1007/s10811-016-0974-5>
52. Morais MG, Stillings C, Wendorff J. Biofunctionalized Nanofibres using *Arthrospira* (Spirulina) Biomass and Biopolymer. *Biomed. Res. Int.* 2015; 1-8. <http://dx.doi.org/10.1155/2015/967814>
53. Swain SN, Biswal SM, Nanda PK, Nayak PL. Biodegradable soy-based plastics: opportunities and challenges. *J. Polym. Environ.* 2004; 12 (1): 35-42. <https://doi.org/10.1023/B:JOOE.0000003126.14448.04>
54. Mekonnen T, Mussone P, Khalil H, Bressler D. Progress in bio-based plastics and plasticizing modifications. *J. Mater. Chem. A,* 2013; 1:13379-98. <https://doi.org/10.1039/c3ta12555f>
55. Rajendran N, Sharanya Puppala S, Raj SM, Angeeleena RB, Rajam C. Seaweeds can be a new source for bioplastics. *J. Pharm. Res.* 2012; 5 (3): 1476-79.
56. Zeller MA, Hunt R, Jones A, Sharma S. Bioplastics and their thermoplastic blends from Spirulina and Chlorella microalgae. *J. Appl. Polym. Sci.* 2013; 130, 3263-75. <https://doi.org/10.1002/app.39559>

