



RESEARCH ARTICLE

Phytochemical screening of two Ilam native plants *Ziziphus nummularia* (Burm.f.) Wight & Arn. and *Ziziphus spina-christi* (Mill.) Georgi using HS-SPME and GC-MS spectroscopy

Mahmoud Bahmani¹, Anahita Jalilian², Iraj Salimikia³, Somayeh Shahsavari¹ & Naser Abbasi^{1*}

¹Biotechnology and Medicinal Plants Research Center, Ilam University of Medical Sciences, Ilam, Iran

²Department of Obstetrics and Gynecology, School of Medicine, Ilam University of Medical Sciences, Ilam, Iran

³Department of Pharmacognosy, School of Pharmacy, Razi Herbal Medicines Research Center, Lorestan University of Medical Sciences, Khorramabad, Iran

*Email: ilamfarma@gmail.com

ARTICLE HISTORY

Received: 24 December 2019

Accepted: 02 February 2020

Published: 07 May 2020

KEYWORDS

Ziziphus nummularia

Ziziphus spina-christi

chemical compounds

HS-SPME

GC-MS

ABSTRACT

Essential oils are aromatic compounds widely used in the perfumery, pharmaceutical and food industries. There are several methods to extract essential oils and extracts. These methods include distillation, pressure or scraping, pressing and razor technique, headspace solid-phase micro extraction (HS-SPME), solvent extraction, extraction by bioactive hydrolyzing enzymes, and CO₂ extraction. The aim of this study was to investigate the amounts of active ingredients of essential oils of medicinal plants *Ziziphus nummularia* and *Ziziphus spina-christi* by HS-SPME so that it can more easily move towards production of effective herbal remedies by identifying the main and active ingredients of the plants. The main chemical compound of *Z. spina-christi* essential oil was found to be trans-caryophyllene (17.31%), followed by alpha-pinene (15.50%), beta-caryophyllene (10.86%), and beta-pinene (7.32%). The main compound of *Z. nummularia* essential oil was tetradecane (16.76%), followed by hexadecane (9.35%), dl-limonene (5.75%), cyclohexan-1-ol, 3 meth (5.54%), trans-caryophyllene (5.47%), and beta-myrcene (5.28%). Chemical compounds of *Ziziphus spina-christi* included carbobicyclic or bicycleheptane, sesquiterpenes derived from germacrene, bicyclic monoterpenes, and monoterpenes, while the main compounds of *Z. nummularia* included monoterpenes, aliphatic hydrocarbons, alkane hydrocarbons, primary terpene compounds, and decarbonated alcohol. Identification of chemical and biological constituents of essential oils of medicinal plants is a valuable way to identify medicinal compounds that can be used to treat diseases by combining the traditional effects and the main compounds in the experimental pharmacy studies.

Introduction

Medicinal herbs are used for the treatment of variety of diseases and disorders (1–7). Herbs exert their effect through the active ingredients and antioxidants available (4–10). *Ziziphus* is an important genus of medicinal herbs that is important in traditional medicine. It is a genus of thorny trees and shrubs belonging to the Rhamnaceae family. There are about 3 species of *Ziziphus* that are distributed in tropical and subtropical regions. The fruit of this tree, also called *Ziziphus*, is edible. These trees are abundant in the wild. The trees are found in the tropical, western,

and southern coasts of Iran and mainly in Ilam, Khuzestan, Sistan and Balouchestan, Hormozgan, Kerman, Fars and Bushehr provinces. The plants are found scattered and people use its fruit (11). *Ziziphus* species include *Ziziphus abyssinica*, *Z. lotus*, *Z. mauritiana*, *Z. mucronata*, *Z. nummularia*, *Z. spina-christi* and *Z. zizyphus* (12). *Zizyphus spina-christi* belongs to Kingdom: Plantae, Order: Rosales. This plant is a tree of approximately 2 meters height with small, heart-shaped and extended leaves and three prominent veins, with its earrings turning into thorn. An important feature is the secretory system producing mucilage and gum, which contains single

© Bahmani et al. (2020). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>).

To cite this article: Bahmani M, Jalilian A, Salimikia I, Shahsavari S, Abbasi N. Phytochemical screening of two Ilam native plants *Ziziphus nummularia* (Burm.f.) Wight & Arn. and *Ziziphus spina-christi* (Mill.) Georgi using HS-SPME and GC-MS spectroscopy. *Plant Science Today* 2020;7(2):275–280.

<https://doi.org/10.14719/pst.2020.7.2.714>

Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, etc. Full list at <http://www.plantsciencetoday.online>

cells or secretory bags found in the skin and core of the stem in the parenchyma tissue of the vascular bundle around the veins of the leaf and petioles as well as outer part of the fruit (13, 14). In traditional medicine, *Z. spina-christi* is used for wound healing and hair growth and as an antimicrobial, antifungal, laxative, anti-bronchitis, analgesic, immune system-boosting, skin-refreshing, and detoxifying agent, to treat bone fracture, cancer, low back pain, to lose weight, etc. (15–17). The plant contains important medicinal compounds such as alkaloids, flavonoids, glycosides, saponins and essential oils (18, 19). It is found in Iran, Pakistan, Afghanistan, Iraq, Egypt and Israel (20–22). *Z. nummularia* or Ramilak is another species of *Ziziphus* is found in Afghanistan, India, Iran, Lebanon, Pakistan, Zimbabwe, Mauritania, Nigeria and Uganda. *Z. nummularia* leaves have been widely used in traditional medicine for the treatment of cold, skin diseases, pain, diabetes, wounds and also as an anti-inflammatory, antibacterial, antifungal, sedative and hypnotic agent (23–29). Phytochemical analysis revealed the presence of a number of phytoconstituents such as flavonoids, tannins, sterols, saponins, pectins, glycosides, alkaloids and tripeptides. Cyclopeptide alkaloids have been identified as active ingredient of *Ziziphus nummularia* (25–29).

Ziziphus spina-christi and *Z. nummularia* are used as edible fruits, jams, health products for skin and hair and medicinal effect. In traditional and Persian medicine of Iran and especially in Ilam province (west of Iran) from fruits and leaves of *Ziziphus spina-christi* and *Z. mauritiana* are widely used as medicinal and hygienic products so, In this study, we tried to understand the effective ingredients of these two plants. Finally, the active ingredients of these herbs will be helpful to produce effective pharmaceutical products.

Materials and Methods

Plant preparation

In October–November 2019, two species *Z. spina-christi* (Fig. 1) and *Z. nummularia* (Fig. 2) were procured from Dehloran County (south of Ilam province, west of Iran). The plants were identified and confirmed using morphological keys of Ilam Province Plant Flora at Ilam University of Medical Sciences Biotechnology and Medicinal Research Center. The collected plants were cleaned and dried in the shade in the open air. The dried plants were pulverized by a plant mixer and analyzed by HS-SPME for chemical composition.

Identification of chemical compounds by SPME-HS method

The essential oil of the plant was extracted by HS-SPME technique. About 2 gms of dried plant (from each plant separately) and its powder were placed in standard vial device and the vial temperature was set at 60–70 °C. These optimum temperature conditions will saturate the vapor content of the substances in the plant essential oil in the headspace of the solid surface (Headspace of the (the vial is a part of the HS-SPME equipment)). The SPME syringe with a lid on it



Fig. 1. *Ziziphus spina-christi*



Fig. 2. *Ziziphus nummularia*

was then placed in the headspace of the vial and the essential oils in the vial were absorbed by the silica phase in the instrument needle. After the silica fiber was allowed to sufficiently saturate with volatile components, the fiber was directly placed into the GC/MS input section and the essential oils in the fiber in the vial were adsorbed due to the temperature of the input and then the essential oils in HS-SPME device entered into the GC/MS apparatus for identification (30).

HS-SPME method

About 2 gm of each plant extract was used for analysis. The device condition was as follows: Gas chromatograph (Agilent 6890N) was coupled with Agilent 5973 Mass detector; Column: HP-5. (30 m length × 0.25 mm (ID) × 0.25 μm (stationary phase thickness)); Injector type: split/splitless and column temperature program: 50 °C, hold time 0.00 min and rate of -°C/min; temperature 200 °C, hold time, 0.00 min and rate of 5 °C/min and temperature 240 °C, hold time 0.00 min and rate of 10 °C/min. Carrier gas: He (99.999%); Injection type: splitless; Library: Wiley 7n; Injector temperature: 250 °C and flow rate: 0.9 mL/min. Extraction mode: (HSSPME); SPME fiber: PDMS 100 μm thickness (SUPELCO); sample weight: 0.5 gm; extraction temperature: 60 °C; extraction time: 20 min; sonication time: 10 min (Euronda

sonication instrument, Italy) and desorption time in GC-MS injector port: 3 min (31).

Results

In this study, essential oils of two plant species *Z. nummularia* and *Z. spina-christi* were extracted by HS-SPME and analyzed by GC-MS. Based on the results, a total of 41 chemical compounds were isolated from *Z. spina-christi* essential oil (Table 1). The results of phytochemical analysis showed that the main

Table 1. Identified compounds of *Ziziphus spina-christi* essential oils of by HS-SPME (GC-MS) method

No.	Retention time	Compound	%
1	5.942	α -Thujene	1.53
2	6.133	α -Pinene	15.50
3	7.07	Sabinene	2.54
4	7.170	β -Pinene	7.32
5	7.508	β -Myrcene	6.26
6	7.88	1-Phellandrene	0.61
7	8.018	Delta 3-Carene	0.68
8	8.22	4-Carene	0.14
9	8.560	β -Phellandrene	10.86
10	8.735	cis-Ocimene	2.02
11	9.03	trans-.beta.-Ocimene	0.26
12	9.348	γ -Terpinene	0.30
13	9.698	trans-Sabinene hydrate	0.65
14	10.242	Terpinolen	0.05
15	10.583	Linalool	0.28
16	11.748	trans-Pinocarveol	0.23
17	12.082	L-Menthone	4.90
18	12.332	Menthofuran	2.43
19	12.737	Menthol	1.28
20	12.99	3-p-Menthanol	0.50
21	13.053	trans-Carane	0.67
22	13.23	A-Terpineol	1.07
23	14.481	Pulegone	0.30
24	14.937	Piperitone	0.21
25	15.898	Carane	4.75
26	16.295	cis-Carane	0.27
27	17.848	α -Muurolene	0.16
28	18.088	α -Copaene	2.46
29	18.338	α -Bourbonene	0.23
30	18.448	β -Cubebene	0.08
31	18.517	β -elemene	0.32
32	19.266	trans-Caryophyllene	17.31
33	19.579	α -Bergamotene	0.64
34	20.121	trans- β -Farnesene	1.70
35	20.814	Germacrene-D	3.84
36	20.957	β -Selinene	0.70
37	21.185	Bicyclogermacrene	4.62
38	21.805	δ -Cadinene	1.15
39	22.207	CIS- α -Bisabolene	0.13
40	23.328	Caryophyllene oxide	0.93
41	23.581	p-Menth-3-en-9-ol	0.10

chemical compounds of *Z. spina-christi* essential oils was trans-caryophyllene (17.31%), followed by alpha-pinene (15.50%), beta-caryophyllene (10.86%), and beta-pinene (7.32%). The other compounds of the essential oil of *Z. spina-christi* are listed in Table 1. Based on the results of GC-MS analysis, a total of 39 chemical compounds were also isolated from the species *Z. nummularia*. The main compound of *Z. nummularia* essential oil was tetradecane (16.76%), followed by hexadecane (9.35%), dl-limonene (5.75%),

cyclohexan-1-ol, 3 meth (5.54%), trans-caryophyllene (5.47%), and beta-myrcene (5.28%). Supplementary information on other chemical compounds of the essential oil of *Z. nummularia* is given in Table 2.

Ziziphus spina-christi essential oils was trans-caryophyllene (17.31%), followed by alpha-pinene (15.50%), beta-caryophyllene (10.86%), and beta-

Table 2. Identified compounds in the essential oil of *Ziziphus nummularia* by HS-SPME (GC-MS)

No.	Retention time	Compound	%
1	6.669	α -Pinene	2.82
2	7.846	β -Pinene	1.78
3	8.298	β -Myrcene	5.28
4	9.561	dl-Limonene	5.75
5	12.018	Nonanal	1.40
6	13.761	Menthone	2.20
7	14.533	Menthol	1.23
8	14.63	Pyrazine, 2-methoxy-3-(2-methylpropyl)	3.60
9	15.19	Dodecane	4.87
10	15.487	Decanal	1.24
11	18.459	Menthyl acetate	1.88
12	18.557	Tridecane	1.45
13	20.311	5-Methyltridecane	1.20
14	20.843	2-Methyltetradecane	1.27
15	21.106	Copaene	1.93
16	21.666	Cyclotetradecane	1.44
17	21.774	Tetradecane	16.76
18	22.46	trans-Caryophyllene	5.47
19	22.877	α -Zingiberene	0.27
20	22.98	α -Guaiene	1.26
21	23.415	trans-Geranylacetone	1.33
22	23.557	Pentatriacontane	0.94
23	24.089	α -Amorphene	0.78
24	24.398	β -Ionone	2.73
25	24.615	Pentadecane	1.93
26	24.746	α -Muurolene	0.99
27	25.369	δ -Cadinene	1.85
28	25.832	Methylundecane	1.30
29	25.986	5-Methylpentadecane	1.13
30	26.455	3-Methylpentadecane	0.38
31	27.192	Hexadecane	9.35
32	28.358	Phytane	1.43
33	28.558	Lanol	0.54
34	29.564	Heptadecane	1.52
35	29.656	3-Cyclohexen-1-ol, 3-methyl-	5.54
36	31.37	Octadecane	1.51
37	31.485	Tritetracontane	1.17
38	31.965	Tetrahydrogeranylacetone	0.87
39	33.153	Dibutyl phthalate	1.60

pinene (7.32%). The other compounds of the essential oil of *Z. spina-christi* are listed in Table 1.

Fig. 3 and 4 comparatively illustrate the chromatograms of the compounds of *Z. spina-christi* and *Z. nummularia* essential oils. Apart from the difference in the percentages of compounds of the two plants', as shown in the chromatogram (No. 3) and Table 1, the plant has 41 chemical compounds in total. The main compounds of *Z. spina-christi* included carbobicyclic or bicycleheptane compounds, sesquiterpene derived from germacrene, monoterpenes as bicyclic monoterpenes. The chromatogram of *Z. nummularia* showed that the plant has a total of 39 chemical compounds of which, the main compounds are monoterpenes, aliphatic hydrocarbons, alkane

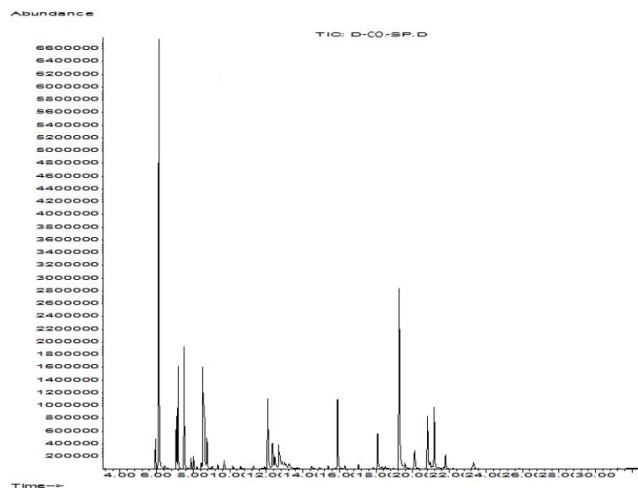


Fig. 3. Chromatogram of the essential oil of *Ziziphus spina-christi*.

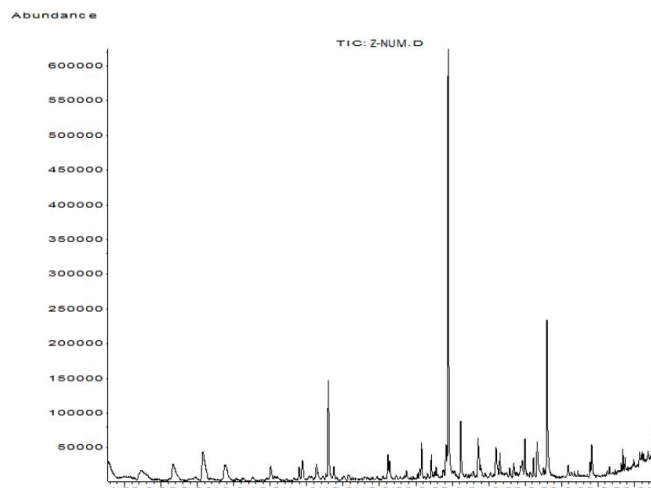


Fig. 4. Chromatogram of the essential oil of *Ziziphus nummularia*.

hydrocarbons, primary terpenes and decarbonated alcohol.

Based on the results of GC-MS analysis, a total of 39 chemical compounds were also isolated from the species *Z. nummularia*. The main compound of *Z. nummularia* essential oil was tetradecane (16.76%), followed by hexadecane (9.35%), dl-limonene (5.75%), cyclohexan-1-ol, 3 meth (5.54%), trans-caryophyllene (5.47%) and beta-myrcene (5.28%). A list of the main constituents along with the chemical formula and the chemical structure of the two plants are outlined in Table 3. View results of the phytochemical analysis show that there are common compounds between the two plants that differ in percentage. The results were reported in Table 4. Different compounds of *Z. spina-christi* are include α -Thujene, Menthofuran, Piperitone, Pulegone, Sabinene and γ -Terpinene. Also different compounds of *Z. nummularia* include Decanal, α -Zingiberene, α -Guaiene, α -Amorphene, phytane and Lanol.

Table 3. Structure and chemical composition of the two plants

<i>Z. spina-christi</i>		<i>Z. nummularia</i>	
Compound	Chemical formula	Compound	Chemical formula
trans-caryophyllene	C ₁₅ H ₂₄	Tetradecane	C ₁₄ H ₃₀
alpha-pinene	C ₁₀ H ₁₆	Hexadecane	C ₁₆ H ₃₄
beta-caryophyllene	C ₁₅ H ₂₄	dl-limonene	C ₁₀ H ₁₆
beta-pinene	C ₁₀ H ₁₆	cyclohexan-1-ol, 3 meth	C ₇ H ₁₂ O

Table 4. Comparison of differences in the percentages of common constituents between the two medicinal plants

Compound	Medicinal plants	
	<i>Z. spina-christi</i> (%)	<i>Z. nummularia</i> (%)
alpha-pinene	15.50%	2.82%
Beta-pinene	7.32%	1.78%
beta-myrcene	6.26%	5.28%
trans-Caryophyllene	17.31%	5.47%
α -Muurolene	0.16%	0.99%

Discussion

According to the studies (33–35), few phytochemical studies have been conducted on *Ziziphus nummularia* and *Z. spina-christi* (32). One of the important factors

in the differences in the chemical composition of a particular species is the growth in different geographical areas and ecological conditions. Despite being part of the plant family Rhamnaceae, These factors have certain differences and similarities with respect to chemical composition and secondary metabolites. In a study, benzaldehyde, phenylacetaldehyde, phenylethylalcohol, benzeneacetonitrile, 2-ethyl hexanoic acid, octanoic acid, 2-methoxy 4- (1-Propanol) -6-acetate phenol, nonanoic acid, decanoic acid, 1-hydroxy 2,4,6-trimethylbenzene and 5-hydroxymethyl-2-furaldehyde were identified from *Z. spina-christi* honey (33). Another study revealed that *Z. spina-christi* seed contained compounds such as 4-hydroxymethyl-1-methyl pyrrolidine-2-carboxylic acid (less polar and major compound) and 4-hydroxy-4-hydroxymethyl-1-methyl pyrrolidine -2-carboxylic acid (34). Chemical compounds of hexane and ethanolic extracts of *Z. nummularia* leaf revealed that hexane extract of the plant included 105 chemical compounds and its ethanolic extract included 56 chemical compounds including palmitic acid, linoleic acid, stearic acid, squalene, stigmasterol campesterol, vitamin E, geranyl linalool isomer, trans-geranylgeran oil, 1-eicosanol, gamma sitosterol, betulin, lupeol and phytol in the n-hexane extract while in addition to palmitic acid, linoleic acid, gamma sitosterol, stigmasterol, phytol, squalene, oleic acid, tricosane, tetradecane, 2-methoxy-4-vinylphenol, ethyl alpha-d-glucopyranoside and behenylbehenate were identified in the extracts of both extract (35). An investigation on the chemical constituents of *Z. nummularia* revealed that 4-hydroxy-cyclohexanone, 2,7-dimethyloctane-3,5-dione, heptacosane and 9,12-octadecadienoic acid methyl ester as the main compounds of the plant (36).

Hexadecanoic acid has antimicrobial, anti-allergic, antioxidant and pesticide effects (37–39). Octadecane has antifungal effect (40) and geranyl linalool isomer has antioxidant effects (41), partly explaining the properties of the two studied plants. The HS-SPME (GC-MS) technique helps to identify the essential oil of the plants. Identifying the plant's active ingredients along with documentation of the traditional and ethnobotanical effects of those plants leads us to a better understanding of *Ziziphus nummularia* and *Z. spina-*

christi pharmacological effects to produce natural and herbal remedies that are effective against diseases.

Acknowledgements

Authors would like to thank the Vice-chancellor for the Research and Technology Deputy of the Ilam University of Medical Sciences and Biotechnology and Medicinal Plants Research Center for funding this study. Grant's number is A-10-2667-1.

Authors' contributions

MB, NA and AJ reviewed the literature and prepared the first draft of manuscript; NA, MB, AJ and IS reviewed the literature, helped in preparing first draft of manuscript, checked and corrected the grammar. This study was designed and performed by MB, SSH and NA. All authors read and approved the final report.

Conflict of interests

The authors declare that they have no conflict of interests.

Funding/Support

Biotechnology and Medicinal Plants Research Center, Ilam University of Medical Sciences has supported this research.

References

1. Abbaszadeh S, Karami N, Bahmani F, Abbasi N, Atefi E. Headache and herbal medicine: An ethnobotanical study of Shahrekord, Southwest of Iran. *Plant Biotechnol Persa*. 2019;1(1):4-9. <http://pbp.medilam.ac.ir/article-1-35-en.html>
2. Zangeneh MM, Ghaneialvar H, Akbaribazm M, Ghanimatdan M, Abbasi N, Goorani S, et al. Novel synthesis of *Falcaria vulgaris* leaf extract conjugated copper nanoparticles with potent cytotoxicity, antioxidant, antifungal, antibacterial and cutaneous wound healing activities under *in vitro* and *in vivo* condition. *Journal of Photochemistry and Photobiology B: Biology*. 2019;197:111556. <https://doi.org/10.1016/j.jphotobiol.2019.111556>
3. Moayeri A, Azimi M, Karimi E, Aidi A, Abbasi N. Attenuation of morphine withdrawal syndrome by *Prosopis farcta* extract and its bioactive component luteolin in comparison with clonidine in rats. *Med Sci Monitor Basic Res*. 2018;24(9):151-58. <https://doi.org/10.12659/MSMBR.909930>
4. Bahmani M. A new method for promoting biologic synthesis and reducing the size of titanium dioxide nanoparticles (TiO₂ NPs) synthesized by *Origanum vulgare*. *Plant Biotechnol Persa*. 2019;1(1):10-12. <http://pbp.medilam.ac.ir/article-1-5-en.html>
5. Bahmani M, Taherikalani M, Khaksarian M, Rafieian-Kopaei M, Ashrafi B, Nazer M, et al. The synergistic effect of hydroalcoholic extracts of *Origanum vulgare*, *Hypericum perforatum* and their active components carvacrol and hypericin against *Staphylococcus aureus*. *Future Science OA*. 2019;5(3):FSO371. <https://doi.org/10.4155/fsoa-2018-0096>
6. Abbasi N, Khosravi A, Aidi A, Shafiei M. Biphasic response to luteolin in MG-63 osteoblast-like cells under high glucose-induced oxidative stress. *Iranian J Med Sci*. 2016;41(2):118-25
7. Jabbari N, Gheibi P, Eftekhari Z. The therapeutic effects of isolated Eugenol of *Syzygium aromaticum*. *Plant Biotechnol Persa*. 2019;1(1):42-44. <http://pbp.medilam.ac.ir/article-1-14-en.html>
8. Karimi E, Abbasi S, Abbasi N. Thymol polymeric nanoparticle synthesis and its effects on the toxicity of high glucose on OEC cells: Involvement of growth factors and integrin-linked kinase. *Drug Design, Development and Therapy*. 2019;13:2513-32. <https://doi.org/10.2147/DDDT.S214454>
9. Bahmani F, Kazemeini H, Hoseinzadeh-Chahkandak F, Farkhondeh T, Sedaghat M. Sedation with medicinal plants: A review of medicinal plants with sedative properties in Iranian ethnobotanical documents. *Plant Biotechnol Persa*. 2019;1(1):13-24. <http://pbp.medilam.ac.ir/article-1-11-en.html>
10. Serna-Escolano V, Serrano M, Valero D, Rodríguez-López MI, Gabaldón JA, Castillo S, et al. Effect of Thymol and Carvacrol Encapsulated in Hp-B-Cyclodextrin by Two Inclusion Methods against *Geotrichumcitri-aurantii*. *Food Microbiology & Safety*. 2019;84(6):1513-21. <https://doi.org/10.1111/1750-3841.14670>
11. Saied SA, Gebaur J, Hammer K, Buerkert A. *Ziziphus spina-christi* (L.) Willd: A multipurpose fruit tree. *Genet Resour Crop Evol*. 2008; 55:929-37. <https://doi.org/10.1007/s10722-007-9299-1>
12. Kadioglu O, Jacob S, Bohnert S, Nass J, Saeed ME, Khalid H, et al. Evaluating ancient Egyptian prescriptions today: Anti-inflammatory activity of *Ziziphus spina-christi*. *Phytomedicine*. 2016;23(3):293-306. <https://doi.org/10.1016/j.phymed.2016.01.004>
13. Jiang JG, Huang XJ, Chen J and Lin QS. Comparison of sedative and hypnotic effects of flavonoids, saponins and polysaccharids extract from semen *Ziziphus jujube*. *Natural Product Research*. 2007;21:310-20. <https://doi.org/10.1080/14786410701192827>
14. Amal H, Eman IA. Effect of *Ziziphus* leaves extract on mice suffering from ehrlich ascites carcinoma. *Nature Sci*. 2010;8:234-44
15. Kaur R, Kapoor K, Kaur H. Plants as a source of anticancer agents. *J Nat Prod Plant Resour*. 2011;1:119-24
16. Abdel-Wahhab MA, Omara EA, Abdel-Gali MM, Hassan NS, Nada SA, Saeed A, et al. A *Ziziphus spina-christi* extract protects against aflatoxin B1-Intitiated hepatic carcinogenicity. *Afr J Tradit Complement Altern Med*. 2007;4:248-56
17. Shahat AA, Pieters L, Apers S, Nazeif NM, Abdel-Azim NS, Berghe DV, et al. Chemical and biological investigations on *Ziziphus spina-christi* L. *Phytother Res*. 2001;9:593-97. <https://doi.org/10.1002/ptr.883>
18. Abalaka M, Mann A, Adeyemo S. Studies on *in-vitro* antioxidant and free radical scavenging potential and phytochemical screening of leaves of *Ziziphus mauritiana* L. and *Ziziphus spina-christi* L. compared with Ascorbic acid. *J Med Genet Genomics*. 2011;3(2):28-34
19. El-Kamali HH, Mahjoub SAT. Antibacterial activity of *Francoeuria crispa*, *Pulicaria undulata*, *Ziziphus spina-christi* and *Cucurbita pepo* against seven standard pathogenic bacteria. *Ethnobot Leaflets*. 2009;(6):6. <https://openiuc.lib.siu.edu/ebf/vol2009/iss6/6>
20. Abbasi AM, Khan MA, Khan N, Shah MH. Ethnobotanical survey of medicinally important wild edible fruits species used by tribal communities of Lesser Himalayas-Pakistan. *J Ethnopharmacol*. 2013; 48:528-36. <https://doi.org/10.1016/j.jep.2013.04.050>
21. Bachaya HA, Iqbal Z, Khan MN, Jabbar A. Anthelmintic activity of *Ziziphus nummularia* (bark) and *Acacia nilotica* (fruit) against Trichostrongylid nematodes of sheep. *J Ethnopharmacol*. 2009; 123:325-29. <https://doi.org/10.1016/j.jep.2009.02.043>
22. Desai AG, Qazi GN, Ganju RK, El-Tamer M, Singh J, Saxena AK, et al. Medicinal plants and cancer chemoprevention. *Curr Drug Metab*. 2008;9:581-91. <https://doi.org/10.2174/138920008785821657>
23. Ullah M, Khan MU, Mahmood A, Malik RN, Hussain M, Wazir SM, et al. An ethnobotanical survey of indigenous medicinal plants in Wana district south Waziristan agency, Pakistan. *J Ethnopharmacol*. 2013;150(3):918-24
24. Goyal M, Ghosh M, Nagori BP, Sasmal D. Analgesic and anti-inflammatory studies of cyclopeptide alkaloid fraction of leaves of *Ziziphus nummularia*. *Saudi Journal of Biological Sciences*. 2013; 20(4):365-71. <https://doi.org/10.1016/j.jep.2013.09.032>
25. Bodroth RP, Das M. Phytochemical screening and antimicrobial activity of ethanol and chloroform extract of *Ziziphus nummularis* Wt. & Arm. *African Journal of Biotechnology*. 2012;11(21):4929-33
26. Yusufoglu HS. Topical anti-inflammatory and wound healing activities of herbal gel of *Ziziphus nummularia* L. (F. Rhamnaceae)

- leaf extract. *International Journal of Pharmacology*. 2011; 7(8):862–67. <https://doi.org/10.3923/ijp.2011.862.867>
27. Ray SD, Ray S, Zia-Ul-Haq M, De Feo V, Dewanjee S. Pharmacological basis of the use of the root bark of *Ziziphus nummularia* Aubrev. (Rhamnaceae) as anti-inflammatory agent. *BMC Complementary and Alternative Medicine*. 2015; 15(1):416.
28. Dureja AG, Dhiman K. Free radical scavenging potential and total phenolic and flavonoid content of *Ziziphus mauritiana* and *Ziziphus nummularia* fruit extracts. *International Journal of Green Pharmacy*. 2012;6(3):187–92. <https://doi.org/10.4103/0973-8258.104929>
29. Rajasekaran S, Jaykar B, Anandan R, Aboobacker KP, Vannamalar S. Anti-diabetic activity of leaves of *Ziziphus nummularia* by dexamethasone induced diabetic rat model. *International Journal of PharmTech Res*. 2013; 5(2):844–51. <https://doi.org/10.1155/2017/4134093>
30. Heather Lord, Janusz Pawliszyn. *Journal of Chromatography A*. 2000; 885: 153.
31. Bahmani M, Taherikalani M, Khaksarian M, Soroush S, Ashrafi B, Heydari R. Phytochemical profiles and antibacterial activities of hydroalcoholic extracts of *Origanum vulgare* and *Hypericum perforatum* and Carvacrol and Hypericin as a Promising Anti-*Staphylococcus aureus*. *Mini Rev Med Chem*. 2019;19(11):923-32. <https://doi.org/10.2174/1389557519666190121124317>
32. Shonouda M, Angeli S, Schutz S, Vidal S. Use of CLSA and SPME-headspace techniques followed by GC-MS analysis to extract and identify the floral odorants. *Pak J BiolSci*. 2008;11,1246-51. <https://doi.org/10.3923/pjbs.2008.1246.1251>
33. Odeh I, AbuU-Lafi S, AL-Najjar I. Determination of Unifloral Honey Volatiles from *Centaureaiberica* and *Ziziphus spina-christi* by Solid-Phase Microextraction and Gas Chromatography-Mass Spectrometry. *Acta Chromatographica*. 2014;26(3):485–93. <https://doi.org/10.1556/AChrom.26.2014.3.7>
34. Said A, Huefner A, Abu Tabl ES, Fawzy G. Isolation and identification of two new cyclic amino acids from the seeds of *Ziziphus spina-christi* L. Willd) by means of ¹H-NMR, ¹³C-NMR, HSQC, HMBC and GC-MS. *IUFS J. Biol*. 2010;69(1):13-23
35. Prajapati S, Singh S. Phytoconstituents of *Ziziphus nummularia* (Burm. f.) Wight & Arn. leaves extracts using GC-MS spectroscopy. *Research & Reviews*. 2019; 9(1):109–18
36. Alfarhan AH, Rajakrishnan R, Al-Shehri MA, Al-Tamimi AB, Al-Obaid S, Khalaf S. Analysis of the cuticular wax composition and ecophysiological studies in an arid plant - *Ziziphus nummularia* (Burm. f.) Wight & Arn. *Saudi Journal of Biological Sciences*. 2020 Jan 1;27(1):318-23. <https://doi.org/10.1016/j.sjbs.2019.09.030>
37. Duke J. Dr Duke's phytochemical and ethnobotanical databases. 2014. Available from: <http://www.ars-grin.gov/duke>. Accessed on 1 December 2019
38. Diezel W, Schulz E, Shanks M, Heise H. Plants oils: Topical application and anti-inflammatory effects (Croton oil test). *Dermatologische Monatsschrift*. 1993;179:173-76.
39. Letawe C, Boone M, Pierard GE. Digital image analysis of the effect of topically applied linoleic acid on acne microcomedones. *Clinical and experimental Dermatology*. 1998;23(2):56-58. <https://doi.org/10.1046/j.1365-2230.1998.00315.x>
40. Kagoura M, Matsui C, Morohashi M. Carcinogenicity study of phytol (3,7,11,15-tetramethyl-2 hexadecen-1-ol) in ICR mice. *J Invest Dermatol*. 1993;101:460
41. Pal DK, Nandi M. CNS activities of *Celesia coromandeliana* Vahl. in mice. *Acta Pol Pharm and Drug Res*. 2005;62:355–61

