





### **REVIEW ARTICLE**

# Role of brassinolide in enhancing plant tolerance to salinity stress

Jyostnarani Pradhan¹\*, Padmanabh Dwivedi², Krishna Kumar Jangid³, Kartik Pramanik⁴\*, Teikur Majaw¹, K Prasad⁵, Chaudhari Vishakha Ramkrushna¹, Aman Jaiswal⁶, Geeta Kumari⁶, Devashish Pathak⁶ & Himani Priya⁷

<sup>1</sup>Department of Botany, Plant Physiology and Biochemistry, College of Basic Sciences and Humanities, Dr. Rajendra Prasad Central Agriculture University, Pusa, Samastipur 848 125, Bihar, India

<sup>2</sup>Department of Plant Physiology, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi 221 005, Uttar Pradesh, India 
<sup>3</sup>School of Water Stress Management, Indian Council of Agricultural Research-National Institute of Abiotic Stress Management, Malegaon (Kh),

Baramati, Pune 413 115, Maharashtra, India

<sup>4</sup>Department of Horticulture, M S Swaminathan School of Agriculture, Centurion University of Technology and Management, Paralakhemundi 761 211, Odisha, India

<sup>5</sup>Department of Horticulture, Tirhut College of Agriculture, Dr. Rajendra Prasad Central Agriculture University, Pusa, Samastipur 848 125, Bihar, India <sup>6</sup>Department of Microbiology, College of Basic Sciences and Humanities, Dr. Rajendra Prasad Central Agriculture University, Pusa, Samastipur 848 125, Bihar, India

<sup>7</sup>Division of Natural Resource Management, Indian Council of Agricultural Research-Indian Agricultural Research Institute, Gauria Karma, Barhi, Hazaribagh 825 405, Jharkhand, India

\*Correspondence email - jyotsna.pradhan@rpcau.ac.in, kartik.pramanik@cutm.ac.in

Received: 02 January 2025; Accepted: 31 August 2025; Available online: Version 1.0: 27 November 2025

Cite this article: Jyostnarani P, Padmanabh D, Krishna KJ, Kartik P, Teikur M, Prasad K, Chaudhari VR, Aman J, Geeta K, Devashish P, Himani P. Role of brassinolide in enhancing plant tolerance to salinity stress. Plant Science Today (Early Access). https://doi.org/10.14719/pst.7032

### **Abstract**

Salinity stress is a major constraint to global agriculture, reducing crop yields, degrading arable land and causing economic losses. In many regions, it is intensified by factors such as seawater intrusion, poor irrigation practices and climate-related changes in water availability. While mild salinity may not severely hinder growth, high salinity disrupts ion homeostasis, induces oxidative stress and impairs physiological processes including water and nutrient transport, leading to stunted growth and reduced biomass. Brassinolide (BL), a naturally occurring plant steroid hormone, plays a pivotal role in mitigating salinity stress by regulating ion balance, enhancing antioxidant defense systems, modulating stress-responsive gene expression (e.g., ion transporters, antioxidant enzymes) and promoting growth and development. Its exogenous application through foliar sprays, seed priming or soil treatments has shown promise in improving salt tolerance. This review outlines the mechanisms of salinity stress in plants and highlights the role of BL in alleviating its effects, with emphasis on signalling pathways, ion homeostasis, regulation of Na/H antiporters and reactive oxygen species (ROS) scavenging.

Keywords: antioxidant defense; Brassinolide; ion transporters; plant growth regulation; salinity stress; stress signalling

### Introduction

The biggest challenge in agriculture in this era is to feed the fastgrowing population from a gradually reducing fertile land area. Around one billion hectares of total area, which accounts approximately 7 % of the earth is currently facing salinity stress issues (1). While many of these problems are because of natural biogeochemical processes, however about 30 % is due to human induced salinization particularly for of irrigation system (2). The use of lower-quality irrigation water exacerbates soil salinity, while additional sources include seawater intrusion, wind-blown (aeolian) salt deposition and salt release from the weathering of parent materials. Climate change further contributes by increasing saltwater inundation in coastal regions and altering water availability, thereby intensifying irrigation demands. Salinity stress refers to the presence of excessive amounts of soluble salts in the soil or water surrounding plant roots, which can adversely affect plant growth and productivity (3). It influences many physiological processes including water uptake, osmotic balance, nutrient acquisition and photosynthesis. High

salt concentrations cause ion toxicity due to increased Na<sup>+</sup>and Cllevels, leading to tissue damage (4). Salinity stress also aggravates oxidative stress, inhibits growth and causes nutrient imbalances. However, certain plants have evolved specific salt tolerance mechanisms to cope with these conditions (5). Brassinolide, a member of the brassinosteroid group–polyhydroxylated steroidal phytohormones–was first identified in the pollen grains of *Brassica napus* in the early 1970s (6, 7). It plays a key role in multiple physiological processes such as growth, development and adaptation to environmental stresses (8, 9). BL is involved in cell expansion and growth by regulating genes related to cell wall synthesis and modification, promoting larger cell size and enhanced tissue growth (10).

BL can increase seed germination by promoting endosperm rupture and reducing the inhibitory effects of Abscisic Acid (ABA) (11). Leaf development is also influenced by BL (12). In terms of reproductive development, BL regulates flower formation, pollen germination and fertilization, thereby supporting successful pollination and contributing to fruit

development and seed production (13). BL can delay senescence and aging processes in plants by regulating genes associated with chlorophyll degradation and senescence, thus extending the functional lifespan of leaves (14). It is very important for plant stress responses, helping plants to tolerate drought, salinity, cold, heat and oxidative stress by regulating antioxidant systems, osmotic balance and genes responsible for stress (6). It acts as a molecular signal, transmitting growth and developmental signals within the plant (15). application of BL has been shown to promote flowering and fruit development in mango (13), strawberry (16), leading to higher fruit set, larger fruits and increased yield in various crops (17). The primary goal of this review paper is to systematically analyze and enumerate the results of recent research, providing a comprehensive understanding of the mechanisms by which BL influences responses to salinity stress. This review critically evaluates the efficacy of BL in enhancing salt tolerance by synthesizing and analysing findings from recent experimental studies, including physiological, biochemical and molecular research on various crop species. The aim is to provide an integrated understanding of BL-mediated mechanisms, highlight current knowledge gaps and identify priority areas for future investigation to guide both applied and fundamental research in salinity stress management.

### **Mechanisms of salinity stress**

Salinity refers to the concentration of dissolved salts in soil or water, commonly measured in parts per thousand (ppt), milligrams per litre (mg L-1) or electrical conductivity (dS m-1) (18). High salinity reduces water uptake, disrupts nutrient balance and can cause sodium and chloride toxicity, ultimately lowering crop yields. As per status report on hydrology, in India, it is prevalent in arid and semi-arid regions such as Rajasthan, Gujarat, Punjab, Haryana and coastal belts (19). Causes include seawater intrusion, saline irrigation water, poor drainage, saline groundwater up flow and unsustainable farming practices. Effective management requires improved irrigation, drainage and salt-tolerant crop cultivation. Salinity stress has harmful effect on plant physiological processes through many mechanisms (20). Firstly, it causes osmotic stress by reducing water availability and hindering cell expansion and development. Secondly, it causes ionic toxicity as excessive salts accumulate in plant tissues, disrupting cellular processes and causing ion imbalances. Plant suffered from salinity stress leads to the evolution of reactive oxygen species (ROS) including superoxide (O), hydroxyl radicals (OH) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). This occurs through mechanisms such as ion imbalance, oxidative burst mediated by NADPH (nicotinamide adenine dinucleotide phosphate, reduced form) oxidases, disruption of the electron transport chain and enzymatic reactions. The production of ROS can cause cellular damage, affecting lipids, proteins and DNA (21). Salinity stress also disrupts the balance of essential mineral ions, affecting nutrient uptake and transport (22). This stress also causes hormonal imbalance, impairs morphology, triggers genetic and molecular deviations and interferes with normal growth and development processes (23). Salinity stress disrupts ion homeostasis and osmotic balance in plants by causing an imbalance of ions and reducing water availability (24). When plant is exposed to high salt concentrations, it experiences an influx of sodium (Na+) and chloride (Cl<sup>-</sup>) ions into its root system. This disturbs the ion homeostasis, as plants typically maintain a precise balance of important ions such as K<sup>+</sup> (potassium), Ca<sup>2+</sup>(calcium) and Mg<sup>2+</sup> (magnesium) within their cells. The excessive accumulation of Na<sup>+</sup> ions disrupts this balance and can interfere with vital cellular processes (25). High salt levels in the surrounding environment create an osmotic imbalance (26). The presence of salts outside the plant cells increases the osmotic potential, reducing the availability of water for plant uptake. As a result, plants unable to maintain adequate water, leading to osmotic moisture stress and cellular dehydration (27).

### Brassinolide as a plant hormone

Brassinolide has vital role in influencing various plant physiological processes including growth, development and stress adaptive response (Table 1 & Fig. 1). It is involved in promoting cell elongation, cell division and differentiation, leading to increased plant growth and biomass (28). Brassinolide also influences seed germination, adventitious root development, flowering and fruit development (29-31). One of the key functions of BL is its role in promoting hypocotyl elongation (32). It stimulates the elongation of cells in the stem, resulting in increased plant height (33). Additionally, BL regulates leaf expansion, leading to larger leaf size and increased photosynthetic capacity. This hormone is also involved in the control of vascular tissue development, which ensures efficient transport of water, nutrients and hormones throughout the plant. It prolongs the life of leaves by delaying

**Table 1.** Function of brassinolide in plant physiological processes

Sl. no.	Role of BL	Mechanism	Crop	Reference
1	Cell expansion and elongation	Modulating gibberellin metabolism	Rice	36
2	Hypocotyl elongation	Expression of auxin responses genes; BZR1/BES1 interacts with auxin response factors (ARFs)	Arabidopsis	37
3	Cell division and differentiation	CycD3 induction pathway; BZR1/BES1 directly bind to the promoter region of CycD3 genes	Arabidopsis	38
4	Seed germination	Break seed dormancy, antioxidative defenses	Brassica juncea	39
5	Regulating fiber elongation	GhBES1.4-mediated regulation of fiber development	Cotton	40
6	Root development	BR signaling via BRI1 KINASE INHIBITOR1	Potato	41
7	Flowering and fruit development	Induced parthenocarpic growth	Cucumber	42
8	Stress responses	Differentially expressed genes (DEGs)	Tomato	43
9	Defense against pathogens	Expression of Br genes; hormonal signaling	Barley	44
10	Senescence regulation	Regulating its chlorophyll metabolic pathway and endogenous hormones	Brassica rapa subsp. chinensis	34
11	Vascular tissue differentiation	Govern stem vascular cell division; expansion; influence cell cycle progression and interact with auxin and other hormones to ensure proper stem growth as well as	Brassica spp.	28

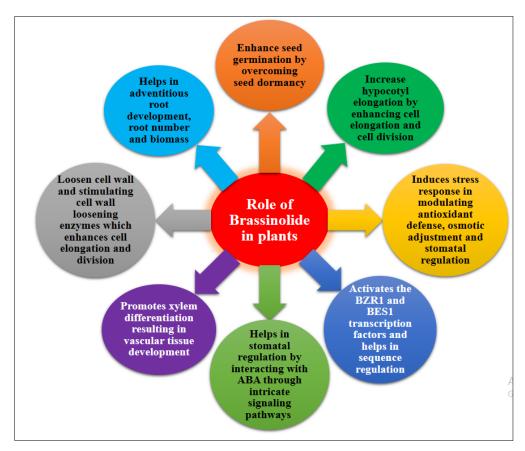


Fig. 1. Function of brassinolide in plant physiological processes.

senescence. It contributes to sustained photosynthetic activity by controlling the expression of genes linked to ageing (34). Brassinolide is known to enhance plant tolerance to various abiotic stresses such as high and low temperatures, drought, salinity and heavy metals. Brassinolide influences stomatal aperture, affecting gas exchange and water loss (35). It helps plants adapt to adverse conditions by influencing the expression of stress-responsive genes and promoting the synthesis of protective compounds. Furthermore, BL has been

found to improve plant resistance against pathogens by activating defense mechanisms.

### Signaling pathways and mechanisms of brasssinolide action

Brasssinolide signalling pathway is complex and tightly regulated, it governs various physiological functions such as cell expansion, cell division, root development, flowering, stress responses and defense against pathogens. Fig. 2 represents a simplified version of this signaling pathway. At the core of BL signaling are the cell surface receptors BRI1 and BAK1, which

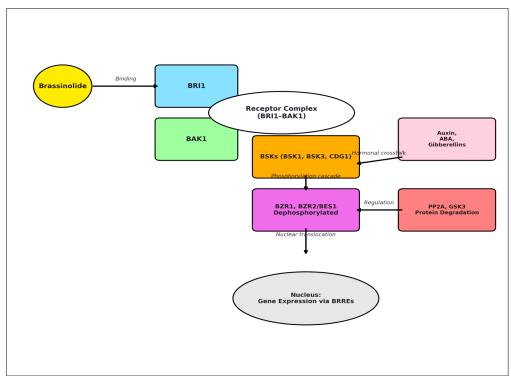


Fig. 2. Signaling pathways and mechanisms of Brassinolide action.

perceive and bind BL. The binding of BL to BRI1 triggers conformational changes and the formation of a receptor complex with BAK1 (15). This receptor complex activates intracellular signalling cascades, known as the brassinosteroid signalling kinases (BSKs) pathway, through a phosphorylation cascade involving multiple protein kinases. Key components of this pathway include BSK1, BSK3 and CDG1 (45). The initiation of this pathway leads to the dephosphorylation and translocation of transcription factors namely BZR1 and BZR2/ BES1 into the nucleus. Once in the nucleus, BZR1 and BZR2/ BES1 control the expression of specific genes by binding to target DNA sequences called brassinosteroid response elements (BRREs) in the promoters of these genes (46). Depending on the context, they can either activate or repress the expression of target genes. These target genes are involved in important processes such as cell elongation, cell division, hormone biosynthesis, stress responses and other physiological processes. Additionally, various hormone-signalling pathways interact with BL signalling including auxin, ABA and gibberellins, allowing for crosstalk and integration of various hormonal signals in plant physiology (47). Furthermore, protein degradation plays an important role in the regulation of BL responses. Proteins like protein phosphatase 2A (PP2A) and glycogen synthase kinase-3 (GSK3) are involved in the degradation of BZR1 and BZR2/BES1, ensuring proper control of BL signalling.

### Impacts of exogenous BL on salinity stress

Exogenous BL application alleviates the detrimental effects of salinity stress (Table 2 & Fig. 3), leading to improved plant adaptation, survival and crop productivity under saline conditions (48). BL improves salt tolerance by promoting ion homeostasis, reducing the accumulation of toxic ions and enhancing the activity of ion transporters (49). BL also enhances water relations by increasing water uptake efficiency and reducing water loss through stomatal regulation (50). It boosts antioxidant defense by improving antioxidant enzyme activity

and defending plants from oxidative injury. Exogenous application of BL modulates gene expression particularly by upregulating stress-responsive genes associated with salinity tolerance. Such application improves photosynthetic efficiency, promotes growth and development by stimulating root and shoot growth and balances stress-responsive hormones.

Alleviation of ion imbalance: Salinity stress increases the accumulation of Na<sup>+</sup>ions in plant tissues, which can disrupt ion homeostasis and impair plant growth (62). Exogenous application of Brassinolide (BL) can alleviate the negative effects of salinity stress on plants by mitigating ion imbalance (51). BL regulates ion uptake, restricts sodium uptake, enhances the potassium-sodium ratio, stimulates ion transporters and modulates ion channel activity (63). It promotes the uptake of minerals while limiting essential excessive sodium accumulation. BL also enhances the expression of ion transporters, such as SOS1 (sodium/hydrogen antiporter) and stimulates potassium uptake, improving the potassium/sodium ratio (64). Furthermore, BL regulates ion channels such as inward-rectifying K<sup>+</sup> channels to maintain ion balance (65). Overall, BL helps in maintaining ion homeostasis, minimizing the negative consequences of salinity stress on plant physiology. Exogenous BL applications witnessed to enhance the absorption of essential mineral nutrients including potassium, calcium and magnesium, which helps to maintain a favourable balance between Na<sup>+</sup> and other cations, reducing the negative impact of excessive Na<sup>+</sup> accumulation (66). BL treatment has been shown to increase the uptake of potassium and enhance its transport to shoots by activation of K<sup>+</sup> transporters and channels, enhanced proton pump activity (H<sup>+</sup>-ATPase) (81) and improved root system architecture (82). As a result, the potassium/sodium ratio is improved, which causes osmotic balance, reducing the negative effects of salinity stress.

**Na<sup>+</sup> exclusion and K<sup>+</sup> retention:** High salt levels in soil results uptake of excessive Na<sup>+</sup> by plant roots, which can be detrimental to plant growth. BL treatment is shown to enhance

Table 2. Role of Brassinolide in salt tolerance mechanism of different crop species

Sl. no.	Exogenous application (concentration)	Mechanism	Crop	Reference
1	24-epibrassinolide and kinetin (100 nM)	Modulation of antioxidant and osmolyte metabolism	Solanum lycopersicum	51
2	24-epiBL (10 <sup>-7</sup> mM)	Maintaining ionic balance (K <sup>+</sup> /Na <sup>+</sup> ); antioxidative enzyme activities	Glycine max L.	52
3	24-epibrassinolide (10 <sup>-6</sup> M)	Signaling of nitric oxide and pathways of antioxidant defense	Brassica juncea L. cv. varuna	53
4	24-epibrassinolide (2 μM)	Promotion of Ca <sup>2+</sup> accumulation in roots	Arabidopsis	49
5	Brassinosteroids	Regulates over expression of key receptor SIBRI1	S. lycopersicum	54
6	24-epibrassinolide (200 mL L <sup>-1</sup> )	Increased leaf gas exchange and water use efficiency	Ficus carica	50
7	28-homobrassinolide	Increasing carboxylation efficiency and the activity of ROS scavenging system	Vigna radiata	55
8	24-epibrassinolide (1.0 µmol L <sup>-1</sup> )	Activation of antioxidant defense systems	Oryza sativa L.	48
9	24-epibrassinolide (0.01 μM)	Activation of antioxidant defense systems	Zea mays	56
10	24-epibrassinolide (100 ppm)	Increased osmolytes activities, antioxidants defence system and limiting Na† uptake while keeping the K† level in leaves stable	Hordeum vulgare L.	57
11	24-epibrassinolide seed priming	Prevented the salinity mediated photosynthetic inhibition	G. max L.	58
12	Brassinosteroids (0.2 mg L <sup>-1</sup> )	NHX-Type Na <sup>+</sup> (K <sup>+</sup> )/H <sup>+</sup> antiporter transcription regulation	Malus hupehensis	59
13	24-epibrassinolide (0.1 μM)	Increasing the expression of AhNHX7 and AhNHX8 and removing ROS	Arachis hypogaea	60
14	24-epibrassinolide, bikinin and brassinazole (0.01 μM – 0.01 μM)	Increased osmolytes activities, antioxidants defence system	H. vulgare L.	61

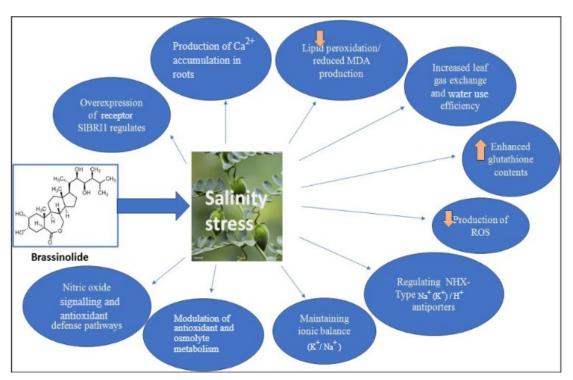


Fig. 3. Impact of BL in plant growth and development under salinity stress.

the exclusion of Na<sup>+</sup> ions from plant roots (47). It helps in regulating the activity of ion transporters, such as the sodium/ hydrogen antiporter (SOS1), which actively pumps Na<sup>+</sup> out of the roots and reduces their accumulation in the shoot tissues (Fig. 3). This sodium exclusion mechanism helps prevent sodium toxicity and maintains a more favourable sodium-topotassium ratio in plant tissues. Salinity stress often disrupts K<sup>+</sup> homeostasis in plants, leading to potassium deficiency (67). However, exogenous BL application can help retain higher levels of potassium in plant tissues under saline conditions (48). BL enhances the activity of K<sup>+</sup> transporters, such as high-affinity potassium transporters (HKTs), which facilitate the absorption and transport of K<sup>+</sup> ions in roots. BL treatment has been reported to influence the expression of genes involved in potassium uptake and translocation, contributing to improved potassium retention in plants.

Regulation of ion channels and transporters: BL is witnessed to regulate the ion transporter expression and activity such as plasma membrane H<sup>+</sup>-ATPases and Na<sup>+</sup>/H<sup>+</sup> antiporters (59). These transporters are responsible for maintaining ion gradients across the cell membrane. BL enhances the activity of plasma membrane H<sup>+</sup>-ATPases, which helps maintain a favourable electrochemical gradient for nutrient uptake and regulates Na<sup>+</sup> exclusion from the cytoplasm. Exogenous BL application influences the activity of ion channels involved in ion transport. It has been reported to regulate the activity of K<sup>+</sup> channels, including inward-rectifying K<sup>+</sup> channels (KIR) and outward-rectifying K<sup>+</sup> channels (KOR) (68). This regulation helps maintain proper potassium ion uptake, transport and distribution, which is important for osmotic adjustment and stress tolerance in plants.

### Reactive Oxygen Species scavenging and antioxidant defense

Exogenous application of BL revealed to alleviate the negative effects of salinity stress on plants by influencing in scavenging of ROS and defense systems of antioxidant (69). Salinity stress often leads to the accumulation of ROS, including  $H_2O_2$ ,  $(O_2-)$ 

and OH, which can cause oxidative damage to plant cells. BL treatment helps in the scavenging of ROS by improving the activities of antioxidant and non-enzymatic antioxidants. BL treatment enhances the activities of various antioxidant enzymes including peroxidase (POD), superoxide dismutase (SOD), catalase (CAT) and ascorbate peroxidase (APX) (70). These enzymes play a crucial role in converting SOD to O2- and finally into  $H_2O_2$ . CAT metabolizes  $H_2O_2$  into water and oxygen, while PODs detoxify  $H_2O_2$  and organic hydroperoxides. Nonenzymatic antioxidant accumulation includes ascorbic acid (vitamin C), glutathione (GSH) and carotenoids (71). These compounds help reduce ROS directly and protect cellular components from oxidative injury.

### Modulation of stress-responsive gene expression

Exogenous BL application has been shown to regulate the action of stress-responsive genes in plants under salinity stress (71). Salinity stress triggers a complex network of gene expression changes to help plants acclimatize and tolerate with the adverse conditions. BL treatment has been found to control the gene expression involved in stress response mechanisms. BL treatment can regulate the expression of genes involved in ion transport and ion channel activities, such as those encoding ion transporters (e.g., SOS1, NHX) and potassium channels (72). BL can influence the associated genic expression with ABA biosynthesis and signaling, as well as those involved in the crosstalk between ABA and brassinosteroids (73). These interactions help regulate stomatal closure, osmotic adjustment and other adaptive responses to salinity stress. BL treatment can activate defense-related genes involved in stress signaling pathways. These genes may include receptor-like kinases (RLKs), pathogenesis related (PR) proteins, heat shock proteins (HSPs) and other stress-responsive genes (74).

# Activation of stress-responsive transcription factors and signaling pathways

BL treatment can activate stress-responsive transcription factors, such as BZR/BES transcription factor SIBZR1 favorable

regulates BR signalling and salt stress tolerance in tomato and Arabidopsis (75). These transcription factors cause the upregulation of stress-related genes by binding to particular DNA sequences in their promoter regions. The upregulated genes are involved in various stress responses including osmotic regulation, detoxification and antioxidant defence. Signalling pathways play a crucial role in the plant's ability to perceive and respond to the stress, leading to adaptive physiological and biochemical changes. One important mechanism involved in plant stress responses is the mitogen-activated protein kinase (MAPK) signalling system. The activation of MAPKs triggers a series of phosphorylation events that regulate the expression of stress-responsive genes, resulting in enhanced stress tolerance (76). Ca<sup>2+</sup> has key role as messengers in stress signalling. This calcium influx activates calcium-dependent protein kinases (CDPKs) and other Ca-binding proteins, which initiate downstream signalling events. The calcium-mediated signalling pathway is involved in regulating various stress-responsive genes and physiological processes including osmotic regulation, ion transport and antioxidant defence. It can modulate the crosstalk between these hormones to regulate stress responses.

### Regulation of NHX-type Na<sup>+</sup>(K<sup>+</sup>)/H<sup>+</sup>antiporter transcription

Exogenous application of BL has been found to alleviate saline stress in plants by influencing the transcription of NHX-type Na<sup>+</sup> (K+)/H+ antiporter genes (59). BL activates the brassinosteroid signaling pathway, leading to the activation of transcription factors such as BZR1. These transcription factors bind to specific promoter regions of NHX-type Na<sup>+</sup>(K<sup>+</sup>)/H<sup>+</sup> antiporter genes, resulting in their transcriptional activation. The synthesized NHX proteins function as ion transporters, facilitating the efflux of Na<sup>+</sup> ions from the cytoplasm and its sequestration into vacuoles (77). This mechanism helps maintain ion homeostasis and prevents the toxic effects of sodium accumulation in plant cells under salt stress conditions. NHX proteins function as ion transporters that mediate the exchange of Na<sup>+</sup> and K<sup>+</sup> ions with protons (H<sup>+</sup>) across membranes, thus reducing the toxic effects of sodium accumulation in plant cells under salt stress conditions (78).

### Efficient ion transport and ion homeostasis

Exogenous application of BL (BL) has been shown to alleviate salt stress in plants by preventing excessive Na<sup>+</sup> accumulation (49). BL treatment influences the activity and expression of ion transporters involved in Na<sup>+</sup> uptake, compartmentalization and efflux (79). It enhances the expression of H<sup>+</sup>-ATPases, which facilitate the efflux of Na<sup>+</sup> from cells, preventing its accumulation. Additionally, BL increases the expression of K<sup>+</sup> transporters, promoting the uptake of K<sup>+</sup> ions. This shift in ion transport helps maintain a favourable K<sup>+</sup>/Na<sup>+</sup> ratio and reduces Na<sup>+</sup> accumulation in cells. BL treatment affects the activity of ion channels involved in Na<sup>+</sup> transport. It has been found to inhibit the activity of non-selective cation channels (NSCCs), which are responsible for Na<sup>+</sup> influx into cells (80). By reducing Na<sup>+</sup> influx, BL helps prevent excessive Na<sup>+</sup> accumulation under salt stress conditions.

### Regulation of Salt Overly Sensitive (SOS) pathway

The SOS pathway is a key signalling pathway involved in salt stress responses. BL treatment enhances the expression and activity of SOS pathway components including SOS1 and SOS3/ SOS2 (59). This activation of the SOS pathway facilitates Na<sup>+</sup> efflux from cells, reducing its accumulation in the cytoplasm. The SOS pathway consists of several components, including SOS1 (a plasma membrane Na<sup>+</sup>/H<sup>+</sup> antiporter), SOS2 (a serine/ threonine protein kinase) and SOS3 (a calcium sensor). Interaction between BL and SOS components leads to the activation of SOS2 kinase activity, which subsequently phosphorylates and activates the SOS1 antiporter. By regulating the SOS pathway, BL helps plants maintain ion homeostasis and minimize the negative effects of salt stress (79).

### **Conclusion**

Exogenous application of BL offers significant potential for enhancing plant stress tolerance, stimulating growth and improving productivity and resilience under salt-affected conditions. Study reported that BL induce tolerance to salinity stress at specific concentration depending upon the crop species (83). When plants suffered salinity stress, BL helps regulate ion homeostasis by increasing the uptake of essential nutrients like potassium while reducing the uptake of toxic ions such as Na<sup>+</sup> and Cl<sup>-</sup>. It also improves osmotic stress tolerance of plants by increasing the accumulation of compatible solutes, preventing water loss and maintaining cellular water potential. Additionally, BL enhances the antioxidant enzymes activities and increases the content of non-enzymatic antioxidants, protecting plants from oxidative damage caused by ROS. The implications of BL application include improved salt tolerance, enhanced productivity, sustainable agriculture in saline environments and the induction of stress priming effects, leading to improved stress tolerance in subsequent salinity stress episodes. Future research should focus on (i) elucidating the molecular mechanisms and signaling pathways underlying BL-mediated stress tolerance, (ii) exploring genotype-specific responses to optimize its use in breeding programs, (iii) developing efficient, cost-effective and scalable application methods for field conditions and (iv) integrating BL treatment with other agronomic and biotechnological approaches to promote climate-resilient and sustainable crop production in salt-affected ecosystems.

## **Acknowledgements**

We would like to acknowledge all the author contribution for documenting the manuscript with genuine earlier research findings on the entitled topic.

### **Authors' contributions**

JP, PD, KKJ did the conceptualization of article. Collection of research data and interpretation was done by JP, TM and KP<sup>1</sup>. AJ, GK, KP<sup>2</sup> and CVR prepared the manuscript. Language editing was done by HP and DP. All authors read and approved the final manuscript.

### **Compliance with ethical standards**

**Conflict of interest:** Authors do not have any conflict of interests to declare.

Ethical issues: None

### References

- Bashir RN, Bajwa IS, Abbas MZ, Rehman A, Saba T, Bahaj SA, Kolivand H. Internet of things (IoT) assisted soil salinity mapping at irrigation schema level. Applied Water Sci 2022;12(5):105. https://doi.org/10.1007/s13201-022-01619-1
- Hopmans JW, Qureshi AS, Kisekka I, Munns R, Grattan SR, Rengasamy P, et al. Critical knowledge gaps and research priorities in global soil salinity. Adv Agro. 2021;169:1-91. https://doi.org/10.1016/bs.agron.2021.03.001
- Isayenkov SV, Maathuis FJ. Plant salinity stress: many unanswered questions remain. Front in Plant Sci. 2019;10:80. https:// doi.org/10.3389/fpls.2019.00080
- Tavakkoli E, Fatehi F, Coventry S, Rengasamy P, McDonald GK. Additive effects of Na+ and Cl-ions on barley growth under salinity stress. J Exp Botany. 2011;62:2189-203. https://doi.org/10.1093/jxb/erq422
- Acosta-Motos JR, Ortuño MF, Bernal-Vicente A, Diaz-Vivancos P, Sanchez-Blanco MJ, Hernandez JA. Plant responses to salt stress: adaptive mechanisms. Agronomy. 2017;7:18. https://doi.org/10.3390/agronomy7010018
- Li S, Zheng H, Lin L, Wang F, Sui N. Roles of brassinosteroids in plant growth and abiotic stress response. Plant Growth Reg. 2021;93:29-38. https://doi.org/10.1007/s10725-020-00672-7https:// doi.org/10.1007/s10725-020-00684-3
- Rao SS, Vardhini BV, Sujatha E, Anuradha S. Brassinosteroids-a new class of phytohormones. Curr Sci. 2002:1239-45. https:// www.jstor.org/stable/24107046
- Li Z, He Y. Roles of brassinosteroids in plant reproduction. Int J Mole Sci. 2020;21:872. https://doi.org/10.3390/ijms21030872
- Manghwar H, Hussain A, Ali Q, Liu F. Brassinosteroids (BRs) role in plant development and coping with different stresses. Int J Mole Sci. 2022;23:1012. https://doi.org/10.3390/ijms23031012
- Bajguz A, Piotrowska-Niczyporuk A. Synergistic effect of auxins and brassinosteroids on the growth and regulation of metabolite content in the green alga *Chlorella vulgaris* (Trebouxiophyceae). Plant Phys Biochem. 2013;71:290-97. https://doi.org/10.1016/ j.plaphy.2013.08.003
- Ali F, Qanmber G, Li F, Wang Z. Updated role of ABA in seed maturation, dormancy and germination. J Adv Res. 2022a;35:199-214. https://doi.org/10.1016/j.jare.2021.03.011
- Kim SY, Warpeha KM, Huber SC. The brassinosteroid receptor kinase, BRI1, plays a role in seed germination and the release of dormancy by cold stratification. J Plant Phys. 2019;241:153031.https://doi.org/10.1016/j.jplph.2019.153031
- 13. Tepkaew T, Khamsuk O, Chumpookam J, Sonjaroon W, Jutamanee K. Exogenous brassinosteroids regulate mango fruit set through inflorescence development and pollen fertility. Horti Sci Techno. 2022;40:481-95.https://doi.org/10.7235/HORT.20220043
- Kovtun IS, Kukharenko NE, Kusnetsov W, Khripach VA, Efimova MV.
   Effect of lactone-and ketone-containing brassinosteroids on photosynthetic activity of barley leaves during aging. Russian J Plant Phys. 2021;68:440-50. https://doi.org/10.1134/S1021443721030080
- 15. Kim EJ, Russinova E. Brassinosteroid signalling. Curr Bio. 2020;30:R294-98. https://doi.org/10.1016/j.cub.2020.02.011
- 16. Ali MM, Anwar R, Malik AU, Khan AS, Ahmad S, Hussain Z, et al. Plant growth and fruit quality response of strawberry is improved after exogenous application of 24-epibrassinolide. J Plant Growth Reg. 2022b;41:1786-99. https://doi.org/10.1007/s00344-021-10422-2
- Khatoon F, Kundu M, Mir H, Nahakpam S. Efficacy of foliar feeding of brassinosteroid to improve growth, yield and fruit quality of strawberry (*Fragaria*× ananassa Duch.) grown under subtropical plain. Commun Soil Sci Plant Analy. 2021;52:803-14. https:// doi.org/10.1080/00103624.2020.1869765
- 18. Corwin DL, Yemoto K. Salinity: Electrical conductivity and total

- dissolved solids. Soil Sci Soc America J. 2020;84(5):1442-61. https://doi.org/10.1002/saj2.20154
- 19. Inda AZ. Status report on hyppology og.
- Shahid MA, Sarkhosh A, Khan N, Balal RM, Ali S, Rossi L, et al. Insights into the physiological and biochemical impacts of salt stress on plant growth and development. Agro. 2020;10:938. https://doi.org/10.3390/agronomy10070938
- Juan CA, Pérez de la Lastra JM, Plou FJ, Pérez-Lebeña E. The chemistry of reactive oxygen species (ROS) revisited: outlining their role in biological macromolecules (DNA, lipids and proteins) and induced pathologies. Int J Mol Sci. 2021; 22(9): 4642. https:// doi.org/10.3390/ijms22094642
- Tammam AA, Rabei AMSM, Pessarakli M, El-Aggan WH. Vermicompost and its role in alleviation of salt stress in plants-I. Impact of vermicompost on growth and nutrient uptake of salt-stressed plants. J Plant Nutr. 2023;46:1446-57. https://doi.org/10.1080/01904167.2022.2072741
- Hussain S, Shaukat M, Ashraf M, Zhu C, Jin Q, Zhang J. Salinity stress in arid and semi-arid climates: Effects and management in field crops. Climate Change Agri. 2019;13:201-26. https://doi.org/10.5772/ intechopen.78427https://doi.org/10.5772/intechopen.87982
- Amin I, Rasool S, Mir MA, Wani W, Masoodi KZ, Ahmad P. Ion homeostasis for salinity tolerance in plants: A molecular approach. Phys Plantarum. 2021;171:578-94. https://doi.org/10.1111/ppl.13185
- Almeida DM, Oliveira MM, Saibo NJ. Regulation of Na<sup>+</sup> and K<sup>+</sup> homeostasis in plants: towards improved salt stress tolerance in crop plants. Genet Mole Bio. 2017;40:326-45. https://doi.org/10.1590/1678-4685-gmb-2016-0106
- Yildiz M, Poyraz İ, Çavdar A, Özgen Y, Beyaz R. Plant responses to salt stress. IntechOpen; 2020. https://doi.org/10.5772/intechopen.93920
- de Oliveira AB, Alencar NL, Gomes-Filho E. Comparison between the water and salt stress effects on plant growth and development. In: Sener A, editor. Responses of Organisms to Water Stress. 2013;4:67-94. https://doi.org/10.5772/54223
- Oh MH, Honey SH, Tax FE. The control of cell expansion, cell division and vascular development by brassinosteroids: a historical perspective. Int J Mole Sci. 2020;21:1743. https://doi.org/10.3390/ ijms21051743
- Soares TF, dos Santos Dias DC, Oliveira AM, Ribeiro DM, dos Santos Dias LA. Exogenous brassinosteroids increase lead stress tolerance in seed germination and seedling growth of *Brassica juncea* L. Ecotoxi Environ Safety. 2020;193:110296. https://doi.org/10.1016/j.ecoenv.2020.110296
- 30. Li Y, Wu Y, Liao W, Hu L, Dawuda MM, Jin X, et al. Nitric oxide is involved in the brassinolide-induced adventitious root development in cucumber. BMC Plant Biology. 2020;20:1-2. https://doi.org/10.1186/s12870-023-04695-w
- Khatoon F, Kundu M, Mir H, Nandita K. Exogenous brassinolide application improves growth, yield and quality of strawberry grown in the subtropics. Erwerbs-Obstbau. 2023;65:2271-79. https://doi.org/10.1007/s10341-023-00981-x
- Zhuang Y, Lian W, Tang X, Qi G, Wang D, Chai G, Zhou G. MYB42 inhibits hypocotyl cell elongation by coordinating brassinosteroid homeostasis and signalling in *Arabidopsis thaliana*. Annals of Botany. 2022;129:403-13. https://doi.org/10.1093/aob/mcab152
- Castorina G, Consonni G. The role of brassinosteroids in controlling plant height in Poaceae: a genetic perspective. Int J Mole Sci. 2020;21:1191. https://doi.org/10.3390/ijms21041191
- Zhu J, Liu X, Huang W, An R, Xu X, Li P. 2, 4-Epibrassinolide delays leaf senescence in Pak choi (*Brassica rapa* subsp. *chinensis*) by regulating its chlorophyll metabolic pathway and endogenous hormones content. Gene. 2023;877:147531. https://doi.org/10.1016/ j.gene.2023.147531
- 35. Shah SH, Parrey ZA, Islam S, Mohammad F, Alamri S, Kalaji HM.

Epibrassinolide application improves *Brassica juncea* L. performance by augmenting gas exchange markers, stomatal behavior, cellular viability, enzymatic activities and yield attributes. South African J Botany. 2023;162:531-44. https://doi.org/10.1016/j.sajb.2023.09.045

- Tong H, Xiao Y, Liu D, Gao S, Liu L, Yin Y, et al. Brassinosteroid regulates cell elongation by modulating gibberellin metabolism in rice. The Plant Cell. 2014;26:4376-93. https://doi.org/10.1105/ tpc.114.132092
- Nemhauser JL, Mockler TC, Chory J. Interdependency of brassinosteroid and auxin signaling in *Arabidopsis*. PLoS Biology. 2004;2:e258. https://doi.org/10.1046/j.1365-313x.2000.00915.x
- Hu Y, Bao F, Li J. Promotive effect of brassinosteroids on cell division involves a distinct CycD3\(\text{Minuter}\) induction pathway in Arabidopsis. The Plant J 2000;24(5):693-701. https://doi.org/10.1046/j.1365-313x.2000.00915.x
- Neha, Twinkle, Mohapatra S, Sirhindi G, Dogra V. Seed priming with brassinolides improves growth and reinforces antioxidative defenses under normal and heat stress conditions in seedlings of *Brassica juncea*. Phy Plantarum. 2022;174:e13814. https:// doi.org/10.1111/ppl.13814
- Liu L, Chen G, Li S, Gu Y, Lu L, Qanmber G, et al. A brassinosteroid transcriptional regulatory network participates in regulating fiber elongation in cotton. Plant Physiology. 2023;191:1985-2000. https:// doi.org/10.1093/plphys/kiac590
- 41. Huang S, Zheng C, Zhao Y, Li Q, Liu J, Deng R, et al. RNA interference knockdown of the brassinosteroid receptor BRI1 in potato (*Solanum tuberosum* L.) reveals novel functions for brassinosteroid signaling in controlling tuberization. Scientia Horticulturae. 2021;290:110516. https://doi.org/10.1016/j.scienta.2021.110516
- 42. Fu FQ, Mao WH, Shi K, Zhou YH, Asami T, Yu JQ. A role of brassinosteroids in early fruit development in cucumber. J Exp Botany. 2008;59:2299-308. https://doi.org/10.1093/jxb/ern093
- Bai C, Zheng Y, Watkins CB, Fu A, Ma L, Gao H, et al. Revealing the specific regulations of brassinolide on tomato fruit chilling injury by integrated multi-omics. Front Nutr. 2021;8:769715. https:// doi.org/10.3389/fnut.2021.769715
- 44. Ali SS, Kumar GS, Khan M, Doohan FM. Brassinosteroid enhances resistance to fusarium diseases of barley. Phytopathology. 2013;103:1260-67. https://doi.org/10.1094/PHYTO-05-13-0111-R
- Kim TW, Guan S, Burlingame AL, Wang ZY. The CDG1 kinase mediates brassinosteroid signal transduction from BRI1 receptor kinase to BSU1 phosphatase and GSK3-like kinase BIN2. Molecular Cell. 2011;43:561-71. https://doi.org/10.1016/j.molcel.2011.05.037
- Wang ZY, Wang Q, Chong K, Wang F, Wang L, Bai M, Jia C. The brassinosteroid signal transduction pathway. Cell Research. 2006;16:427-34. https://doi.org/10.1038/sj.cr.7310054
- Santner A, Estelle M. Recent advances and emerging trends in plant hormone signalling. Nature. 2009;459:1071-78. https:// doi.org/10.1038/nature08122
- Mu DW, Feng NJ, Zheng DF, Zhou H, Liu L, Chen GJ, Mu B. Physiological mechanism of exogenous brassinolide alleviating salt stress injury in rice seedlings. Sci Reports. 2022;12:20439. https://doi.org/10.1038/s41598-022-24747-9
- Wang X, Chai J, Liu W, Zhu X, Liu H, Wei X. Promotion of Ca<sup>2+</sup> accumulation in roots by exogenous brassinosteroids as a key mechanism for their enhancement of plant salt tolerance: A meta-analysis and systematic review. Int J Mole Sci. 2023;24:16123. https://doi.org/10.3390/ijms242216123
- 50. Mardinata Z, Edy ST, Ulpah S. Biochemical responses and leaf gas exchange of fig (*Ficus carica* L.) to water stress, short-term elevated CO<sub>2</sub> levels and brassinolide application. Horticulturae. 2021;7:73. https://doi.org/10.3390/horticulturae7040073
- 51. Ahanger MA, Mir RA, Alyemeni MN, Ahmad P. Combined effects of

- brassinosteroid and kinetin mitigates salinity stress in tomato through the modulation of antioxidant and osmolyte metabolism. Plant Phys Biochem. 2020;147:31-42. https://doi.org/10.1016/j.plaphy.2019.12.007
- 52. Otie V, Udo I, Shao Y, Itam MO, Okamoto H, An P, Eneji EA. Salinity effects on morpho-physiological and yield traits of soybean (*Glycine max* L.) as mediated by foliar spray with brassinolide. Plants. 2021;10:541. https://doi.org/10.3390/plants10030541
- Gupta P, Seth CS. 24-epibrassinolide regulates functional components of nitric oxide signalling and antioxidant defense pathways to alleviate salinity stress in *Brassica juncea* L. cv. Varuna.
   J Plant Growth Reg. 2023;42:4207-22. https://doi.org/10.1007/ s00344-022-10884-y
- 54. Wang S, Liu D, Jin T, Du J, Tian A, Huang Q, et al. Manipulation of brassinosteroid signaling via overexpression of its major receptor SIBRI1 regulates salt tolerance in tomato. Scientia Horticulturae. 2023;322:112457. https://doi.org/10.1016/j.scienta.2023.112457
- Hayat S, Hasan SA, Yusuf M, Hayat Q, Ahmad A. Effect of 28-homobrassinolide on photosynthesis, fluorescence and antioxidant system in the presence or absence of salinity and temperature in *Vigna radiata*. Environ Exp Botany. 2010;69:105-12. https://doi.org/10.1016/j.envexpbot.2010.03.004
- Rattan A, Kapoor D, Kapoor N, Bhardwaj R, Sharma A. Brassinosteroids regulate functional components of antioxidative defense system in salt stressed maize seedlings. J Plant Growth Reg. 2020;39:1465-75. https://doi.org/10.1007/s00344-020-10097-1
- Liaqat S, Umar S, Saffeullah P, Iqbal N, Siddiqi TO, Khan MI. Protective effect of 24-epibrassinolide on barley plants growing under combined stress of salinity and potassium deficiency. J Plant Growth Reg. 2020;39:1543-58. https://doi.org/10.1007/s00344-020-10163-8
- Soliman M, Elkelish A, Souad T, Alhaithloul H, Farooq M. Brassinosteroid seed priming with nitrogen supplementation improves salt tolerance in soybean. Phys Mole Bio Plants. 2020;26:501-11. https://doi.org/10.1007/s12298-020-00765-7
- 59. Su Q, Zheng X, Tian Y, Wang C. Exogenous brassinolide alleviates salt stress in *Malus hupehensis* Rehd. by regulating the transcription of NHX-Type Na<sup>+</sup> (K<sup>+</sup>)/H<sup>+</sup> antiporters. Front Plant Sci. 2020;11:38. https://doi.org/10.3389/fpls.2020.00038
- Li W, Sun J, Zhang X, Ahmad N, Hou L, Zhao C, et al. The mechanisms underlying salt resistance mediated by exogenous application of 24-epibrassinolide in peanut. Int J Mole Sci. 2022;23:6376. https://doi.org/10.3390/ijms23126376
- Groszyk J, Szechyńska-Hebda M. Effects of 24-epibrassinolide, bikinin and brassinazole on barley growth under salinity stress are genotype-and dose-dependent. Agro. 2021;11:259. https:// doi.org/10.3390/agronomy11020259
- Basu S, Kumar A, Benazir I, Kumar G. Reassessing the role of ion homeostasis for improving salinity tolerance in crop plants. Physiologia Plantarum. 2021;171:502-19. https://doi.org/10.1111/ ppl.13112
- 63. El-Banna MF, Al-Huqail AA, Farouk S, Belal BE, El-Kenawy MA, Abd El-Khalek AF. Morpho-physiological and anatomical alterations of salt affected thompson seedless grapevine (*Vitis vinifera* L.) to brassinolide spraying. Horticulturae. 2022;8:568. https://doi.org/10.3390/horticulturae8070568
- Chourasia KN, Lal MK, Tiwari RK, Dev D, Kardile HB, Patil VU, et al. Salinity stress in potato: Understanding physiological, biochemical and molecular responses. Life. 2021;11:545. https://doi.org/10.3390/life11060545
- Yu B, Liu N, Tang S, Qin T, Huang J. Roles of glutamate receptor-like channels (GLRs) in plant growth and response to environmental stimuli. Plants. 2022;11:3450. https://doi.org/10.3390/ plants11243450
- 66. Saraiva MP, Maia CF, Batista BL, Lobato AD. Ionic homeostasis and

redox metabolism upregulated by 24⊠epibrassinolide are crucial for mitigating nickel excess in soybean plants, enhancing photosystem II efficiency and biomass. Plant Biology. 2023;25:343-55. https://doi.org/10.1111/plb.13496

- 67. Kumari S, Chhillar H, Chopra P, Khanna RR, Khan MI. Potassium: A track to develop salinity tolerant plants. Plant Phys Biochem. 2021;167:1011-23. https://doi.org/10.1016/j.plaphy.2021.09.031
- Deng J, Kong L, Zhu Y, Pei D, Chen X, Wang Y, et al. BAK1 plays contrasting roles in regulating abscisic acid-induced stomatal closure and abscisic acid-inhibited primary root growth in *Arabidopsis*. J Inte Plant Bio. 2022;64:1264-80. https:// doi.org/10.1111/jipb.13257
- Zhong W, Xie C, Hu D, Pu S, Xiong X, Ma J, et al. Effect of 24epibrassinolide on reactive oxygen species and antioxidative defense systems in tall fescue plants under lead stress. Ecotoxi Environ Safety. 2020;187:109831. https://doi.org/10.1016/ j.ecoenv.2019.109831
- Guo C, Chen Y, Wang M, Du Y, Wu D, Chu J, Yao X. Exogenous brassinolide improves the antioxidant capacity of *Pinellia ternata* by enhancing the enzymatic and nonenzymatic defense systems under non-stress conditions. Front Plant Sci. 2022;13:917301. https:// doi.org/10.3389/fpls.2022.917301
- Shah AA, Khan WU, Yasin NA, Akram W, Ahmad A, Abbas M, et al. Butanolide alleviated cadmium stress by improving plant growth, photosynthetic parameters and antioxidant defense system of *Brassica oleracea*. Chemosphere. 2020;261:127728. https://doi.org/10.1016/j.chemosphere.2020.127728
- Guo J, Dong X, Han G, Wang B. Salt-enhanced reproductive development of *Suaeda salsa* L. coincided with ion transporter gene upregulation in flowers and increased pollen K<sup>+</sup> content. Front Plant Sci. 2019;10:333. https://doi.org/10.3389/fpls.2019.00333
- Hu D, Wei L, Liao W. Brassinosteroids in plants: crosstalk with small-molecule compounds. Biomolecules. 2021;11:1800. https://doi.org/10.3390/biom11121800
- Chen X, Xue H, Zhu L, Wang H, Long H, Zhao J, et al. ERF49 mediates brassinosteroid regulation of heat stress tolerance in *Arabidopsis* thaliana. BMC biology. 2022;20:254. https://doi.org/10.1186/s12915-022-01455-4https://doi.org/10.1186/1741-7007-3-1
- 75. Jia C, Zhao S, Bao T, Zhao P, Peng K, Guo Q, et al. Tomato BZR/BES transcription factor SIBZR1 positively regulates BR signaling and salt stress tolerance in tomato and *Arabidopsis*. Plant Sci. 2021;302:110719. https://doi.org/10.1016/j.plantsci.2020.110719
- Tian X, He M, Mei E, Zhang B, Tang J, Xu M, et al. WRKY53 integrates classic brassinosteroid signaling and the mitogen-activated protein kinase pathway to regulate rice architecture and seed size. The Plant Cell. 2021;33:2753-75. https://doi.org/10.1093/plcell/koab137
- 77. Solis CA, Yong MT, Zhou M, Venkataraman G, Shabala L, Holford P,

- et al. Evolutionary significance of NHX family and NHX1 in salinity stress adaptation in the genus *Oryza*. Int J Mole Sci. 2022;23:2092. https://doi.org/10.3390/ijms23042092https://doi.org/10.3390/ijms23042092
- 78. Gupta A, Shaw BP, Roychoudhury A. NHX1, HKT and monovalent cation transporters regulate K⁺ and Na⁺ transport during abiotic stress. In: Transporters and plant osmotic stress. Academic Press; 2021. pp. 1-27. https://doi.org/10.1016/B978-0-12-817958-1.00002-5
- Sun Z, Zou Y, Xie C, Han L, Zheng X, Tian Y, et al. Brassinolide improves the tolerance of *Malus hupehensis* to alkaline stress. Front Plant Sci. 2022;13:1032646. https://doi.org/10.3389/ fpls.2022.1032646
- Cao Y, Song H, Zhang L. New insight into plant saline-alkali tolerance mechanisms and application to breeding. Int J Mole Sci. 2022;23:16048. https://doi.org/10.3390/ijms232416048
- Azhar N, Su N, Shabala L, Shabala S. Exogenously applied 24epibrassinolide (EBL) ameliorates detrimental effects of salinity by reducing K+ efflux via depolarization-activated K+ channels. Plant and Cell Phys. 2017;58(4):802-10. https://doi.org/10.1093/pcp/ pcx026
- Sharma AK, Pradhan J, Kumar S, Pramanik K, Kastury C, Kumari G, et al. Root responses under water deficit stress: unraveling the impact on wheat crop and the ameliorating role of brassinolide. J Environ Bio. 2024;45(1):87-95. http://doi.org/10.22438/jeb/45/1/MRN-5169
- Purty RS, Kumar G, Singla-Pareek SL, Pareek A. Towards salinity tolerance in Brassica: an overview. Physiology and Molecular Biology of Plants. 2008;14(1):39-49. https://doi.org/10.1007/s12298-008-0004-4

#### Additional information

 $\label{per review} \textbf{Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.}$ 

**Reprints & permissions information** is available at https://horizonepublishing.com/journals/index.php/PST/open\_access\_policy

**Publisher's Note**: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etcSee https://horizonepublishing.com/journals/index.php/PST/indexing\_abstracting

**Copyright:** © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (https://creativecommons.org/licenses/by/4.0/)

**Publisher information:** Plant Science Today is published by HORIZON e-Publishing Group with support from Empirion Publishers Private Limited, Thiruvananthapuram, India.