



RESEARCH ARTICLE

Iron release characteristics in groundnut growing red and black calcareous soils of Cuddalore district

Porkodi G^{1*}, R Shanmugasundaram², MP Sugumaran³, P Ramamoorthy⁴, K Bharathi Kumar⁵, M Kiruba⁶, R Brindhavathy⁷ & G Anand^{6*}

¹Department of Soil Science and Agricultural Chemistry, Agriculture College and Research Institute, Kudumiyanmalai, Pudukkottai District 622 104, Tamil Nadu, India

²Department of Soil Science and Agricultural Chemistry, Tamil Nadu Agricultural University, Coimbatore 641 003, Tamil Nadu, India

³ICAR-Krishi Vigyan Kendra, Cuddalore 606 001, Tamil Nadu, India

⁴Department of Soil Science and Agricultural Chemistry, Don Bosco College of Agriculture, Ranipettai 631 151, Tamil Nadu, India

⁵Regional Research Station, Vridhachalam, Cuddalore 606 001, Tamil Nadu, India

⁶ICAR-Krishi Vigyan Kendra, Salem 636 203, Tamil Nadu, India

⁷Oil Seed Research Station, Tindivanam 604 002, Tamil Nadu, India

*Email: porkodi.g@tnau.ac.in, anandext@tnau.ac.in



ARTICLE HISTORY

Received: 17 October 2024

Accepted: 23 October 2024

Available online

Version 1.0 : 23 December 2024



Check for updates

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>)

CITE THIS ARTICLE

Porkodi G, Shanmugasundaram R, Sugumaran MP, Ramamoorthy P, Kumar KB, Kiruba M, Brindhavathy R, Anand G. Iron release characteristics in groundnut growing red and black calcareous soils of Cuddalore district. *Plant Science Today*.2024;11(sp4):01-05. <https://doi.org/10.14719/pst.5916>

Abstract

The current study was assessed the nutrient release pattern of iron using different sources of iron fertilizer at various levels in groundnut-growing red and black calcareous soils of the Cuddalore district at regular sampling intervals. The study design was completely randomized block design with different levels of iron sulphate (FeSO₄) at 25 and 50 kg ha⁻¹, iron-enriched farmyard manure (Fe-EFYM) at 37.5 and 50 kg ha⁻¹ and iron-ethylene diamine dihydroxy phenylacetic acid (Fe-EDDHA) at 1 and 5 kg ha⁻¹ three replicates at different sampling intervals of incubation. The release pattern of iron was analyzed. The study revealed that the releasing rate of iron increased with the advancement of the incubation period up to 42 days and thereafter declined in both type of calcareous soil. The maximum release of iron was found with the application of 50 kg FeSO₄ ha⁻¹ as Fe- EFYM, which recorded mean values of 26.92 and 26.13 mg kg⁻¹ on 42nd day in red and black calcareous soils, respectively. Application of Fe-EDDHA at 5 kg ha⁻¹ recorded a gradual increase in DTPA iron availability from 7th day (17.35 mg kg⁻¹ and 12.03 mg kg⁻¹ in red and black calcareous soil, respectively) to 42nd day (21.49 mg kg⁻¹ and 18.87 mg kg⁻¹ in red and black calcareous soil respectively). Then it showed a steady decline in both calcareous soils. The results of the current study showed that the efficient usage of iron in both calcareous soils was greatly increased by the combined application of organic and inorganic fertilizers.

Keywords

calcareous soil; FeSO₄; Fe-EFYM; Fe- EDDHA; iron; nutrient release

Introduction

Iron (Fe) is the fourth most common element in the soil. Among micronutrients, Fe was the first micronutrient identified as essential for plant growth. In the metabolic processes of plants, Fe is necessary for several respiratory enzymes and metabolic activities related to photosynthesis. Legumes extensively use iron for nodule formation to fix atmospheric nitrogen. The protein content of peanut kernels was also increased by Fe fertilization. Green plant tissue contains 50-100 mg of iron per kilogram of dry weight (1). Therefore, Fe is one of the most prominent micronutrients in plants (2).

Mineral Fe is involved in numerous metabolic activities such as photosynthesis and cell respiration. Regardless of the Fe nutritional status, over 80% of the Fe in green leaves is concentrated in the chloroplasts. Iron has the potential to form chelates and can undergo reversible oxidation-reduction reactions, being the two important features underlying its range of physiological effects (1, 3). Fe is not incorporated into the actual chlorophyll structure, although it plays a catalytic role in numerous reactions that lead to chlorophyll biosynthesis. Fe regulates the common precursors of heme and chlorophyll synthesis (4).

Iron can be complexed by organic compounds, which improves its solubility and mobility (5). Compounds that are particularly effective in chelating Fe are found in organic amendments. Humic acids, amino acids, phenolics, hydroxamates and catechol siderophores are the most common chelators of Fe found in organic sources (6). In addition to providing chelators that increase Fe solubility, organic sources also encourage microbial activity, providing effective siderophores (7, 8). Iron sulphate (FeSO_4) is a widely used fertilizer because of its fast solubility and low cost; however, nutrient fixation in the soil is a primary issue. In such situations, other methods, such as the use of synthetic chelates, can be used to increase the lack of micronutrient utilization. To improve iron availability in the soil and correct iron chlorosis in plants, organic manures, Fe chelates, and inorganic fertilizers such as FeSO_4 were applied to iron-deficient soil through soil application and foliar spraying, respectively. In calcareous soil, the application of Fe chelates like Fe-EDDHA, Fe-EDTA and Fe-DTPA was highly effective for sustaining soil solution Fe. With the soil having a high pH and calcareousness, the recovery ability of Fe from FeSO_4 was negligible (9-11). Using organic manures that serve as natural chelates for enrichment, rather than expensive synthetic chelates such as iron-ethylene diamine tetra acetic acid (Fe-EDTA) and iron-ethylene diamine dihydroxy phenyl acetic acid (Fe-EDDHA) Fe-EDDHA, appears to be a cost-effective strategy. There are numerous iron fertilizers for preventing iron chlorosis, but their high cost typically prevents the widespread use of iron sources in agriculture (12). Thus, higher nutrient release is possible by combining manure and inorganic fertilizers.

Few investigations on the various sources and levels of iron release have been published. Consequently, a laboratory incubation experiment was conducted to study the pattern of Fe release after the addition of different sources and levels of iron fertilizer.

Materials and Methods

Incubation studies were carried out in the Department of Soils and Environment, Agricultural and College and Research Institute, Madurai, India, during 2021-2022 to study the release pattern of Fe fractions from different fertilizer sources *viz.*, FeSO_4 , Fe-EFYM and Fe-EDDHA in red and black calcareous soils. The black calcareous soil samples from Magulam village in Mangalur Block and red calcareous soil samples from Arasakuzhi village in Vridhachalam Block in the Cuddalore district were collected and assessed for the availability of iron

in the soil. Soil samples from the above-mentioned areas were collected, processed and utilized for incubation studies. The initial soil analytical report for black and red calcareous soil is shown in Table 1. In black calcareous soil, the soil was silty clay with an alkali pH and the electrical conductivity of the soil was less than 4 dS m^{-1} which indicates that the soil was free from salinity. The organic carbon status was found to be low with a cation exchange capacity of $34.5 \text{ c mol (p+) kg}^{-1}$. The fertility status was found to be low in available nitrogen, medium in available phosphorus and high in available potassium. The available iron status of the soil was below its critical level.

Table 1. Initial soil properties of the experimental soil

Parameters	Black calcareous soil	Red calcareous soil
Textural class	Silty Clay	Sandy Clay Loam
Soil reaction (1: 2.5 Soil: water)	8.35	8.67
Electrical conductivity (dS m^{-1})	0.82	0.49
Organic carbon (g kg^{-1})	3.60	3.96
Free calcium carbonate (%)	11.5	11.9
Available nitrogen (kg ha^{-1})	257.6	242
Available phosphorus (kg ha^{-1})	15.6	10.54
Available potassium (kg ha^{-1})	336	276
Available iron (ppm)	5.28	5.02

The farmyard manure was physically mixed @1:10 with FeSO_4 and manure for enrichment in a plastic bag. The component was properly combined and moistened to the required amount using deionized water to ensure moist friable manure. The FYM were mixed with $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ and the moisture level was maintained at about 60% during the enrichment process (13). The polyethylene bags were clipped with rubber bands and placed on a laboratory worktable at room temperature for a month. To speed up decomposition, the mixture was rotated once a week, kept damp and given two hours to release CO_2 and relieve any CO_2 stress on microbial activity (14). The polyethylene bags were additionally weighed every three days, the weight recorded and the difference between the two weights was made up by adding deionized water to maintain a constant moisture level. The composition of FYM and Fe-EFYM is presented in Table 2. A 500 ml container containing 200 g of soil samples was used to study the laboratory incubation research for the soil's Fe release pattern. Under laboratory conditions, soil samples were periodically maintained at field capacity (21% gravimetric).

Soil samples were drawn from each container at seven days intervals (7, 14, 21, 28, 35, 42, 49, 56 and 63 days of incubation) and analyzed for soil DTPA-Fe using standard procedures (15). The incubation study was conducted using a completely randomized design that was replicated thrice with different treatment combinations, *viz.*, T1 - Control, T2 - 25 kg ha^{-1} FeSO_4 , T3- 50 kg ha^{-1} FeSO_4 , T4 - FeSO_4 as Fe-enriched farmyard

Table 2. Composition of FYM and Fe-EFYM

Nutrients	Content (%)	
	FYM	Fe-EFYM
Nitrogen	0.76	1.24
Phosphorus	0.69	0.72
Potassium	0.67	0.73
Zinc	0.0005	0.0059
Copper	0.0006	0.00062
Manganese	0.009	0.018
Iron	0.09	0.6

manure @ 37.5 kg ha⁻¹, T₅ - FeSO₄ as Fe-enriched farmyard manure @ 50 kg ha⁻¹, T₆ - 1 kg ha⁻¹ Fe- EDDHA, T₇ - 5 kg ha⁻¹ Fe- EDDHA.

The data obtained from the incubation study were subjected to statistical analysis using AGRSS software version 7.01. The level of significance was set at P < 0.05. Critical difference (CD) values were calculated for the P < 0.05 whenever the “F” test was found significant (16).

Results

To study the availability of Fe in red and black calcareous soils, an incubation experiment was conducted with different sources of Fe (FeSO₄, Fe-EFYM and Fe-EDDHA) and different levels of Fe on the availability of Fe at different incubation time intervals. The results revealed that different sources and levels of Fe had a significant effect on the release pattern of nutrients at different incubation intervals.

The treatments comprised of control (T₁), 25 kg FeSO₄ ha⁻¹ (T₂), 50 kg FeSO₄ ha⁻¹ (T₃), 37.5 kg FeSO₄ ha⁻¹ as Fe-EFYM (T₄), 50 kg FeSO₄ ha⁻¹ as Fe-EFYM (T₅), 1 kg ha⁻¹ Fe-EDDHA (T₆) and 5 kg ha⁻¹ Fe-EDDHA (T₇). The samples were incubated for 65 days at the field capacity level. Once a week, soil samples were collected and over time, the available iron was determined.

The findings showed that increasing levels of iron greatly increased the release of iron in red and black calcareous soils over time (from the 7th days to the 42nd days of incubation). Using different sources of iron, iron availability increased with time and levels of iron. In both calcareous soils, the amount of available iron increased with the advancement of the incubation period up to the 42nd day and decreased from the 49th day onwards.

Increasing levels of Fe addition through different sources increased Fe availability and the highest available Fe was recorded with the addition of 50 kg FeSO₄ ha⁻¹ as Fe-EFYM (T₅), followed by the addition of 37.5 kg FeSO₄ ha⁻¹ as Fe-EFYM (T₄). The rate of iron release was higher in treatment T₅ (50 kg FeSO₄ ha⁻¹ as Fe-EFYM) until 42nd day and thereafter declined from the 49th days. The maximum release of iron was found to be with the application of 50 kg FeSO₄ ha⁻¹ as Fe-EFYM, with recorded values of 26.92 and 26.13 mg kg⁻¹ in red and black calcareous soils, respectively.

The maximum mean of iron release was found in T₅ (50 kg FeSO₄ ha⁻¹ as Fe-EFYM) and the mean values ranged from 24.59 to 23.88 mg kg⁻¹ in red and black calcareous soils. Next to this treatment, T₄ (37.5 kg FeSO₄ ha⁻¹ as Fe-EFYM) recorded highest available iron concentration of 22.11 and 20.70 mg kg⁻¹ in red and black calcareous soil respectively followed by T₃, T₇, T₂ and T₆. The addition of FYM had a significant influence on the amount of iron released. While FeSO₄ applied at 25 kg ha⁻¹ (T₂) and 50 kg ha⁻¹ (T₃) showed a mean value of 17.14 and 20.98 mg kg⁻¹ of available iron content in red calcareous soil, respectively. In black calcareous soil registered a mean value of 12.66 and 17.09 mg kg⁻¹ of available Fe content in black calcareous soil supplied 25 kg FeSO₄ ha⁻¹ (T₂) and 50 kg FeSO₄ ha⁻¹ (T₃) respectively.

Among the treatment T₆ and T₇, addition of Fe-EDDHA at the rate of 5 kg ha⁻¹ (T₇) registered the maximum mean

available iron content of 19.07 and 15.57 mg kg⁻¹ in red and black calcareous soils respectively. Followed by this, the treatment applied with 1 kg of Fe- EDDHA (T₆) recorded 15.97 and 10.57 mg kg⁻¹ in red and black calcareous soil.

In control, the lowest availability of Fe content was observed, which recorded the values of 5.20 and 4.66 mg kg⁻¹ in red and black calcareous soils respectively. A gradual decrease was observed during the incubation period.

Discussion

The experiment on the release pattern of Fe nutrients from the applied sources provides a thorough understanding of the applicability and performance of additional fertilizers in crop production. Understanding how micronutrients are converted in the soil through the application of enhanced manure (FYM) is crucial because it affects the availability of these essential and significant plant nutrients. Most of the Fe required by the plants must be supplied by the soil fertilizer reaction products, not the applied water-soluble micronutrient fertilizer. This assumes that the significance of Fe transition in soil is true. A study was conducted on the Fe-release patterns of FeSO₄, Fe-EFYM and Fe-EDDHA. Modified soils were evaluated over time to determine whether they could support agricultural production.

In red and black calcareous soil, Fe release was at its maximum with increasing time intervals up to 42 days before declining. With the application of 50 kg FeSO₄ ha⁻¹ as Fe-EFYM, the maximum release of Fe in red and black calcareous soils was 26.92 and 26.13 mg kg⁻¹ at 42nd days, respectively (Fig. 1 & 2). The availability of Fe increased with increasing levels of Fe through FYM and the highest DTPA Fe status was recorded with the addition of 50 kg Fe-EFYM. The addition of iron-containing fertilizers and the mineralization and release of Fe from organic matter during decomposition have been shown to significantly increase the availability of Fe in the soil. A rise in soil microbial activity brought on by additional organic material addition may have contributed to the release of Fe from complex organic compounds (17-19). The formation of organometallic complexes with Fe may have decreased its fixation by clay minerals or its precipitation as FePO₄, increasing the availability of Fe with the addition of organic matter (20).

DTPA-Fe had a more noticeable impact than other critical elements, which may be associated with the residual Fe that was left behind in the soil after the crop was harvested in proportion to the depletion. Several researchers have noted

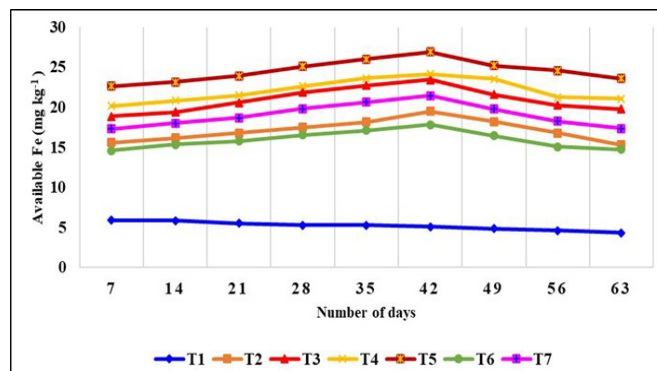


Fig. 1. Effect of levels and source of iron on releasing pattern of iron in red calcareous soil

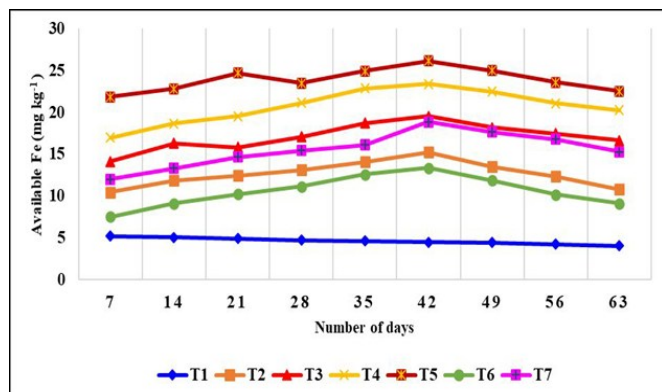


Fig. 2. Effect of levels and source of iron on releasing pattern of iron in black calcareous soil

that the addition of iron to soil results in higher iron concentrations (21-23).

Because FYM is responsible for generating a variety of organic acids that may have acted as chelating agents and prevented the release of iron from accumulating with calcium in the soil, the increase in iron due to FYM may be attributed to their chelating effect. Furthermore, it is consistent with the findings of Katyal *et al.*, (24) who found that the organic acids or chemicals produced during the breakdown of organic matter react with iron to form soluble organo-iron complexes that prevent iron from being fixed by soil components. The increased iron availability might be due to the role of humic substances in the manure, which increased iron availability by preventing their transformation into insoluble hydroxides, chelation action, and release in labile forms (25). A similar finding was also reported in soil by Latha *et al.*, (26) and who found a significant influence of Fe-fortified vermicompost on iron availability due to the complexation or chelation action, contribution from manures and prevention of iron fixation (27, 28).

Conclusion

From this study, it was concluded that with the combined application of organic and inorganic fertilizers, an increase in iron availability was observed in black and red calcareous soil. On the 42nd day, the DTPA-Fe availability was higher when the Fe was applied with an organic source, that is, the application of 50 kg of FeSO₄ ha⁻¹ as Fe-EFYM recorded maximum availability both in red and black calcareous soil.

Acknowledgements

I thank Dr. R. Shanmugasundaram for mentoring and providing technical support for this study and the authors are grateful to the Agriculture College and Research Institute, Madurai for providing research area and lab facilities.

Authors' contributions

GP and RS conceived of the project and designed the experiments. GP and RS analysed the data. RS, MPS and PR assisted the data, prepared the figures and tables and prepared the manuscript. KB, GA and MK validated the statistical data All authors approved the final version of the manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interest to declare.

Ethical issues: None

References

- Mengel K, Kirkby EA, Kosegarten H, Appel T. Further elements of importance. Principles of plant nutrition. 2001:639-55. https://link.springer.com/chapter/10.1007/978-94-010-1009-2_19
- Bauer P, Hell R. Translocation of iron in plant tissues. In Iron nutrition in plants and rhizospheric microorganisms 2006;25:279-88. Dordrecht: Springer Netherlands. https://link.springer.com/content/pdf/10.1007/1-4020-4743-6_13?pdf=chapter%20toc
- Marschner H. Mineral nutrition of higher plants 2nd edn. Institute of Plant Nutrition University of Hohenheim: Germany. 1995.
- Pushnik JC, Miller GW. Iron regulation of chloroplast photosynthetic function: mediation of PS I development. Journal of Plant Nutrition. 1989;12(4):407-21. <https://doi.org/10.1080/01904168909363962>
- Cesco S, Nikolic M, Romheld V, Varanini Z, Pinton R. Uptake of Fe from soluble Fe-humate complexes by cucumber and barley plants. Plant and Soil. 2002;241(1):121-8. <https://doi.org/10.1023/A:1016061003397>
- Chen L, Dick WA, Streeter JG, Hoitink HA. Fe chelates from compost microorganisms improve Fe nutrition of soybean and oat. *Plant and Soil*. 1998;200(2):139-47. Available from: <https://doi.org/10.1023/A:1004375430762>
- Yehuda Z, Shenker M, Romheld V, Marschner H, Hadar Y, Chen Y. The role of ligand exchange in the uptake of iron from microbial siderophores by gramineous plants. *Plant Physiology*. 1996;112(3):1273-80. <https://doi.org/10.1104/pp.112.3.1273>
- O'Hallorans JM, Lindemann WC, Steiner R. Iron characterization in manure amended soils. *Communications in Soil Science and Plant Analysis*. 2005;35(15-16):2345-56. <https://doi.org/10.1081/CSS-200030671>
- Jaloud AA, Al Rabhi M, Bashour I. Availability and fractionation of trace elements in arid calcareous soils. *Emirates Journal of Food and Agriculture*. 2013;25(9):702-12. <https://pdfs.semanticscholar.org/375/357a38445ee2bbc543d1a5cb70d8718a565.pdf>
- Faraz A, Maqsood MA, Aziz T, Cheema MA. Water soluble iron (Fe) concentration in alkaline and calcareous soils influenced by various Fe sources. *Pakistan Journal of Agricultural Sciences*. 2014;51(2):407-11. <https://www.cabidigitallibrary.org/doi/full/10.5555/20153260729>
- Safarzadeh S, Kasmaei LS, Abadi ZA. Effect of organic substances on iron-release kinetics in a calcareous soil after basil harvesting. *Journal of the Serbian Chemical Society*. 2018;83(10):7-8. <https://doi.org/10.2298/JSC171115019S>
- Banijamali SM, Feizian M, Alinejadian Bidabadi A, Mehdipour E. Effect of magnetite nanoparticles on vegetative growth, physiological parameters, and iron uptake in *Chrysanthemum morifolium* 'Salvador'. *Journal of Ornamental Plants*. 2019;9(2):129-42. https://journals.iau.ir/article_665138_5eb80c10b335130a336ff25148a44d56.pdf
- Laan P, Smolders A, Blom C. The relative importance of anaerobiosis and high iron levels in the flood tolerance of *Rumex* species. *Plant and Soil*. 1991;136(2):153-61. <https://doi.org/10.1007/BF02150046>
- Regar KL, Yadav J. Effect of enriched FYM and P2O5 levels on physio-chemical properties of soil after harvesting of rice. *International Journal of Chemical Studies*. 2019;7(1):2520-6. <https://www.academia.edu/download/81465803/7-1-509-691.pdf>
- Lindsay WL, Norvell W. Development of a DTPA soil test for zinc,

- iron, manganese, and copper. *Soil Science Society of America Journal*. 1978;42(3):421-8. <https://doi.org/10.2136/sssaj1978.03615995004200030009x>
16. Gomez KA, Gomez AA. Statistical procedures for agricultural research. *John Wiley & Sons*. 1984.
 17. Chen Z, Wang Y, Xia D, Jiang X, Fu D, Shen L, Wang H, Li QB. Enhanced bioreduction of iron and arsenic in sediment by biochar amendment influencing microbial community composition and dissolved organic matter content and composition. *Journal of Hazardous Materials*. 2016;311:20-9. <https://doi.org/10.1016/j.jhazmat.2016.02.069>
 18. Poonia T, Kumar S, Kumawat SM. Crop management practices influence the nodule characteristics, yield attributes, and yield of groundnut: Groundnut yield influence by crop management practices. *Journal of Agri Search*. 2022;9(1):16-23.
 19. Priyadarshini P, Chitdeshwari T, Sudhalakshmi C. Iron availability in calcareous and non-calcareous soils as influenced by various sources and levels of iron. *Madras Agricultural Journal*. 2019;106(1):12-24. <https://doi.org/10.29321/maj.2019.000265>
 20. Kumar M, Kar A, Raina P, Singh SK, Moharana PC, Chauhan JS. Spatial variability of available soil nutrients in the Sekhawati region of Thar Desert, India. *Journal of the Indian Society of Soil Science*. 2019;67(1):21-33. [10.5958/0974-0228.2019.00003.3](https://doi.org/10.5958/0974-0228.2019.00003.3)
 21. Cakmak I, Pfeiffer WH, McClafferty B. Biofortification of durum wheat with zinc and iron. *Cereal Chemistry*. 2010;87(1):10-20. <https://doi.org/10.1094/CCHEM-87-1-0010>
 22. Dhaliwal SS, Sadana US, Khurana MPS, Dhadli HS, Manchanda JS. Enrichment of rice grains with zinc and iron through ferti-fortification. *Indian Journal of Fertilisers*. 2010;6(7):28-35. <https://www.cabidigitallibrary.org/doi/full/10.5555/20103236091>
 23. Bairwa S, Yadav PK. Influence of FYM, inorganic fertilizers, and micronutrients on soil nutrient status and plant nutrient contents and their uptake by African marigold (*Tagetes erecta* Linn.). *International Journal of Current Microbiology and Applied Sciences*. 2017;6(2):1362-70.
 24. Katyal JC, Sharma BD. DTPA-extractable and total Zn, Cu, Mn, and Fe in Indian soils and their association with some soil properties. *Geoderma*. 1991;49(1-2):165-79. [https://doi.org/10.1016/0016-7061\(91\)90099-F](https://doi.org/10.1016/0016-7061(91)90099-F)
 25. Bocanegra MP, Lobartini JC, Orioli GA. Plant uptake of iron chelated by humic acids of different molecular weights. *Communications in Soil Science and Plant Analysis*. 2006;37(1-2):239-48. <https://doi.org/10.1080/00103620500408779>
 26. Latha MR, Savithri P, Indirani R, Kamaraj. Influence of zinc-enriched organic manures on the yield, dry matter production, and zinc uptake of maize. *Acta Agronomica Hungarica*. 2001;49(3):231-6. <https://doi.org/10.1556/AAgr.49.2001.3.3>
 27. Mane JT, Jagtap PB, Patil WV. Effect of different manures on periodical availability of zinc and iron in calcareous soil. *Journal of Pharmacognosy and Phytochemistry*. 2020;9(6):191-4.
 28. Meena M, Jegadeeswari D, Selvi D, Sankari A. Fortification of organic manures with iron on nutrient release characteristics. *International Journal of Plant & Soil Science*. 2022;34(19):312-20. <https://doi.org/10.9734/ijpss/2022/v34i1931118>