

RESEARCH ARTICLE



Inheritance of fertility restoration in A4 cytoplasm of Pigeonpea (*Cajanus cajanifolius* L.)

Thanga Hemavathy Arumugam ^{1*}, Kavitha Shanmugam ², Sakila Muthusamy ³, Vijaya Geetha Venugopal⁴, M. Kanimoli Mathivathana⁵ & Santhi Madhavan Samyuktha⁶

¹Department of Pulses, Tamil Nadu Agricultural University, Coimbatore 641 003, Tamil Nadu, India

²Tamil Nadu Agricultural University, Coimbatore 641 003, Tamil Nadu, India

³Krishi Vigyan Kendra (KVK), Sirugamani 639 115, Tamil Nadu, India

⁴Krishi Vigyan Kendra (KVK), Tamil Nadu Agricultural University, Tindivanam 604 002, Tamil Nadu, India

⁵College of Agricultural Technology, Theni 625 562, Tamil Nadu, India

⁶Dhanalakshmi Srinivasan Agriculture College, Perambalur 621 212, Tamil Nadu, India

*Email: hemavathytnau@gmail.com

ARTICLE HISTORY

Received: 29 September 2024 Accepted: 29 December 2024 Available online Version 1.0:07 February 2025 Version 2.0:13 February 2025

Check for updates

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonepublishing.com/ journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc See https://horizonepublishing.com/journals/ index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an openaccess article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (https://creativecommons.org/licenses/ by/4.0/)

CITE THIS ARTICLE

Arumugam TH, Shanmugam K, Muthusamy S, Venugopal VG, Mathivathana MK, Samyuktha SM. Inheritance of fertility restoration in A4 cytoplasm of Pigeonpea (*Cajanus cajanifolius* L.). Plant Science Today. 2025; 12(1): 1-4. https:// doi.org/10.14719/pst.6120

Abstract

A set of three medium-duration CMS lines ICPA 2043, ICPA 2047 and ICPA 2048 were used in study to investigate fertility restoration by making twenty-eight test crosses at the Department of Pulses, TNAU, Coimbatore. ICPA 2047 exhibited fertility restoration ranging from 0-40 % with negligible pod setting except for the cross ICPA 2047 × ICP 3181. In contrast, sixteen F1s derived from ICPA 2048 showed pollen fertility ranging from 10-48 % except for the cross ICPA 2048 × ICP 9224. Similarly, sixteen F1s derived from ICPA 2043 demonstrated 15-38 % pollen fertility, with ICPA 2043 × LRG 41 showing 98 % fertility and good pod setting. Among the hybrids, ICPA 2043 × LRG 41 exhibited high fertility, ranging from 95.58 % to 98.67 % over two years. This suggests that dominant fertility-restoring genes from the restorer parent were incorporated into the hybrid, which showed consistent fertility restoration across different years. In the current investigation, the restorer line ICP 3181 demonstrated monogenic inheritance of fertility restoration in crosses with ICPA 2047 and ICPA 2048, following 3:1 ratio. Conversely, the cross ICPA 2043 × LRG 41 exhibited a digenic duplicate dominance inheritance pattern. These results suggest that the fertility restoration in pigeonpea can be inherited monogenically or digenically, depending on the hybrid. The identified hybrids, particularly ICPA 2043 × LRG 41 demonstrate significant potential for improving fertility restoration and enhancing productivity in pigeonpea breeding programs.

Keywords

A4 cytoplasm; fertility restoration; inheritance; male sterile lines; pigeonpea

Introduction

To address the stagnant productivity of the global pigeonpea crop, caused by various biotic and abiotic factors as well as a lack of high-yielding varieties over the past three decades, heterosis breeding has been employed in pigeonpea breeding research over 60 years ago. Increasing pigeonpea yield has been the primary goal of the national breeding efforts (1). The only way to achieve yield maximisation in this frequently cross-pollinated exception (due to insect pollination) among the grain legumes of the world was assumed to be through the exploitation of hybrid vigour. Any crop must have stable male sterile lines as the female parents and stable restores as the pollen parents in order to generate good-yielding hybrids. Fertility restorer (rf) genes suppress male sterility, enabling the use of CMS systems for hybrid

seed production. In pigeonpea, there are several cytoplasmic nuclear male sterility (CMS) systems.

Despite the introduction of several hybrid varieties, farmers' fields do not experience significant yield improvements. This is due to factors such as partial fertility restoration and high genotype-environment interactions. Most morphological characteristics of C. cajanifolius A4 cytoplasm are similar to those of cultivated varieties (2). According to reports, the CMS lines that contain A4 cytoplasm are extremely stable over a wide range of conditions and periods without displaying any morphological abnormalities. The current study presents a comparative evaluation of fertility restoration in hybrids containing A4 cytoplasm and discusses the importance of these findings for pigeonpea hybrid breeding. From C. cajanifolius, three CMS lines namely, ICPA 2047, ICPA 2048 and ICPA 2043 were utilised. Even though these CMS lines were recorded, differences were found in pollen fertility restoration, which may be the result of different intergenomic and cytoplasmic genome interactions between the cultivars that were converted to the A-line and the wild source cytoplasm (3). Understanding restorer genetics is crucial for developing new restorers for available CMS lines. Consequently, the goal of the current inquiry is to learn how CMS lines descended from A 4 origins.

Materials and Methods

A set of three CMS lines [1] ICPA 2043, ICPA 2047 and ICPA 2048, were used for the current study. To attain the higher yield in the A4 cytoplasm the medium maturity group for fertility restoration with proper stability these CMS lines were used for the current study. These CMS lines were used to create 28 test crosses. At the Department of Pulses, TNAU, Coimbatore, 28 test crosses using germplasm lines were made during *kharif* 2016 (Table 1). F1 seeds were separately gathered and produced during the 2017 *kharif* season. All of the CMS lines and testers were planted inside the nylon net at the Department of Pulses, CPBG, TNAU, Coimbatore in order to shield the experimental materials from pollinating insects. In order to prevent pollen shedders during hybridisation, individual CMS line plants were tested for male sterility.

At flowering, new pollen from the 28 crossings in a Line × Tester mating strategy was used to manually pollinate the malesterile lines on more than 500 occasions. With a mean crossing **Table 1.** CMS lines used for crossing in medium duration success of 60 %, 40-80 pods were collected from each cross. All the hybrid combinations were planted during kharif 2018 in an unreplicated state to examine the stability of fertility restoration. Two seeds per hill were planted with the recommended spacing of 90 × 30 cm and this produced a 90% plant stand. To raise a robust crop, conventional cultural practices were used. Five completely developed but unopened flower buds were chosen randomly from each cross of 30 plants and their anthers were crushed in a 2 % acetocarmine solution to examine the pollen fertility. Three microscopic fields [3] were examined under a light microscope to examine the pollen productivity of each plant. The pollen grains that were completely dyed were regarded fertile, whilst the pollen grains that were empty or only partially stained were thought to be male-sterile. The plants were divided into three groups according to (4) with some modifications: fertile (>90 % pollen fertility with good pod setting), partial fertile (11-89 % pollen fertility with poor pod setting) and sterile (0-10 % pollen fertility with negligible pod setting). A measure of fertility restoration was the proportion of male fertile plants in each F1 hybrid. Based on the pollen fertility status of the twenty-eight test crosses, three hybrids from ICP 3181, ICP 9224 and LRG 41 (exhibiting >80 % pollen fertility restoration and good pod setting) were chosen and their morphological traits were used as descriptors to identify the true parent as well as F₁s. In order to obtain seeds for the F₂ generation, all of the F1 plants were selfed using insect-proof nylon net cages. They were then crossed [1] to their respective A-lines to produce BC1 seeds and three new crosses were also made in 2016. A Chi-square test was used to evaluate the goodness of fit to various predicted ratios in the F₂, BC_1F_1 and test cross generations.

Results and Discussion

A stable CGMS (cytoplasm Male Sterility System) and effective restorers are crucial for developing CGMS hybrids in pigeonpea (2). The confirmation of potential fertility restorers cannot be done based on pollen fertility alone due to the stability of restorers. However, the studying pod set through self-pollination and open pollination under isolation is considered as an essential criterion for identification of restorer in pigeonpea (5). Consistent with earlier research (6) at Patencheru, test crosses on male sterile plants following hand pollination showed high pod setting success (60 %) (Table 2). This demonstrates that a

S.No.	CMS lines	Source	Plant type	Days to 50% flowering	Anther morphology	Mean pollen fertility (%)
1	ICPA 2047	A ₄	NDT	115	Yellow, scaly	0.0
2	ICPA 2048	A ₄	NDT	120	Yellow, scaly	0.0
3	ICPA 2043	A ₄	NDT	114	Yellow, scaly	0.0
			Lis	t of test crosses made from	the A4 CMS lines	
1.	ICPA 2	047 × ICP 52	5403	16	ICPA 2047×IC	P 2047
2.	ICPA 2	047 × ICP 52	5454	17	ICPA 2048 × ICF	P 525403
3.	ICPA 2	047 × ICP 54	5457	18	ICPA 2048 × ICF	° 525454
4.	ICPA	2047 × ICP 9	224	19	ICPA 2048 × ICF	P 525457
5.	ICPA 2	2047 × ICP 10	0697	20	ICPA 2048 × IC	P 9224
6.	ICPA 2	2047 × ICP 10	0788	21	ICPA 2048 × IC	P 10788
7.	ICPA 2	047 × ICP 52	5438	22	ICPA 2048 × ICF	P 525438
8.	ICPA 2	047 × ICP 52	5440	23	ICPA 2048 × ICF	9 525521
9.	ICPA 2	2047 ×ICP 52	5471	24	ICPA 2048 × IC	P 12321
10.	ICPA 2	047 × ICP 52	5421	25	ICPA 2048 × IC	P 3181
11	ICPA 2	2047 × ICP 12	2321	26	ICPA 2048 × IC	P 3666
12	ICPA	2047 × ICP 1	135	27	ICPA 2043 × ICI	PB 2043
13	ICPA	2047 ×ICP 2	224	28	ICPA 2043 × L	.RG 41
14	ICPA	2047 × ICP3	181			
15	ICPA	2047 × ICP 3	666			

 $\label{eq:table_$

S.No.	Crosses	Bud pollinated	Seeds harvestee	% I success
	F ₁ Hyb	rids		
1	ICPA 2047 / ICP 3181	94	51	54.25
2	ICPA 2048 / ICP 9224	95	57	60.00
3	ICPA 2043 / LRG 41	118	67	56.77
	Test	crosses		
4	ICPA 2047// ICPA 2047/ ICP 318	1 194	84	43.29
5	ICPA 2048/ ICPA 2048/ ICP 9224	188	69	36.70
6	ICPA 2043/ ICPA 2043/LRG 41	254	125	49.21

plant could set a large number of pods even with only a small percentage (10% or more) of viable pollen grains.

Among the twenty-eight F1 hybrids produced by the CMS line ICPA 2047 from sixteen crossings, only one cross, ICPA 2047 × ICP 3181, showed significant seed setting at 43.29 %. The rest exhibited minimal pod setting.

Among the ten hybrids derived from ICPA 2048 x ICP 9224, showed notable seed setting with 36.70 %.

For ICPA 2043 × LRG 41, two F1s descended from ICPA 2043 revealed 49.21 % of seed setting. The test cross between ICPA 2043 and LRG 41, one of the twenty-eight hybrids produced from the three CMS lines ICPA 2047, ICPA 2048 and ICPA 2043, displays 98 % fertility and good pod setting (Table 3). These crosses were subsequently assessed in isolation in *kharif* 2017 and obtained good pod setting because of restoration ability.

All of the plants in the three ICPA 2043 × LRG 41 hybrids were showing fertile during the course of two years, *kharif* 2017 and *kharif* 2018. (Table 3). This demonstrates that the hybrid successfully integrated dominant fertility-restoring genes from the restorer parent and that the hybrid had shown excellent fertility restoration stability over two years. This hybrid may be tested over years in different locations will lead to the potential application of the knowledge in the hybrid pigeonpea breeding programme.

LRG 41 stood out among the testers and displayed perfect male sterility preservation and fertility restoration in the A4 cytoplasm, whereas ICP 3181 and ICP 9224 also had fertility

 $\ensuremath{\text{Table 3.}}\xspace$ Fertility restoration of the Hybrid ICPA 2043 x LRG 41 for two consecutive years

		ICPA 2043			
Tester	_	Total plants	Fertility restoration (%)		
LRG 41	Dept. of Pulses, kharif 2017	120	94		
	Dept. of Pulses, <i>kharif</i> 2017 Dept. of Pulses, <i>kharif</i> 2018	240	98		
Me	an (across two Years)	180	96		

ICPA 2048// ICPA 2048/ ICP 9224

ICPA 2043//ICPA 2043/LRG 41

The successful transmission of restorer genes between genotypes is governed by the underlying genetics of fertility restoration, which is crucial for hybrid breeding programs. This study revealed that the restorer ICP 3181 exhibited monogenic inheritance (3:1) when crossed with ICAP 2047, a trait similarly observed in crossed with ICPA 2048 (Table 4). The fertility restoration of in hybrids, ICPA 2047 × ICP 3181 and ICPA 2048 × ICP 9224 was controlled by a single dominant gene based on the F2 value. In contrast, the hybrid ICAP 2043 × LRG 41 exhibited a di -genic duplicate dominance inheritance of fertility restoration (Table 4). A study was conducted on the inheritance of fertility restoration in three diverse medium-duration CMS lines of pigeonpea (7). Various cytoplasmic sources with the same set of male parents expressed fertility restoration differently (7). Research indicates that one or two fertility-recovering genes were in charge of restoring fertility in the A4 CMS lines of pigeonpea (8). In soyabean, it was found that the monogenic to digenic gene in CMS lines of NJCMS2 A controlled fertility restoration (8).

The variable responses of CMS lines with the same cytoplasm in test crosses may result from the accumulation of recessive nuclear genes from recurrent male ancestors during backcrossing. The accumulation might lead to changes in the cytoplasmic genes governing male sterility (9). The presence of modifier genes that affect the expression and penetrance of fertility-restoring genes may also contribute to differences in the segregation pattern (10). In the current study, the nuclear background of male-sterile and fertility-restoring lines had an impact on the inheritance of male fertility restoration in CMS lines of pigeonpea. 30 male sterile lines from Zea mays (maize) were divided into several groups based on their ability to restore fertility (1). Due to the limited number of CMS lines in this study, grouping based on fertility restoration patterns was not feasible, potentially limiting the breadth of conclusions regarding genetic variability. The fertility of hybrids between the same female and particular males in sorghum (Sorghum vulgare) varied, according to (11). Similarly, the variation in fertility restoration

 X^{2} 3:1=3.14^{NS}

X² 3:1=3.74^{NS}

Table 4. Segregation ratios for male fertile and sterile plants in F1 and F2 generations of crosses involving A4 CMS lines and fertility restorers in Pigeonpea

	C+	Generation —	Number of plants					~	
	Cross		Total	Fer	tile	Sterile	Ratio	L	ni-square probability
	47 × ICP 3181	F 1	16	1	6	0	-		-
ICPA 204	47 × ICP 3101	F ₂	340	24	18	92	3:1		0.98
	48 × ICP 9224	F_1	15	1	5	0	-		-
ICPA 204	48 × ICP 9224	F ₂	315	29	90	25	3:1		0.99
		F_1	17	1	7	0	-		-
ICPA 2043 × LRG 41		F_2	421	38	37	34	15:1		0.99
		Po	llen fertilit	y of testcr	oss popula	ations			
S.No.	т	Test crosses		No. of plants					Segregation ratio
5.NU.	rest crosses			Total	Fertile	Partia	fertile	Sterile	
1	ICPA 2047/	/ ICPA 2047/ ICP 3181		96	68	()	28	X ² 3:1=2.78 ^{NS}

Non-significant at p=0.05 & 0.01 level respectively

2

3

95

111

62

94

0

0

33

17

observed in the current study could be attributed to the presence or absence of key fertility genes. Variations in fertility restoration observed in the current study could be attributed to the presence or absence of fertility genes (12). Variations in sunflower fertility restoration have been reported using several testers and the same cytoplasm (*Helianthus annus* L.) (13). Studies at ICRISAT (International Crops Research Institute for the semi-arid Tropics) revealed the presence of one to three dominant genes for restoring male fertility in all CMS sources of pigeonpea.

Conclusion

According to the research, ICPA 2043 from A4 cytoplasm is a consistent male sterile line throughout a number of seasons because of genetic uniformity and environmental adaptability. The LRG 41 variety demonstrates good fertility restoration, unlike cultivars ICP 3181 and ICP 9224, which exhibited variability in fertility restoration among testers. This inconsistency may be attributed to genetic impurities in the male parents, potentially resulting from spontaneous outcrossing and difficulties in maintaining genetic stocks under natural pollination Understanding the inheritance pattern of fertility restoration facilitated the development of new restorers for the same cytoplasmic source. The differences in behaviour of the three fertility restorer lines ICP 3181, ICP 9224 and LRG 41 were attributed to the interactions of the three female parents' different nuclear genes. The pigeonpea variety LRG 41 was identified as a stable restorer, having significant implications for future breeding programs.

Future directions

Hybrid pigeonpea, with its potential for higher yields and improved quality, can play a vital role in meeting this demand and ensuring food security. In conclusion, hybrid pigeonpea has demonstrated substantial potential for yield enhancement, offering numerous benefits to both farmers and the broader agricultural industry. Continued research, for fertility restorer development and promotion of these hybrids can pave the way for a more productive and sustainable future in pigeon pea cultivation.

Acknowledgements

The authors acknowledge the institution's provision of laboratory and field facilities for conducting the research.

Authors' contributions

ATH has contributed to writing the original draft, reviewing and editing, methodology, data curation and conceptualization. SK has done review and editing and data curation. MS contributed through review and editing, data curation and methodology. WG conducted formal analysis. MKM contributed to formal analysis, review and editing. SM Samyuktha also conducted formal analysis, review and editing. All authors read and approved the final manuscript.

Compliance with ethical standards

Conflict of interest: The authors declare that there is no conflict of interest.

Ethical issues: None

References

- Saroj SK, Singh MN, Vishwakarma MK, Singh T, Mishra VK. Identification of stable restorers and genetics of fertility restoration in late-maturing pigeon pea [*Cajanus cajan* (L.) Millspaugh]. Plant Breed. 2015;134(6):696-702. https://doi.org/10.1111/pbr.12309
- Yadav MK, Patel C, Singh RS, Singh KK, Balasubramanian R, Mall RK. Assessment of climate change impact on different pigeonpea maturity groups in north Indian condition. J Agrometeorol. 2021;23:82-92. https://doi.org/10.54386/jam.v23i1.92
- Kalaimagal T, Muthiah A, Kumaresan D. Inheritance of fertility restoration for A2 cytoplasm of pigeonpea [*Cajanus scarabaeoides* (L.) Thouars]. Crop Res. 2012;43(1, 2 and 3):185-88.
- Bohra A, Singh IP, Yadav AK, Pathak A, Soren KR, Chaturvedi SK, et al. Utility of informative SSR markers in the molecular characterization of cytoplasmic genetic male sterility-based hybrid and its parents in pigeon pea. Natl Acad Sci Lett. 2015;38:13-19. https://doi.org/10.1007/s40009-014-0288-6
- Ranjani MS, Jayamani P. Identification of fertility restorers for A2 cytoplasm-based CGMS lines and development of short duration hybrids in pigeon pea (*Cajanus cajan* (L.) Mill sp.). J Genet. 2023;102:28. https://doi.org/10.1007/s12041-023-01426-w
- Chauhan RM, Parmar LD, Patel PT, Tikka SBS. Fertility restoration in cytoplasmic genetic male sterile line of pigeon pea [*Cajanus cajan* (L.) Millsp.] Derived from *Cajanus scarabaeoides*. Indian J Genet Plant Breed. 2004;64(2):112-14.
- Dalvi VA, Saxena KB, Madrap IA. Fertility restoration in cytoplasmic nuclear male-sterile lines derived from three wild relatives of pigeon pea. J Hered. 2008;99(6):671-73. https://doi.org/10.1093/ jhered/esn034
- Bai YN, Gai JY. Development of a new cytoplasmic-nuclear malesterility line of soybean and inheritance of its male-fertility restorability. Agricultural and Food Sciences 2006;125(1):85-88. https://doi.org/10.1111/j.1439-0523.2006.01191.x
- Nadarajan N, Ganesh S, Petchiammal KI. Fertility restoration studies in short duration red gram [*Cajanus cajan* (L.) Mill sp.] hybrids involving CGMS system. Madras Agric J. 2008;95(7-12):320-27.
- Patil MB, Khule AA, Satish K, Chauhan RM, Patel PT. Fertility restoration studies in short duration red gram [*Cajanus cajan* L.) Mill Sp.] hybrids involving CGMS system. Ecol Environ Conserv. 2016;22:313-17.
- Sultana R, Saxena KB. Identification of new fertility restorers for development of early maturing pigeon pea hybrids. Legume Res. 2017;40(4):639-42. https://doi.org/10.18805/ijar.v0iOF.7647
- Kumaresan A, Jayamani P. Identification of restorers and maintainers for A2 and A4 cytoplasm based CGMS lines in early maturing pigeon pea (*Cajanus cajan* (L.) Mill sp.). Electron J Plant Breed. 2019;10:706-11. https://doi.org/10.5958/0975-928X.2019.00090.5
- Vanishree, Byre Gowda M, Ramesh S, Ramappa HK. Restorers and maintainers of A2 and A4 cytoplasm-based CMS lines in pigeon pea. Int J Curr Microbiol App Sci. 2018;7(12):480-88. https:// doi.org/10.20546/ijcmas.2018.712.060