



REVIEW ARTICLE

Botanicals for managing insect pests in rice: An eco-friendly strategy for sustainable rice production

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Abstract

Rice, a staple crop for over half the global population, faces severe yield losses due to insect pests, such as the rice stem borer (Scirpophaga incertulas), brown planthopper (Nilaparvata lugens) and leaf folder (Cnaphalocrocis medinalis) and minor insect pests, which are significant threats to global food security. Traditional reliance on chemical pesticides for pest control has negatively impacted the environment, human health and non-target organisms. This has necessitated a shift toward sustainable pest management strategies that minimize chemical inputs. Botanicals, derived from plants as essential oils, extracts and secondary metabolites, have emerged as an ecofriendly alternative due to their biodegradability, targeted efficacy and reduced environmental footprint. These plant-based compounds act through various mechanisms, including antifeedant activity, growth inhibition, oviposition deterrence, ovicidal effects and toxicity, effectively disrupting pest lifecycles without harming non-target species. Studies underscore the potential of botanicals in suppressing pest populations and mitigating crop losses, making them vital components of Integrated Pest Management (IPM) strategies. When incorporated into IPM programs alongside biological control agents, cultural practices and resistant rice varieties, botanicals enhance pest control efficacy while preserving ecosystem balance and promoting biodiversity. Their use aligns with sustainable agriculture principles, offering a viable path to reduce pesticide dependency and ensure long-term agricultural resilience. This review highlights the critical role of botanicals in IPM for rice cultivation, emphasizing their potential to mitigate pest impacts while supporting environmentally sustainable and economically viable food production systems.

Keywords

botanicals; eco-friendly pest management; key pests; natural enemies; rice

Introduction

Rice, *Oryza sativa* (L), belonging to the family Poaceae, is one of the major staple food crops in the world, farming the staple diet for half of the worlds' population with the production of 523.9 million tonnes in an area of 166.31 million hectares. In India, rice production 2023 reached 1308.37 lakh tonnes, occupying 40 to 43 % of the total cultivable area (1). Rice is widely cultivated in tropical and sub-tropical regions and its significance as a food crop has earned it the title "King of Cereals"(1). Rice is recognized as a nutri-

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tious grain devoid of fats and cholesterol, making it an ideal component of a well-rounded diet (2). It serves as a rich source of carbohydrates, vitamins, proteins, calcium, thiamine, niacin, iron, riboflavin and fibre, with low sugar. Its gluten-free nature renders it a crucial option for individuals adhering to gluten-free diets and those with diabetes (2).

More than 175 species of insect pests have been identified on rice crops. Nearly 20 insect species are known to be critical in rice crops and are regularly noticed in tropical and subtropical regions in Asia (3). More than 25 % yield loss was recorded due to rices' infestation of various insect pests. Several previously considered minor pests have become prominent, causing severe yield loss in rice. The major insect pests of rice include yellow stemborer (Scirpophaga incertulas), leaf folder (Cnaphalocrocis medinalis), brown plant hopper (Nilaparvata lugens), whitebacked planthopper (Sogatella furcifera), gall midge (Oroseolia oryzae) and rice ear head bug (Leptocorisa acuta) (4). Rice farmers mostly prefer to use synthetic insecticides to control insect pests. However, continuous and indiscriminate use of insecticides has led to the development of resistance and resurgence of pest populations and residues in the final produce. Therefore, finding other alternatives for chemical pesticides is urgently needed. Botanicals may be the best insecticide alternatives due to their eco-friendly (5). Botanicals may also be considered plant protection inputs for future rice production (6).

Botanicals may improve the physical stability of food products and their applications enhance the stability of rice-based foods by inhibiting oxidative reactions (7). The bioactive molecules present in several plant extracts act as toxicants, repellents, phagodeterrents, ovideterrents, growth regulators, etc., offering viable alternatives to conventional chemical pesticides (8). Botanicals are cost-effective, target-specific, biodegradable and environmentally friendly, with a broad spectrum of activity and minimal harmful effects (9). Integrating botanicals for nutrient management with pesticide properties has been shown to enhance rices' growth and yield parameters (10). Despite regulatory and market challenges, botanicalbased biopesticides have proven to be a safer, eco-friendly and sustainable pest management option for rice (11). Botanicals have been used in Indian agriculture for over a century to minimize the substantial yield losses caused by most insect pests in rice (12).

Overview of Botanicals in Pest Management

Botanicals have been derived from plant materials that act as insecticides and repellents, bactericides, fungicides, herbicides and nematicides. Around the world, there are about 2,121 plant species have been reported to have pest control capabilities, including 1,005 species with insecticidal, 384 antifeedant, 297 repellent and 31 growth-inhibiting properties, of which only a few have been validated and commercialized (13). Around 735 botanical-based pesticide products have been registered by various companies. Among them, 443 were Azadirachtin and 290 were Pyrethrum-based products. Plant extracts and their

secondary metabolites, such as nicotine, Pyrethrum, rotenone, derris and sabadilla, are examples of "first generation" insecticides known as "botanicals" (13). Some of these plant chemicals lay the groundwork for developing synthetic "second-generation" insecticides. For example, Pyrethrum extracted from Chrysanthemum flowers has become the fundamental unit for developing synthetic pyrethroids (14). These second-generation insecticides are nerve toxins affecting both the target and non-target organisms. Neem-based products are considered second-generation insecticides, extensively used in managing various insect pests.

Among botanicals, neem products have an imperative role in IPM strategies of crops (15). Using plant products in pest management has a long history and is an essential indigenous traditional pest management practice in India (16). Botanicals can have tri-trophic effects, offering pest control equivalent to synthetic insecticides. Additionally, their proximity to natural enemies like predators and parasitoids indicates their potential for managing insect pests in sustainable organic agriculture (17).

Impact of Botanicals on Borers and Defoliators in Rice Stem borer complex

Among the various insect pests of rice, stem borers are considered the most dangerous. Three species of the Noctuidae family and eighteen species of the Pyralidae family of stem borer have been documented in India (18). The yellow stem borer (Scirpophaga incertulas) is a monophagous pest that damages all rice growth stages (19). The rice stems borer larvae bore into the stem of the rice plant and arrest the supply of water and nutrients to the upper part of the plants, which resulted in "Dead hearts" during the vegetative stage and "white ears" during the reproductive stage (20). The plants' compensatory mechanism will compensate the dead hearts at the vegetative stage of the crop, but it was not possible at the reproductive stage, in which 4 % yield loss was observed for every 1 % of the white ear (21). At severe epidemic outbreaks, up to 63 % of tillers were damaged, 65 % of dead heads, and 95 % of white heads were observed (22). The yield losses by yellow stem borer alone ranged from 1% to 19 % at the early stage and 38-80 % at the late stage in rice (23).

Among various botanicals tested against stem borers, the application of Neem seed kernel extract (5 %) and a combination of neem oil (1 %) and pungam oil (1 %) recorded the lowest incidence of dead heart symptoms at 4.07 and 4.93 %, respectively in organic rice (24). Botanical extracts prepared from *Azadirachta indica, Vitex negundo, Ipomoea carnea and Adhatoda zeylanica* resulted in decreased rice stem borer population and increased larval mortality (25). The efficacy of extracts derived from *Neorautanenia mitis* and *Derris elliptica* was effective against the rice stem borer *Chilo partellus*(26). Three botanical leaf extracts *viz.*, tobacco, neem and karonja applied @ 15mL/L showed considerable population reduction in stem borers under field conditions with reduced dead hearts (38.38 %) and whiteears (58.08 %)(27) (Fig. 1).

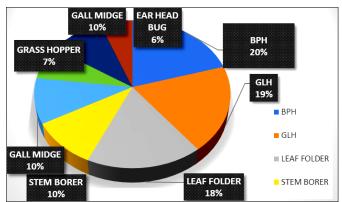


Fig 1. Comparative efficacy botanicals against significant insect pests of rice.

The neem seed kernel extract application @ 5 % concentration significantly reduced the dead hearts and white ears due to rice stem borer (28). Azadirachtin, derived from neem, acts as a potent insect growth regulator that interferes with the moulting process of stem borer larvae. Similarly, pongam extracts contain flavonoids and limonoids that exhibit insecticidal properties against rice stem borers (29). Garlic extracts, rich in sulfur compounds, act as repellents and disrupt the feeding behaviour of stem borers (30). The plant extracts with animal byproduct viz., cow urine + Azadirachta indica, cow urine + Melia azedarach, cow urine + Lantana camara, cow urine + Jatropha gossypiflora, cow urine + eucalyptus oblique, cow urine + Cannabis sativa and Azadirachtin were found to effective against rice stem borer and among them, result suggested that neemoz @ 2500 mL/ha was the most effective in reducing the infestation of stem borer, S. incertulas in paddy followed by cow urine + Melia azedarach @ 7500 mL/ha (31). The application of neemarin @ 1500 ppm @ 3 L/ha reduced (5.60 %) of white ears in rice (32). Foliar application of nimbecidine and neemarin was efficient and costeffective in decreasing the occurrence of S. incertulas in rice (33).

Rice gall midge

The gall midge, *Orseolia oryzae*, is often known as Wood-Mason, which results in an annual yield loss of 0.8 % of the overall production (34). Rice gall midges cause severe damage by forming a hollow cavity or tubular gall at the base of the infested tiller. The increased application of urea significantly increased the incidence of gall midges (35). The metabolic changes in rice plants due to gall midge feeding in different rice varieties were studied and they found that the metabolites could be considered a biomarker for insect-plant interaction, providing insights into the biochemical responses of rice plants to gall midge infestation and its impact on yield (36).

Plant-based essential oils like eucalyptus and cedarwood oil effectively suppressed gall midge incidence in rice (36). The efficacy of neem seed kernel extracts against the African rice gall midge was studied (35) and they found that the NSKE significantly reduced the gall midge damage and increased the grain yield. Applying neem oil @ 3 % significantly reduced the damage caused by rice gall midges(37). Spraying of neem oil @ 5 % significantly decreased

the gall midge infestations in rice (38). A foliar spray of neem oil @ 2 % applied thrice @ 25, 40 and 55 days after transplantation substantially diminished the population of rice gall midges (39). The application of various neem derivatives was effective against cecidomyiid pests in rice (40) (Fig. 1).

Leaf folder

The rice leaf folder belonging to the order Lepidoptera represents a significant defoliating pest affecting various stages of the rice crop. This pest emerged seriously across Asian countries, leading to considerable yield losses, particularly in tropical and subtropical regions (41). Eight species of rice leaf folders have been recorded among them. Cnaphalocrocis medinalis is the most prevalent one, which caused substantial yield losses in rice (42). Leaf folders cause damage by folding rice leaves and feeding within, resulting in reduced photosynthetic area that can lead to significant yield loss. Moreover, the damaged leaves become vulnerable entry sites for fungal and bacterial pathogens (41). Yield losses due to leaf folders were ranged from 3 % to 10 % (43). A positive correlation between the extent of leaf damage and yield losses due to rice leaf folder indicated that 17.5 % of leaf damage led to a 16.5 % reduction in grain yield. In comparison, a higher damage rate of 26.6 % of leaves corresponded to a 21.3 % loss in yield was observed (44).

The percent reduction over control for leaffolder was more (76.23) in NSKE @ 5 %, followed by neem oil (1 %) + pungam oil (1 %), neem oil (2 %) alone (68.44) and pungam (2 %) oil alone (57.07) (45). A Foliar application with a combination of seaweed (Sargassaum wightii) extract (8 %) with neem leaf extract (5 %) exhibited the highest mortality of 66.66 % leaf folder larvae (46). Azadirachtin was most effective in reducing the incidence of leaf folders compared to untreated controls, showcasing the potential of neem-based products in managing rice leaf folder populations effectively under field conditions (47). Azadirachtin and neem extracts affect the rice leaf folder larvae and reduce food ingestion, detoxifying the enzyme activity of dysoxylum triterpenes (48). The combination of botanical insecticides and bacterial toxins affected the digestive enzymes like protease, amylase and lipase in rice leaf folder larvae, showing a synergistic effect at low doses, decreases in enzyme activity, indicating impaired larval digestion and nutrition, which could be an effective strategy for controlling leaf folder in rice (49).

Application of oil-based formulations extracted from the seeds of Custard apple (*Annona squamosa*), Punnai (*Calophyllum inophyllum*) and Mahua (*Madhuca indica*) each @ 1 % concentration reduced the rice leaf folder infestation (50). Neem seed kernel extract(NSKE) @ 5 % showed juvenile hormone mimic action and reduced the development of *C. medinalis* larvae when mixed with 0.16 percent teepol (51). Neem oil applied @ 1 % concentration on rice plants decreased the occurrences of leaf folder (52). Alternatively, neem cake (de-oiled) amendment in the soil at 150 kg/ha followed by neem oil (2 %) spray at 10-day intervals was found effective against leaf folder (53).

The extract from custard apple leaves showed decreased rice leaf folder infestation (50). Research indicates that the juvenile hormone mimicry activity of Neem Seed Kernel Extract (NSKE) and the inhibitory effect of NSKE against the growth and development of leaf folder larvae (Table 1).

Botanicals Against Sucking Pests in Rice and their Impact on Yield

Brown plant hopper

The brown plant hopper (BPH), *Nilaparvata lugens*is an important sucking pest in rice (56). It directly damages rice

Table 1. Botanicals for the management of borers and defoliators in rice

S.No.	Botanical plant	Plant part	Mode of action				
Stem borer complex							
1	Neem (Azadirachta indica)	Seeds, leaves, Extracted as neem oil,	Antifeedant, growth inhibitor, oviposition deterrent				
2	Datura (Datura alba)	leaf extract	Demonstrated contact toxicity				
3	Ginger (Zingiberofficinale)	Rhizome extract	Insecticidal				
4	Castor (Ricinus communis)	Leaf extract	Repellent and deterrent				
5	Marigold (<i>Tagetes</i> sp.)	Aqueous root extract	Toxicity				
6	Tobacco (Nicotiana tabacum)	Tobacco stems	Insecticidal				
7	Garlic (Allium sativum)	Garlic oil	Repellent				
8	Cedarwood (Cedrus sp.)	Cedar oil	Antifeedant				
9	Chinaberry (Melia azedarach)	Seed oil	Antifeedant and insecticidal activity				
10	Chinaberry (Meliatoosendan)	Seed oil	Antifeedant/ Insecticidal activity				
11	Puna oil tree (Pongamiaglabra)	Mixture of Pongamia and neem oil (1:1)	Antifeedant and insecticidal activity				
12	Chinese azalea (Rhododendron molle)	Root leaf, flower aqueous and alcohol	Toxicity				
13	Thunder God vine (Tripteryguimwiliford)	Root and bark powder Toxicity and Antifeedant activity					
14	Karanja (<i>Pongamia pinnata</i>)	Seed oil and extracts Antifeedant, insect growth regulator					
15	Sweet flag (Acorus calamus)	Rhizomes	Neurotoxicant, repellent				
16	Chinese chaste tree (Vitex negundo)	Leaves and extracts	Antifeedant and ovicidal activity				
17	Tulasi (Ocimum sanctum)	Leaves and essential oil	Repellentand antifeedant				
Gall midge							
1	Neem (Azadirachta indica)	Seeds, leaves, Azadirachtin and other Insect growth regulator; affects moulting and la limonoids development					
2	Chinaberry (Melia azedarach)	Fruits, leaves and Meliatoxins, 2 %	6 Antifeedant, larvicidal				
3	Sweetflag (Acorus calamus)	Rhizomes and β-Asarone Repellentand oviposition deterrent					
4	Garlic (Allium sativum)	Bulbs, Allicin and Diallyl disulfide	Fumigant and antifeedant				
5	Tobacco (Nicotiana tabacum)	Leaves and Nicotine Neurotoxic to insects					
6	Citronella (Cymbopogon nardus)	Leaves, Citronellal and Geraniol Repellent affects larval feeding					
7	Turmeric (Curcuma longa)	Rhizomes and Curcumin Oviposition deterrent, larvicidal					
Defoliators	3						
1	Neem (Azadirachta indica)	Seeds, leaves extracted as neem oil and Azadirachtin	Antifeedant, growth inhibitor, oviposition deterrent				
2	Garlic (Allium sativum)	Cloves, Aqueous and oil extracts and	Antifeedant, repellent, toxic to larvae				
3	Tobacco (<i>Nicotiana</i> sp.)	Leaves and Nicotine sulfate extract Neurotoxin, antifeedant					
4	Chilli (<i>Capsicum</i> sp.)	Fruits and Capsaicinoid extracts	Repellent, toxic to larvae, antifeedant				
5	Pyrethrum (Chrysanthemum cinerariifolium)	Flowers and pyrethrin extract	Neurotoxin, knockdown effect				

Other defoliators

Six botanical preparations *viz.*, neem oil, mahogany oil, a mixture of neem and mahogany oil, bishkatali leaf extract, pitraj leaf extract and a mix of bishkatali and pitraj leaf extract were tested against rice his (*Dicladispa armigera*). among these, neem oil was most effective in controlling the pest (54). Neemazal, eucalyptus oil, lemongrass oil, cedarwood oil and camphor oil were evaluated to manage rice grasshoppers (*Oxyanitidula*) in rice. The neemazal was highly effective (Fig. 1) (55).

crops by feeding on the phloem and indirectly by transmitting viral diseases, such as grassy stunt and wilted stunt viruses (57). In rice-growing areas of India, frequent crop failures were attributed to BPH outbreaks (58). The BPH outbreaks resulted in "Hopper burn" symptoms in the main field, leading to significant yield losses (59). The yield loss by BPH in rice ranges from 10 to 90 % (60). The application of NSKE @ 5 % and a combination of neem oil @ 1 % and pungam oil @ 1 % were most effective against BPH,

with a more than 50 % reduction in BPH over the control (24). The BPH population was reduced when NSKE @ 5 % and Vitex leaf extracts @ 10 % were sprayed (61).

A significant decrease in BPH nymph emergence was observed with 2 % neem seed kernel extract and 1 % karanj oil foliar application (62). The extracts derived from Ageratum conyzoides, Barringtonia asiatica, Melia azedarach and Tephrosia vogelii caused significant mortality in brown planthopper populations (63). Applying eucalyptus oil @ 1000 mL/ha was effective against BPH and resulted in higher grain yield (64). The extracts of Piper retrofractum and Tagetes erecta show the highest mortality and feeding inhibition in the brown plant hopper population (65). Pyrethrum application increased BPH mortality within four days of application (66).

Neem seed kernel extract at 7.5 % reduced BPH population and increased grain yield (9). Spraying of Vitex leaf extract at 5 % showed a good knockdown effect against BPH under laboratory conditions (67). Multineem 300 ppm applied @ 1.25 L/ha was highly effective for managing BPH in rice (68). Applying 5 % bitter gourd leaf extract, 5 % garadi leaf extract and 5 % custard apple leaf extract was highly effective for BPH in rice, resulting in more nymphal mortality and higher feeding inhibition (69). The BPH was highly susceptible to neem oil, resulting in reduced food intake and nymphal mortality (70). Application of neem oil 3 % and NSKE 5 % was found superior to chemical pesticides against Nilaparvata lugens (71). A foliar spray of 2 % neem leaf extract reduced the overall population of BPH by 40 % (72). BPHs' longevity and survival were markedly decreased when the rice crop was sprayed with neem oil 3 % (73). BPHs' oviposition and survival rate were reduced when the crop was treated with petroleum ether-based extract of neem seed at 5 % (Fig. 1) (74).

White-backed plant hopper

The White-backed plant hopper (WBPH) is also a critical sucking pest in rice. It commonly feeds on the phloem tissues and causes a reduction in leaf area, plant height, dry weight, chlorophyll contents, photosynthetic rate and yield (75). Both adults and nymphs suck the plant sap and inject their toxic saliva into the plant, which results in the drying of leaves and hopper burn symptoms (58). The yield loss due to WBPH ranges from 11 to 39 percent in rice (76). Neem oil (1 %), mahua oil (1 %), neem seed kernel extract (5 %) and lemongrass leaf extract (5 %) affected the biological activities of WPBH, which resulted in reduced growth and development, decreased survival rates and lower adult emergence(77). Ethanol extracts of Anredera cordifolia leave at 1, 1.5 and 2 % caused nymphal and adult mortality @ 52.50 %, 56.25 % and 61.25 %, respectively, after 14 days of application. Multi-neem (300ppm) applied @ 1.25 L/ha was the most effective in managing WBPH in rice (68).

The application of NSKE (5%), neem oil (3%), neem leaf extract (3%), vitex leaf extract (3%), periwinkle leaf extract (3%), palmarosa oil (0.05%) and jatropha oil (1%) resulted in higher nymphal mortality, lower adult survival and reduced honeydew excretions (78). Spraying neem

kernel extract 5 % along with teepol (0.16 %) reduced the incidence of WBPH (79). Spraying of neem oil (1 %) @ $7.5\,L/$ ha and root soaking of rice seedlings with NSKE (5 %) decreased the incidence of WBPH (80). Similarly, research indicates that applying 5 % neem cake extract reduced the emergence of WBPH (81). The relative effectiveness of aqueous custard apple leaf extracts exhibited the reduced population in rice (9). The efficacy of vitex, Pongamia and calotropis leaf extract @ 10 % was highly effective against WBPH in rice (77).

Green leaf hopper

The green leafhoppers (GLH), particularly the Nephotettix virescenes and Nephotettix nigropictus, are common in any rice production system. In addition to direct feeding damage, they also served as vectors for viral diseases like ricetungro leaf yellowing viruses, resulting in reduced grain yield (81).A decreased nymphal survival rate and adult emergence due to applying Pongamia leaf extract @ 10 percent (50). Reduced growth development of GLH was observed when theyfed on rice plants treated with commercial neem formulations (82). Significant decreases in GLH populations were achieved by applying 8 % NSKE (83). Neem oil reduced Nigropictus virescens populations and tungro virus transmission capabilities, resulting in stable grain yield (70). Spraying custard apple leaf extract also reduced the spread of tungro viral disease in rice (84). The combination of neem oil (1%) and pungam oil (1%) resulted in 50 % reduction in GLH population over control in the organic rice experiment (24).

Ear head bug

The rice ear head bug, *Leptocorisa acuta*, is one of the most serious and harmful sucking insect pests during the maturity stages of the rice crop. Damages occur primarily during the flowering stage, where both nymphs and adults suck the sap from rice grains, resulting in shrivelled and chaffy grains. The feeding sites of ear head bugs promote the growth of sooty mould fungus, causing significant yield losses of up to 30 % (85). Direct feeding damage to ear head bugs reduces grain yield by up to 40 % in severe cases (86).

The extracts from Piperaceae plants showed a quick knockdown effect in addition to their repellent properties against rice ear head bugs in rice (87). Chloroform extract of *Pipercrocatum* was highly effective against rice ear bugs (88). Application of neem oil 3 % and NSKE 5 % showed a very low incidence of *Leptocorisa* spp in rice (71). Neem oil 2 % and neem seed kernel extract 5 % reduced *Leptocorisa* acuta to 69 and 39 % in rice (89) (Table 2). Foliar spraying of neem oil (3 %) at 15, 30 and 60 days after transplanting showed minimum black bug (*Scotinopharalurida*) incidence (6.60 %) and more grain yield (5279 kg/ha) in rice (90).

Impact of Botanicals on Natural Enemies

Natural enemies are essential components of integrated pest management in organic farming (91). Applying botanical pesticides on rice crops showed a synergistic effect and indirectly benefitted the natural enemies by

Table 2. Botanicals for the management of sucking pests in rice

S. No.	Botanical plant	Plant part	Mode of action	
Plant ar	nd leaf hoppers			
1	Neem (Azadirachta indica)	Seeds, leaves and neem oil 0.5-2.0 % azadirachtin	Antifeedant, growth inhibitor, oviposition deterrent and insecticidal activity	
2	Barnyard grass (Echinochloa crusgalli)	Transaconitic acid isolated from E.crusgalli	Antifeedant activity	
3	Morchand (Eclipta alba)	Root and shoot extracts	Antifeedant activity	
4	Cleome (Gynandropsis pentaphylla)	Petroleum seed extract	Reduced oviposition	
5	Lantana (<i>Lantana camara</i>)	Flower extract in water	Toxicity, when tested as a topical application	
6	Marigold (Tagetespafula)	Aqueous root extract	Toxicity	
7	Ginger (Zingiber officinale)	Rhizome extract	Insecticidal	
8	Basil (Ocimum sanctum)	Plant extract	Repellent	
9	Mugwort (Artememisla kurramensis)	Seed oil alone or in combination with organic materials	Insecticidal activity	
10	Polang/und (Calophylluminophyllum)	1 %seed oil of Indian lurvel	Check population and RTV transmission	
11	Amalghota/ croton (Croton sparsiflorum)	Seed oil	Reduced survival and longevity	
12	Mahua (Madhucaindica)	Seed oil	Reduced survival of insects	
13	Eucalyptus (<i>Eucalyptus</i> sp.)	Eucalyptus oil	Insect repellent	
14	Citronella (<i>Cymbopogon</i> sp.)	Citronella oil	Repelling pests	
15	Lemongrass (Cymbopogon citratus)	Essential oil Repelling pests		
16	Custard apple (Annona squamosa)	Seed oil	Effectively reduce survival of adults	
Ear hea	d bug			
1	Neem (<i>Azadirachta indica</i>)	Seeds and leaves - Extracts contain Azadirachtin, salannin and Nimbin 10 Insect growth regulator; antifeedant, disrupts mo and reproduction		
2	Pyrethrum (Chrysanthemum cinerariifolium)	Flowers - Pyrethrins 1 %	Neurotoxin to insects, causing paralysis and death	
3	Garlic (Allium sativum)	Cloves - Allicin, diallyl disulfide Repellent, antifeedant and toxic to insects		
4	Eucalyptus (<i>Eucalyptus</i> sp.)	Leaves - Eucalyptol, cineole Fumigant, repellent, insecticidal activity		
5	Turmeric (<i>Curcuma longa</i>)	Rhizomes and Curcumin Repellent, antifeedant, affects insect growth and development		
6	Tobacco (Nicotiana tabacum)	Leaves - Nicotine	Neurotoxin affects the nervous system of insects	

weakening antagonistic pests (49). The application of Azadirachtin does not affect the coccinellid predators in the rice ecosystem. Neem oil 50 EC spray did not impact the predatory green lacewing bugs (Chrysoperla carnea) in rice (IRRI1992) (92). The Application of Azadirachtin to manage rice pests showed less impact on the population reduction of parasitoid predators in the rice ecosystem (93). Neem oil and various non-edible oil sprays were considered safer for beneficial insects (94). Neemax, Rakshak and Fortune Aza were lesser risk to the predators of plant hoppers such as the velid bugs (Microvelia douglasiatrolineata), mirid bugs and Trichogramma japonicum egg parasitoids (95). A higher spider population of 1.30-1.70 per hill was observed at 3 Days after treatment (DAT) and 1.50-2.00 per hill at 7 DAT was recorded in neem oil (1 %) + pungam oil (1 %) as foliar spray in organic rice (91). The use of botanicals is one of the essential components in organic plant protection strategy, is safe for biological control agents and saves the environment from pesticide pollution (96).

Mode of Action of Botanicals on Rice Insects

Understanding the mode of action of botanicals through

physical, biological and chemical interactions between the insect and the product is crucial for managing pests successfully (13). The mode of action of botanicals involves repulsiveness, inhibition, dehydration of proteins, etc., depending on the botanical ingredient and nature of the insect pests. Neem-based pesticides have antifeedant and repellent properties. Additionally, moulting aberrant, oviposition deterrent and endocrine system disruption. Pyrethrum-based bio-pesticides damaged the insect nerve cells, followed by paralysis and death (14). A combination of botanicals was more effective, significantly reducing enzyme activity, including acetylcholinesterase (AChE), glutathione S-transferase (GST), and cytochrome P450 monooxygenases. The synergistic effect of botanicals was observed when combined with bacterial toxins in disrupting the digestive processes of most rice pests, resulting in impaired nutrition and eventual mortality (49).

Neem exhibits a broad range of activities against most rice pests, acting as a feeding deterrent, repellent, growth regulator and oviposition inhibitor (96). Apart from neem formulations, the solvent-free neem formulation is also effective against sucking pests (96). Pyrethrin interfer-

ence in the sodium and potassium ion exchange mechanism within insect nerve fibres was noticed, thereby disrupting nerve impulse transmission. Nevertheless, due to the rapid metabolization property of pyrethrins, many insects experienced paralysis and death (13). In agricultural pest management, using plant-based products and identifying toxic principles in the plant parts are imminent research areas to focus (14). Alkaloids derived from *sabadilla* were toxic to insect pests, disrupting nerve cell membranes and leading to loss of nerve function, paralysis and death (11). Findings on the mode of action of plant-derived compounds have led to the identification of novel compounds with unique modes of action to combat pest re-

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Authors' contributions

KG and BA wrote the first draft of the paper. KG and RPS conceptualized, reviewed, and edited the review paper holistically. MS, VM, SV, SVS, and MM reviewed the paper and shared their input for upscaling. All authors read and approved the final manuscript.

Compliance with ethical standards

Table 3. Mode of action of botanicals on rice insect pests

S. No	Plant	Compound	Mechanism of action	Impact of insect system
1	Mentha sp. Lavendula sp.	Essential oils	Inhibition of acetylcholinesterase (AChE)	Cholinergic system
2	Nicotiana sp.Haloxylonsalicornicum, Stemona Jap	Nicotine	Cholinergic acetylcholine nicotinic receptor Agonist/ antagonist	Cholinergic system
3	Chrysanthemum cinerariaefolium	Pyrethrin	Sodium and potassium ion exchange disruption	Mitochondrial system
4	Lonchocarpus sp.	Rotenone	Inhibitor of cellular respiration(mitochondrial complex I electron transport inhibitor (METI)	Mitochondrial system
5	Ryania sp.	Ryanodine	Affect calcium channels	Mitochondrial system
6	Cedrus sp. Eucalyptus sp.	Essential oils	Octopaminergic receptors	Octopaminergic system
7	Thymus vulgaris	Thymol	Block octopamine receptors by working through tyramine receptors cascade	Octopaminergic system
8	Schoenocaulon officinale	Sabadilla	Affect nerve cell membrane action	Mitochondrial system
9	Nicotiana tabacum	Nicotine	Mimics acetylcholine, causing overstimulation of the nervous system	Nervous system
10	Azadiractina indica	Azadirachtin	Hormonal balance disruption	Hormonal system
11	Allium sativum	Allicin	Antifeedant and repellent; also has fungicidal and bactericidal effects	Digestive system

sistance and resurgence (12) (Table 3).

Conclusion

Botanicals represent a viable and sustainable component of integrated pest management in rice. Future research should focus on identifying novel plant-derived compounds, optimizing formulation techniques and conducting long-term field studies to evaluate the efficacy of botanicals within IPM frameworks. Botanicals offer a promising avenue for enhancing rice yields through natural and sustainable pest management methods. Integrating botanicals into rice cultivation can improve crop growth and yield while increasing resilience to environmental stresses. Future research should focus on exploring the potential of botanical compounds, optimizing application techniques and understanding the unique mode of action for better pest control efficiency. Development of guidelines for the use of botanicals can further promote their adoption, ensuring food security and environmental health. Botanical pesticides may play a significant role in food production and postharvest protection in developing countries like India. Hence, there is tremendous potential in the future for business start-ups to develop new botanical formulations for eco-friendly pest management.

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References

- Agarrwal R, Bentur JS, Nair S. Gas chromatography mass spectrometry-based metabolic profiling reveals biomarkers involved in rice-gall midge interactions. J Integr Plant Biol. 2014;56:837–48. https://doi.org/10.1111/jipb.12244
- Alice J, Sujeetha R. The biological and behavioral impact of some indigenous plant products on rice white backed plant hopper (WBPH) Sogatella furcifera (Horvath) (Homoptera: Delphacidae). Inde. 2008;1:193–96. https://doi.org/10.57182/ jbiopestic.1.2.193-196
- Amandeep GS, Verma PK, Varma A. Evaluation of indigenous plant extracts with cow urine against rice stem borer and rice leaf folder in basmati rice. Pharma Innov J. 2021;10:354–59.
- Amoabeng BW, Gurr GM, Gitau CW, Nicol HI, Munyakazi L, Stevenson PC. Tri-trophic insecticidal effects of African plants against cabbage pests. PLoS One. 2013;8:e78651. https://doi.org/10.1371/journal.pone.0078651
- Ananthi GBG, Roy K, Chen B, Lim JB. Testing, simulation and design of back-to-back built-up cold-formed steel unequal angle sections under axial compression. Steel Compos Struct. 2019;33:595–614. https://doi.org/10.12989/scs.2019.33.4.595
- 6. Assabgui R, Lorenzetti F, Terradot L, Regnault-Roger C, Malo N,

Wiriyachitra P, et al. Efficacy of botanicals from the Meliaceae and Piperaceae. ACS Publ. 1997;54:145–56. https://doi.org/10.1021/bk-1997-0658.ch004

- Athulya R, Maheswari TU, Padmakumari A, Madhav MS, Devi GU. Reaction of rice varieties to yellow stem borer, *Scirpophaga incertulas* (Walker) populations. Integr Pest Manag Rev. 2022;3:155–60.
- Balamurugan S, Kannan R. Larvicidal and insect growth regulator activity of a brown algal seaweed, Sargassum wightii
 (Greville) against rice leaf folder. Int J Entomol Res. 2022;7:10–
 14.
- Banerjee S. In Andhra Pradesh there are three broods in rabi season. The first brood is noticed from about the end of May. Entomol India. 1964;92:1938–63.
- Bhattacharyya P, Bordoloi D. Insect growth retardant activity of some essential oil bearing plants. J Econ Entomol. 1986;90:1102 –05.
- Bhojane S, Desai V, Wade P, Jalgaonkar V, Wankhede S, Desai S. Management of major pests infesting rice (*Oryza sativa* L.) using botanicals. J Entomol Zool Stud. 2020;8:1962–64.
- 12. Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. Biological activities and medicinal properties of neem (*Azadirachta indica*). Curr Sci. 2002;11:1336–45.
- Chatterjee S, Mondal P. Management of rice yellow stem borer, Scirpophaga incertulas Walker using some biorational insecti- cides. J Biopesticides. 2014;7:143. https://doi.org/10.57182/ jbiopestic.7.0.143-147
- Cheng X, Zhu L, He G. Towards understanding of molecular interactions between rice and the brown planthopper. Mol Plant. 2013;6:621–34. https://doi.org/10.1093/mp/sst030
- Dash A, Senapati B, Tripathy M. Effect of neem derivatives alone and in combination with synthetic insecticides on gall midge and its parasitization. J Pest Sci. 1994;85:359–66.
- Dash S, Mohapatra L, Swain S, Swain D. Bio-efficacy of newer insecticides in combination with neem product against plant hoppers of rice. J Entomol Zool Stud. 2019;7:1152–55.
- 17. Dhaliwal G, Jindal V, Dhawan A. Insect pest problems and crop losses: changing trends. Indian J Ecol. 2010;37:1–7.
- Dhaliwal GS, Multani JS, Singh S, Kaur G, Dilawari VK, Singh J. Field evaluation of azadirachtin-rich neem formulations against Cnaphalocrocis medinalis (Guenee) and Scirpophaga incertulas (Walker) on rice. Pesticide Res J. 2002;14:69–76.
- Dodia D, Patel I, Patel G. Botanical pesticides for pest management. Pest Manag Sci. 2010;77:3500-07.
- 20. Durairaj C, Venugopal M. Effects of neem and nochi on rice bug *Leptocorisa acuta*. Aust J Bot. 1993;53:607–19.
- Febritami G, Usyati N, Dono D. Toxicity of four plant extracts (Ageratum conyzoides L., Barringtonia asiatica L., Melia azedarach L., Tephrosia vogelii L.) against brown planthopper (Nilaparvata lugens). Cropsaver J Plant Prot. 2018;1:1–8. https://doi.org/10.24198/cs.v1i1.16970
- Ganesan K, Somasundaram E, Sanbagavalli S, Kumar P. Ecofriendly management practices for insect pests in organic farming. In: Kempuraj T, Balasubramanian R, Philip H, Selvaraj KN, editors. Ecofriendly management technologies for pests and diseases; 2017. 117–27.
- Ganesan K, Somasundaram E. Ecofriendly pesticides for the management of insect pests in organic rice. Int J Agric Sci. 2022;14:11408–10.

Ganesan K, Soundararajan RP, Sanbagavalli S. Indigenous technical knowledge (ITK) in pest management. In: Kennedy JS, Ganesan K, Senguttuvan K, Rajabaskar D, Kuttalam S, editors. Functional insect pest management. Coimbatore: A.E. Publications; 2014. p. 191–98.

- 25. Golec AFC. Effect of neem (*Azadirachta indica* A. Juss) insecticides on parasitoids. Rev Peru Biol. 2007;14:69–74.
- Gunasena H. Indigenous vegetables and legumes in Sri Lanka.
 Acta Hortic. 2007;752: 111–14. https://doi.org/10.17660/actahortic.2007.752.14
- Hajjar MJ, Ahmed N, Alhudaib KA, Ullah H. Integrated insect pest management techniques for rice. Sustainability. 2023;15:4499. https://doi.org/10.3390/su15054499
- Islam MS, Das SD, Islam KS, Rahman AR, Huda MN, Dash PK. Evaluation of different insecticides and botanical extracts against yellow stem borer, *Scirpophaga incertulas* in rice field. Int J Bio Sci. 2013;9:2731. https://doi.org/10.12692/ijb/3.10.117-125
- Isman MB. Botanical insecticides, deterrents and repellents in modern agriculture and an increasingly regulated world. Annu Rev Entomol. 2006;51:45–66.https://doi.org/10.1146/ annurev.ento.51.110104.151146
- 30. Jan U. Evaluation of botanicals against grasshopper, *Oxyanitidula Walker* infesting rice, *Oryza sativa* L. in Kashmir. Cropsaver J Plant Prot. 2019;25:112–15.
- January B, Rwegasira GM, Tefera T. Efficacy of selected biopesticides and botanical extracts in managing rice stem borer, *Chilo partellus* (Lepidoptera: Crambidae) in Tanzania. J Econ Entomol. 2018;93:732–33. https://doi.org/10.9734/jaeri/2018/44015
- Kardinan A, Wahyono TE, Tarigan N. Persistence of botanical insecticide residue of Pyrethrum and neem in rice plant. J Econ Entomol. 2017;96:1054–65. https://doi.org/10.21082/bullittro.v28n2.2017.191-198
- Khan BV, Harrison DG, Olbrych MT, Alexander RW, Medford RM. Nitric oxide regulates vascular cell adhesion molecule 1 gene expression and redox-sensitive transcriptional events in human vascular endothelial cells. Proc Natl Acad Sci USA. 1996;93:9114 –19. https://doi.org/10.1073/pnas.93.17.9114
- 34. Krishnaiah N, Kalode M. Efficacy of selected botanicals against rice insect pests under field conditions. Indian J Plant Prot. 1990;18:197–205.
- Krishnaiah N, Kalode M. Evaluation of neem oil, neem cake and other non-edible oil cakes against rice pests. J Appl Entomol. 1984;121:121–28.
- 36. Ladja FT, Santoso T, Nurhayati E. Potensi cendawan entomopatogen *Verticillium lecanii* dan *Beauveria bassiana* dalam mengendalikan wereng hijau dan menekan intensitas penyakit tungro. J Penelitian Pertanian Tanaman Pangan. 2015;30:139–66.
- 37. Lakshmi VJ, Katti G, Krishnaiah N. Safety of neem formulations and insecticides to *Microvelia douglasi atrolineata* Bergroth (Heteroptera: Veliidae), a predator of planthoppers in rice ecosystem. J Biol Control. 1997;15:33–36.
- Laskowski W, Gorska-Warsewicz H, Rejman K, Czeczotko M, Zwolinska J. How important are cereals and cereal products in the average Polish diet. Nutrients. 2019;11:679. https:// doi.org/10.3390/nu11030679
- Litsinger J, Alviola A, Cruz DC, Canapi B, Batay-An III E, Barrion A. Rice white stemborer *Scirpophaga innotata* (Walker) in southern Mindanao, Philippines. I. Supplantation of yellow stemborer *S. incertulas* (Walker) and pest status. Int J Pest Manag. 2006;52:11

 –21. https://doi.org/10.1080/09670870600552497
- Mahalingam L, Reed D. Bipolar device packaging. Electrical, thermal and mechanical stress considerations. Microelectronics J. 1987;18:65. https://doi.org/10.1016/s0026-2692(87)80465-2

- 41. Mariappan V, Jayaraj S, Saxena R. Effect of non-edible seed oils on survival of *Nephotettix virescens* (Homoptera: Cicadellidae) and on transmission of rice tungro virus. J Econ Entomol. 1988;81:1369–72. https://doi.org/10.1093/jee/81.5.1369
- 42. Mayabini Jena MJ. Integrated pest management with botanical pesticides in rice with emphasis on neem products. Fitoterapia. 2005;78:205–10.
- 43. Mohan K, Gopalan M, Balasubramanian G. Studies on the effects of neem products and monocrotophos against major pests of rice and their safety to natural enemies. Indian J Plant Prot. 1991;19:23–30.
- 44. Mohapatra P, Ponnurasan N, Narayanasamy P. Tribal pest control practices of Tamil Nadu for sustainable agriculture. Phytoparasitica. 2009;32:433–43.
- Muralidharan K, Pasalu I. Assessments of crop losses in rice ecosystems due to stem borer damage (Lepidoptera: Pyralidae).
 Crop Prot. 2006;25(5):409–17. https://doi.org/10.1016/j.cropro.2005.06.007
- Narasimhan V, Mariappan V. Effect of plant derivatives on green leafhopper (GLH) and rice tungro (RTV) transmission. J Appl Zool Res. 1988;62:1381–87.
- Nathan SS, Choi MY, Paik CH, Seo HY. Food consumption, utilization and detoxification enzyme activity of the rice leaf folder larvae after treatment with *Dysoxylum* triterpenes. Pestic Biochem Physiol. 2007;88:260–67. https://doi.org/10.1016/j.pestbp.2007.09.002
- Nathan SS, Chung PG, Murugan K. Combined effect of biopesticides on the digestive enzymatic profiles of *Cnaphalocrocis medinalis* (Guenee) (the rice leaf folder) (Insecta: Lepidoptera: Pyralidae). Ecotoxicol Environ Saf. 2006;64:382–89. https://doi.org/10.1016/j.ecoenv.2005.04.008
- Nathan SS, Kalaivani K, Murugan K, Chung PG. Efficacy of neem limonoids on *Cnaphalocrocis medinalis* (Guenée) (Lepidoptera: Pyralidae) the rice leaffolder. Crop Prot. 2005;24:760–63. https://doi.org/10.1016/j.cropro.2005.01.009
- 50. Norton GW, Heong K, Johnson D, Savary S. Rice pest management: issues and opportunities. Rice in the global economy: strategic research and policy issues for food security. Los Banos: IRRI; 2010. p. 433–43.
- 51. Nugroho LH, Pratiwi R, Soesilohadi R, Subin ER, Wahyuni S, Hartini YS, Lailaty IQ. Repellent activity of *Piper* spp. leaves extracts on rice ear bugs (*Leptocorisa oratorius*) and the characters of its volatile compounds. Annu Res Rev Biol. 2020;35:34–45. https://doi.org/10.9734/arrb/2020/v35i930269
- 52. Nuryanti N, Martono E, Ratna E, Dadang D. The bioactivities of selected Piperaceae and Asteraceae plant extracts against brown planthopper (*Nilaparvata lugens* Stal). Appl Entomol Zool. 2018;11:563–70.
- 53. Ogah E, Ogbodo E. Comparative efficacy of neem seed extract with carbofuran in the management of African rice gall midge, *Orseolia oryzivora* Harris and Gagne (Diptera: Cecidomyidae). J Biol Agric Healthcare. 2012;2:147–53.
- 54. Ogah E, Omoloye A, Nwilene F, Nwogbaga A. Effect of neem seed kernel extracts in the management of rice stem borers in the field in Nigeria. Niger J Biotechnol. 2011;23:114–17.
- Powell K, Gatehouse A, Hilder V, Van Damme E, Peumans W, Boonjawat J, et al. Different antimetabolic effects of related lectins towards nymphal stages of *Nilaparvata lugens*. Entomol Exp Appl. 1995;75:61–65. https://doi.org/10.1111/j.1570-7458.1995.tb01910.x
- Prakash A, Rao J, Nandagopal V. Future of botanical pesticides in rice, wheat, pulses and vegetables pest management. J Biopesticides. 2008;1:154–69. https://doi.org/10.57182/ jbiopestic.1.2.154-169
- 57. Prakash A, Tiwari S, Rao J. Exploitation of natural plant products

- for management of pests and diseases in rice ecosystems. Proc Symp Growth Dev Resource Conserv. 1990;73:274–77.
- 58. Pustika AB, Kobarsih M. Plant density and water management effect to leaf folder and narrow brown spot severity in irrigated rice. E3S Web Conf. 2021;56:651–76. https://doi.org/10.1051/e3sconf/202130601014
- 59. Raguraman S, Rajasekaran B. Effect of neem products on insect pests of rice and the predatory spider. Madras Agri J. 1996;86:117–25. https://doi.org/10.29321/maj.10.a01046
- Rajappan K, Ushamalini C, Subramanian N, Narasimhan V, Abdul Kareem A. Effect of botanicals on the population dynamics of *Nephotettix virescens*, rice tungro disease incidence and yield of rice. Phytoparasitica. 2000;28:109–13. https://doi.org/10.1007/bf02981739
- 61. Rajasekaran B, Jayraj S, Raghuraman S, Narayanswamy T. Use of neem products for the management of certain rice pests and diseases. J Essential Oil Res. 1987;18(3):315–17.
- Ramaraju K, Sundarababu P. Effect of plant derivatives on brown planthopper, white-backed planthopper, green leafhopper of rice. J Food Sci Technol. 1990;27:296–300.
- 63. Rao J, Prakash A, Ghosh S, Sinha R. Deterioration of seed quality by tarsonemid mites infestation in rice fields. Oryza. 1998;35:297 –99.
- 64. Rao PRM, Rao PS. Effect of biocides on brown planthopper adults on rice. Int Rice Res Newsl. 1979;4:20.
- Rashid M, Ahmed N, Jahan M, Islam K, Nansen C, Willers J, Ali M. Higher fertilizer inputs increase fitness traits of brown planthopper in rice. Sci Rep. 2017;7:4719. https://doi.org/10.1038/s41598 -017-05023-7
- Ravisankar N, Raja R, Din M, Elanchezhian R, Chaudhuri SG. Evaluation of green manure intercropping in wet seeded and transplanted rice under island ecosystem. Oryza. 2007;44:231– 33.
- 67. Reddy AV, Devi RS, Reddy DVV. Evaluation of botanical and other extracts against plant hoppers in rice. J Biopesticides. 2012;5:57. https://doi.org/10.57182/jbiopestic.5.1.57-61
- Relyea RA. New effects of roundup on amphibians: Predators reduce herbicide mortality; herbicides induce antipredator morphology. Ecol Appl. 2012;22:634–47. https://doi.org/10.1890/11-0189.1
- Roy P, Uddin MM, Islam KS, Das KR. Efficacy of different botanical and chemical insecticides against rice hispa (*Dicladispa armigera*). Prog Agric. 2017;28:64–72. https://doi.org/10.3329/pa.v28i2.33466
- Rubia-Sanchez E, Suzuki Y, Arimura K, Miyamoto K, Matsumura M, Watanabe T. Comparing Nilaparvata lugens and Sogatella furcifera (Horvath) (Homoptera: Delphacidae) feeding effects on rice plant growth processes at the vegetative stage. Crop Prot. 2003;22:967–74. https://doi.org/10.1016/s0261-2194(03)00112-1
- 71. Sachan S, Singh D, Chaudhary A. Field evaluation of insecticides against rice stem borer and leaf folder. Ann Plant Prot Sci. 2006:14:469–70.
- 72. Saikia P, Parameswaran S. Eco-friendly strategies for the management of rice leaffolder, *Cnaphalocrocis medinalis* Guenee. Ann Pl Prot Sci. 2002;10(1):12–16.
- 73. Samalo A, Senapati B, Satpathy C, Jacob T. Effect of neem derivatives on the incidence of some major insect pests in wet season rice. Green Pesticides Handbook. 1993;256:2166–74.
- 74. Samrit R, Chaudhari B, Chopkar P, Uparkar SSA. Evaluation of bio-pesticides, botanicals and plant extracts against brown plant hopper (*Nilaparvata lugens*) of rice. Appl Entomol Zool. 2020;3:155–60.

 Sangamithra S, Vinothkumar B, Manoharan T, Muthukrishnan N, Rathish S. Evaluation of bioefficacy, phytotoxicity of imidacloprid 17.1% SL against plant and leaf hoppers and its safety to non-target invertebrates in rice. J Entomol Zool Stud. 2018;6:230 –34. https://doi.org/10.20546/ijcmas.2018.701.397

- Savary S, Willocquet L, Elazegui FA, Castilla NP, Teng PS. Rice pest constraints in tropical Asia: quantification of yield losses due to rice pests in a range of production situations. Plant Dis. 2000;84:357–69. https://doi.org/10.1094/pdis.2000.84.3.357
- Saxena R, Justo H, Rueda B. Neem seed bitters for management of planthopper and leafhopper pests of rice. Biopesticides Int. 1987;4:63–84.
- Saxena R, Justo Jr H, Epino P. Evaluation and utilization of neem cake against the rice brown planthopper, *Nilaparvata lugens* (Homoptera: Delphacidae). J Econ Entomol. 1984;77:502–07. https://doi.org/10.1093/jee/77.2.502
- Saxena R, Khan Z. Effect of neem oil on survival of Nilaparvata lugens (Homoptera: Delphacidae) and on grassy stunt and ragged stunt virus transmission. J Econ Entomol. 1985;78:647–51. https://doi.org/10.1093/jee/78.3.647
- Schmutterer H, Saxena R, Heyde VDJ. Morphogenetic effects of some partially purified fractions and methanolic extracts of neem seeds on *Mythimna separata* (Walker) and *Cnaphalocrocis medinalis* (Guenee). Z Angew Entomol. 1983;95:230–37. https:// doi.org/10.1111/j.1439-0418.1983.tb02637.x
- 81. Seni A, Naik B. Influence of different abiotic factors on the incidence of major insect pests of rice (*Oryza sativa* L.). J Agromete-orol. 2018;20:256–58. https://doi.org/10.54386/jam.v20i3.559
- 82. Seni A, Naik BS. Evaluation of some insecticides against brown plant hopper, *Nilaparvata lugens* (Stal) in rice, *Oryza sativa* L. Int J Bio-resource Stress Manag. 2017;8:268–71. https://doi.org/10.23910/ijbsm/2017.8.2.1685
- 83. Seni A. Impact of certain essential oils and insecticides against major insect pests and natural enemies in rice. Wheat Barley Res. 2019;11:252–56. https://doi.org/10.25174/2249-4065/2019/95533
- 84. Shah FM, Razaq M, Ali Q, Ali A, Shad SA, Aslam M, Hardy IC. Action threshold development in cabbage pest management using synthetic and botanical insecticides. Entomol Generalis. 2020;40:325–29.
 - https://doi.org/10.1127/entomologia/2020/0904
- 85. Simmonds M, Manlove J, Blaney W, Khambay B. Effect of botanical insecticides on the foraging and feeding behavior of the coccinellid predator *Cryptolaemus montrouzieri*. Phytoparasitica. 2000;28:99–107. https://doi.org/10.1007/bf02981738

- 86. Singh R, Srivastava R. Prospects of neem (*Azadirachta indica* A. Juss.) in the management of insect pests of stored grain in developing countries. J Eco-friendly Agric. 2015;10:1–14.
- 87. Sison P. Some notes on the white pyralid moth borer (*Scirpophaga innotata* Walk.) and suggestions for its control. Philippine Agric Rev. 1929;22:333–43.
- 88. Sogawa K. The rice brown planthopper: feeding physiology and host plant interactions. Annu Rev Entomol. 1982;27:49–73. https://doi.org/10.1146/annurev.en.27.010182.000405
- 89. Soti A, Bhandari S, Acharya A, Shrestha S. Screening of early maturing rice varieties against rice earhead bug, *Leptocorisa oratorius* Fabricius (Hemiptera: Alydidae) in IRD Center Jhapa, Nepal. J Entomol Zool Stud. 2020;8:1363–66.
- Soundararajan RP, Chandrasekaran M, Chitra N. Strategies for organic insect pest management in pulses. In: Dutta P, Chakraborty A, editors. Current trends in plant health management. Tripura: Biotica Publishers; 2021. p.1–17.
- 91. Soundararajan RP, Chitra N. Effect of global warming in agriculture and insect pest. In: Singh KK, Aleya L, Singh V, Singh M, editors. Manmade disasters. New Delhi: APH Publishing Corporation; 2010. p. 205–35.
- 92. Soundararajan RP, Lakshmanan V. Neem in pest management of horticultural crops. In: Singh KK, Phogat S, Tomar A, Dhillon RS, editors. Neem A treatise. New Delhi: I.K. International Publishing House Pvt. Ltd.; 2009. p. 396–413.
- Tiwari S, Bisht D, Srivastava A, Pipal A, Taneja A, Srivastava M, Attri S. Variability in atmospheric particulates and meteorological effects on their mass concentrations over Delhi, India. Atmos Res. 2014;145:45–56. https://doi.org/10.1016/j.atmosres.2014.03.027
- 94. Venugopal PS, Elamathi S, Subrahmanian K, Sassikumar K, Elanchezhyan K, Anandhi. Ecological engineering methods for the management of rice black bug, *Scotinophara lurida* Burmeister (Pentatomidae: Hemiptera): an emerging pest in rice. Ecol Eng. 2023;54:2.
- 95. Watanabe T, Kitagawa H. Photosynthesis and translocation of assimilates in rice plants following phloem feeding by the planthopper *Nilaparvata lugens* (Homoptera: Delphacidae). J Econ Entomol. 2000;93:1192–98. https://doi.org/10.1603/0022-0493-93.4.1192
- Yi J, Qiu M, Liu N, Tian L, Zhu X, Decker EA, McClements DJ. Inhibition of lipid and protein oxidation in whey-protein-stabilized emulsions using a natural antioxidant: Black rice anthocyanins.
 J Agric Food Chem. 2020;68:10149–56. https://doi.org/10.1021/acs.jafc.0c03978