



REVIEW ARTICLE

Enhancing seed shelf life through modified atmospheric packaging: Mechanisms, benefits and future prospects

K Parameswari¹, T Kavichakravarthi^{2*}, R Brindavathy³, V Vijayageetha⁴, S Kavitha⁵, V Vakeswaran², R Kalaiyarasi⁶, P V Suriyaprakash² & J Harishbalaji²

¹Department of Sericulture, Forest College and Research Institute, Mettupalayam 641301, Tamil Nadu, India

²Department of Seed Science and Technology, Tamil Nadu Agricultural University, Coimbatore 641003, Tamil Nadu, India

³Oilseeds Research Station, Tindivanam 604 102, Tamil Nadu, India

⁴Krishi Vigyan Kendra, Tamil Nadu Agricultural University, Tindivanam 604 002, Tamil Nadu, India

⁵Centre for students welfare, Tamil Nadu Agricultural University, Coimbatore 641003, Tamil Nadu, India

⁶Department of Oilseeds, Tamil Nadu Agricultural University, Coimbatore 641003, Tamil Nadu, India

*Correspondence email - kaviagritn@gmail.com

Received: 14 January 2025; Accepted: 25 February 2025; Available online: Version 1.0: 08 May 2025

Cite this article: Parameswari K, Kavichakravarthi T, Brindavathy R, Vijayageetha V, Kavitha S, Vakeswaran V, Kalaiyarasi R, Suriyaprakash PV, Harishbalaji J. Enhancing seed shelf life through modified atmospheric packaging: Mechanisms, benefits and future prospects. Plant Science Today (Early Access). <https://doi.org/10.14719/pst.7224>

Abstract

Efficient seed storage is crucial for maintaining seed quality, viability and germination potential, which directly impact agricultural productivity and food security. However, prolonged storage leads to seed deterioration due to both biotic and abiotic factors, resulting in significant post-harvest losses. Modified atmospheric packaging (MAP) is a sustainable non-chemical technology that enhances seed storability by altering the gaseous environment around seeds. MAP operates through active packaging, which injects desired gases and passive packaging, where the product's respiration modifies the atmosphere. By reducing oxygen (O₂) levels, minimizing moisture absorption and inhibiting microbial and insect activity, MAP extends seed shelf life. Typically, O₂ is replaced with nitrogen (N₂) or carbon dioxide (CO₂), limiting oxidative reactions and metabolic activity. MAP benefits multiple seed types, including cereals, legumes, oilseeds and horticultural crops. Physically, it reduces seed moisture, delaying deterioration and preserving seed coat color. Physiologically, it prolongs seed longevity by reducing the accumulation of reactive oxygen species (ROS) and conserving seed reserves. Biochemically, it minimizes lipid peroxidation, maintains enzyme activities like alpha-amylase and catalase and reduces electrolyte leakage. Additionally, MAP controls storage pests and pathogens; elevated CO₂ inhibits insect proliferation and fungal contamination, mitigating mycotoxin risks. This review examines the strategic role of MAP in seed conservation for sustainable agriculture by comparing MAP with traditional storage methods and assessing its impact on seed viability and longevity.

Keywords: modified atmospheric packaging; packaging technology; seed storage; seed quality; shelf life

Introduction

A seed is a ripened ovule consisting of an embryo and a protective seed coat. Seeds contain reserve materials such as starch, oils and proteins, which are utilized during germination and are typically found in the endosperm or cotyledons (1). They represent the fruits of plant breeding and differ from grains, which are meant for food or raw materials. After harvesting, the seeds are dried to optimum moisture content (depending upon crops) and stored to maintain the initial seed quality until they are needed for planting. Proper storage conditions can preserve seed vigour and germination, which directly impact crop yield (2). Seed storage methods have evolved from ancient to modern times, with traditional practices like bamboo bins and earthen pots (3) and modern practices like ultra-cold storage and cryopreservation (4). Seed storage serves as a buffer between demand and production, maintains

genetic resources and preserves seed viability (5).

However, during the extended storage period, the seed undergoes deterioration, which is an undesirable and detrimental attribute of agriculture. Oxidative damage caused by reactive oxygen species (ROS) is a primary factor in seed aging, leading to protein oxidation, chromosomal abnormalities and DNA damage (6). Three-phase of hypothesis for seed deterioration was reported (7). This process is a separate event from seed development and germination (8). Several biotic (insect Infestation and pathogen infection) and abiotic (temperature, relative humidity, oxygen and moisture content) factors contribute to seed deterioration, making it very difficult to maintain viability during storage. Seed quality depends upon initial seed quality, temperature, moisture content and mycoflora (9). It is estimated that 25-33 percent of the world grain crop, including seeds, is lost each year during

storage (FAO). These losses reduce the efforts taken to increase agricultural production and the availability of quality seeds. Proper seed management practices during the storage period can minimize these losses (10), which can be implemented through modified atmospheric packaging (MAP). It has emerged as a promising approach to extending seed storability and maintain seed quality by reducing moisture content and oxygen levels and preventing insect infestation and pathogen infection (11).

MAP is defined as introducing carbon dioxide (CO₂) or nitrogen (N₂) into a storage container and restricting the concentration of oxygen (O₂) (12). It is the practice of packaging a perishable product that is kept in an atmosphere that has been modified. Hence the composition altered from the original atmosphere (13). MAP in seed storage is the best alternative to the use of chemicals and insecticides, which leave carcinogenic residues in the treated products (14). MAP differs from other non-chemical preservation methods, such as vacuum packaging and controlled atmosphere storage (CAS). While vacuum packaging eliminates oxygen, it lacks flexibility in gas exchange, which can be crucial for seeds requiring some metabolic activity (15). CAS, on the other hand, maintains a continuously monitored balance of gases but is more resource-intensive and costly (16). MAP provides gas modifications without the need for continuous monitoring, making it both adaptable and cost-effective.

MAP is preferred over conventional seed storage techniques due to its ability to create gaseous environments that optimize seed viability and longevity (17). It offers a sustainable chemical-free alternative that reduces oxidative stress and microbial contamination (18). This technology is particularly relevant in addressing climate-related storage challenges, such as fluctuating temperatures and increased humidity, which accelerate seed deterioration (19). By mitigating these environmental stresses, MAP enhances food security and supports resilient agricultural practices in the face of climate change. MAP's ability to maintain the catalase, peroxidase and alpha-amylase activity when the onion seeds are stored under CO₂ storage conditions (20). Similarly, the impact of MAP on groundnut kernel storage and found that N₂ storage significantly extended the shelf life while maintaining a higher germination rate (21). These studies reinforce the potential of MAP as a viable alternative to conventional storage.

Methodology

This review utilized Scopus, Web of Science and Google Scholar to ensure a comprehensive selection of studies. Inclusion criteria focused on peer-reviewed articles published since 2010 that examined the role of MAP in seed storage and conservation. Studies without empirical data or those centered solely on food packaging were excluded. Keyword searches included terms such as "MAP in seed storage," "seed viability under MAP," and "enzyme activity under MAP."

Packaging techniques for modified atmospheric

The packaging of modified atmospheric (MA) condition can be established with two approaches, such as

Active packaging

Active packaging involves creating a modified atmosphere inside a container by first applying a vacuum and then replacing the air with a specific gas mixture. This method can incorporate ethylene scavengers or emitters, which work together to establish the desired MA within the sealed package rapidly. Compared to the passive technique, the active packaging process is more efficient and less time-consuming (22). It is commonly used for preserving vegetable seeds, fresh-cut fruits and nuts (23).

Passive packaging

Passive packaging, also known as commodity-generated MAP, forms naturally within a sealed container due to the respiration process of the product, where O₂ is consumed and CO₂ is released. In this method, the packaging material allows oxygen to permeate into the container while excess CO₂ escapes. When the respiration rate of the product aligns with the gas permeability properties of the packaging film, a beneficial MA is created inside the container (24). This type of packaging is particularly suitable for storing potato tubers (25).

Applications of modified atmospheric packaging

MAP is a widely used technology for extending the shelf life and maintaining the quality of various perishable products, including fruits (26), vegetables (27), marine foods (28) and preserving food quality (29). It is highly beneficial for increasing the shelf life of processed foods. It reduces the availability of O₂ to microbes, retard their growth and allowing food to be preserved for an extended period. It also reduces all metabolic activity and respiration rates within the packing. It preserves the food item's original form and makes it appealing to the consumer. It also reduces the loss of moisture from the packaged food product. In agriculture, MAP helps in preserving seed quality during storage. It slowed the deterioration process and maintained higher physiological quality (30). During storage, there was less reduction in enzyme activity, protein content and seed leachate (31). Gases used in modified atmospheric packaging

Oxygen

It promotes the growth of aerobic bacteria while inhibiting the growth of anaerobic bacteria. In modified atmosphere packaging (MAP), oxygen plays a key role in maintaining myoglobin in its oxygenated form, oxymyoglobin. Usually, seed deteriorates quicker at higher storage temperatures, seed moisture content, or in the presence of high O₂ levels (32). A high O₂ level at storage reduces seed longevity (33).

Nitrogen

It is an inert, tasteless gas with minimal antibacterial properties. Because of its limited solubility in water, the addition of N₂ in MAP food helps avoid pack collapse caused by high CO₂ concentrations. Furthermore, by replacing O₂ in the pack, N₂ can slow oxidative rancidity while also inhibiting the growth of aerobic microbes (18).

Carbon dioxide

It is soluble in both water and lipids and it is primarily responsible for the bacteriostatic effect observed on microbes cultured in MA conditions. The antimicrobial effects of CO₂ were first noted in 1877 by Pasteur and Joubert, who found it could kill *Bacillus anthracis* (34). The CO₂ prolongs the microbial lag phase and slows growth during the logarithmic phase of development. For practical purposes, in most diets, gaseous CO applied to a living tissue would exist predominantly as soluble CO₂ gas and carbonic acid (35).

Gas composition and film permeability

The composition of gases in MAP plays a critical role in seed preservation. High CO₂ levels suppress microbial growth and insect activity, while reduced O₂ levels limit oxidative stress and metabolic degradation (36). N₂, being an inert gas, helps maintain a stable atmosphere by displacing O₂, reducing lipid peroxidation and enzymatic degradation (37). MAP effectiveness is significantly influenced by film permeability. It determines how gases like O₂ and CO₂ can pass through the film, which in turn affects the respiration rates of the produce and the overall atmosphere inside the package. Different types of polymeric films, such as polyvinyl chloride (PVC) and polyvinylidene chloride (PVDC), are utilized in Modified Atmosphere Packaging (MAP). These films have varying gas transmission rates (GTR) and water vapor transmission rates (38). The GTR of a film increases with the storage temperature and the magnitude of this increase varies with the film type and thickness. This helps in maintaining the quality of the produce for a longer period by reducing respiration rates, ethylene sensitivity, decay and physiological changes (39).

Nanomaterials can significantly enhance the barrier properties of packaging films, thereby reducing permeability to gases such as O₂ and CO₂. This helps in maintaining the desired atmosphere inside the package, which is crucial for preserving seed viability. Nanomaterials, such as silver nanoparticles, have antimicrobial properties. Incorporating these into packaging films can help protect seeds from microbial contamination, reducing the risk of seed-borne diseases. Nanomaterials can improve the mechanical strength of packaging films, making them more resistant to physical damage. This is particularly important for protecting seeds during transportation and storage (40).

Modified atmospheric packaging on seed quality during storage

Physical seed quality attributes

Moisture content: The primary factor contributing to seed deterioration during storage is high moisture content, which leads to reduced germination capacity (41). Seeds are inherently hygroscopic, meaning their moisture content fluctuates in response to changes in atmospheric humidity and temperature. When seeds are stored in gunny or cloth bags, they readily reach a state of moisture equilibrium with the surrounding environment. However, MAP conditions help to prevent moisture absorption from the external environment, thereby extending seed viability.

Research has demonstrated the effectiveness of MAP in reducing moisture content and preserving seed quality over time. In groundnut, the kernels stored under N₂ recorded low moisture content (7.0%) compared to other storage conditions at the end of 6 months of storage period (21). For instance, kabuli chickpea seeds stored in the MAP with a combination of 80% CO₂, 20% N₂ and 0% O₂ showed a low moisture content of 8.03% at the end of 14-month storage period, compared to the control (8.42%) and the germination percentage was recorded at a maximum of 92.38% due to low moisture content (42). Similarly, green gram seeds stored in an environment with 40%CO₂ maintained a low moisture content of 8.42% at the end of 6-month storage period (43). In another study, the onion seeds exposed to MAP for 12 months, with an initial moisture content of 6.10%, were subjected to 15 treatment combinations. Among these, the combination of 80% CO₂, 5 % O₂ and 15 % N₂ showed a minimal increase in moisture content, reaching only 6.17% at the end of the storage period. In contrast, seeds stored in cloth bags recorded the highest moisture content of 8.17% (44) (Table 1). Through MAP, the moisture content can be maintained within the permissible limit during storage, thereby preventing deterioration and extending seed viability (11, 45).

Seed coat color: The MAP techniques have demonstrated significant effects on preserving seed coat color by reducing O₂ levels, which slows the rate of oxidative reactions. The oxidative reactions are a primary cause of color deterioration as they affect anthocyanin pigment, which is responsible for seed coat color (46). The low-O₂ environments enhance pigment stability, thereby preventing browning or darkening of the seed coat (47, 48).

Research in various legumes and fruits illustrates the benefits of MAP in preserving seed quality. For instance, lentil seeds are stored in aluminum-laminated bags with N₂, CO₂ and air at temperatures of 15°C and 35°C. The N₂ and CO₂ present in atmosphere maintained seed coat color, germination percentage and hydration capacities over extended storage periods across temperature variations (49). Similarly, faba bean seeds are stored under CO₂, N₂, O₂ and vacuum conditions at 30°C in darkness for a year. The N₂ storage exhibited reduced color darkening and minimized tannin losses, maintaining the quality of seeds during long-term storage (50) (Table 1). Furthermore, pomegranate seeds stored in MAP at 1°C effectively retained anthocyanin levels (47). These findings underscore the importance of low-O₂ environments for pigment preservation and MAP conditions, significantly slowing down the anthocyanin degradation and helping to retain color and quality attributes in seeds and fruits during storage.

Physiological seed quality attributes

Seed longevity is intricately linked to various physiological traits such as germination percentage, root and shoot length, dry matter production, vigour index and speed of germination (45). A significant factor affecting seed longevity is reactive oxygen species (ROS). They act as a dual role in seed physiology, serving as signaling molecules at controlled levels to support germination but causing oxidative damage and cellular degradation when they

Table 1. Effect of Modified Atmospheric Packaging on the physical and physiological quality of seeds

SI. No.	Seeds and their storage duration	Impact of Modified Atmospheric Packaging	Reference
Physical seed quality	1. Groundnut (6 months)	At the end of 6 months of storage, kernels stored in N ₂ recorded low moisture content (7.0%) compared to other storage conditions	(21)
	2. Chickpea (14 months)	Low moisture content (8.03%) recorded at the end of 14 months of storage period compared to control (8.42%) under 80% CO ₂ + 20% N ₂ + 0% O ₂ gas combination	(42)
	3. Green gram (6 months)	Storing seeds under 40% CO ₂ gas, moisture content (8.42%) is maintained at the end of the storage period compared to control	(43)
	4. Onion (12 months)	In the 80% CO ₂ + 5 % O ₂ + 15 % N ₂ gas combination, there was the minimum increase in moisture content (6.17%) at the end of the storage period compared to the control (8.17%)	(44)
	5. Lentil (12 months)	The N ₂ and CO ₂ atmospheres maintained the seed coat colour, germination percentage and hydration capacities throughout the storage period	(49)
	6. Faba bean (12 months)	Storing seeds under N ₂ , reduces colour darkening and tannin losses, which would be useful in maintaining the quality of seeds during long-term storage.	(50)
Physiological seed quality	7. Groundnut (8 months)	Stored under N ₂ at -5°C condition, maintained germination above 70% even after 8 months of storage period	(54)
	8. Groundnut (12 months)	40 percent CO ₂ atmosphere storage maintains the vigour and viability up to 12 months of storage	(53)
	9. Amaranth (7 years)	Under the N ₂ condition, the viability and vigour are maintained for 7 years under the ambient storage condition	(55)
	10. Horse gram (6 months)	Maximum germination (89%) is observed at the end of storage period by storing under 50% CO ₂ atmosphere	(56)

accumulate excessively (51). MAP plays a crucial role in mitigating these effects by reducing O₂ levels and enhancing CO₂ and N₂ concentrations, which in turn slows down seed respiration rates and limits ROS production, thus preserving the seed's food reserves available to the seeds (52).

Research highlights the effectiveness of MAP in extending seed viability across various species. Groundnut pods or kernels with 8% moisture content stored either in the storage environment of 10°C with 40% RH or 40% CO₂ concentration can maintain vigour and viability for up to 12 months (53). Another study on groundnut stored with a 100% N₂ gas at -5 °C condition maintained germination above 70% even after 8 months of storage period (54). When the amaranth seed stored with N₂ gas in laminated aluminium foil pouches maintained viability and vigour for nearly 7 years under the ambient storage condition (55). Similarly, horse gram seeds are stored in 50% of CO₂ and recorded maximum germination of 89% at the end of the six-month storage period (56) (Table 1). Overall MAP, especially under conditions involving N₂, CO₂ and vacuum, effectively preserves the seed vitality.

Biochemical seed quality attributes

Seed storage studies reveal that biochemical changes during deterioration significantly impact seed quality and viability. During aging, enzymes viz., alpha-amylase, peroxidase, catalase, dehydrogenase and superoxide dismutase are positively correlated with seed quality (45) and decrease in various crops like wheat (57), soybean (58), onion (59) and groundnut (60). These enzymes play a crucial role in maintaining seed viability by scavenging free radicals produced during deterioration. Conversely, lipase activity increases with storage time and is negatively correlated with seed quality (60). Additionally, seed deterioration is associated with increased lipid peroxidation and higher electrical conductivity (EC) of seed leachates (59). Understanding these enzymatic and biochemical changes

can help to predict seed storage life and develop strategies to maintain seed quality during storage.

Alpha-amylase breaks down starch into simpler sugars, providing energy for seed germination (61). Reduced O₂ levels in MAP can slow down the activity of alpha-amylase, thereby reducing the rate of starch breakdown. This helps in conserving energy reserves and prolonging seed viability. The activity of peroxidase and catalase enzymes in seeds decreases during storage and leading to a decline the seed viability (60). This decline is associated with an increased accumulation of hydrogen peroxide (H₂O₂) and malondialdehyde content, as well as decreased germination and membrane deterioration (62). The reduced O₂ leads to lower ROS production. Peroxidase activity helps maintain cellular homeostasis by breaking down any formed ROS, thus protecting the seeds from oxidative damage. Similar to peroxidase, catalase activity is crucial in MAP for mitigating oxidative stress. The enzyme's activity ensures that any H₂O₂ produced during metabolic processes is efficiently broken down, preventing cellular damage (63). During stressful conditions; superoxide dismutases are produced and act as a defense mechanism against oxidative stress in plants and seeds. Additionally, these superoxide dismutases possess free radical scavenging properties by stabilizing the carbon bonds in free radicals (64). When the onion seeds were stored under CO₂ storage conditions with seed coating, they maintained storage potential by recording higher catalase, peroxidase and alpha-amylase activity (20). The dehydrogenase enzyme activity is a stable marker to estimate the degree of viability in seeds during storage. During storage, the enzyme activity is reduced and it is maintained under MAP conditions. The groundnut seeds exposed to the gas combination of 60% N₂ + 0% O₂ + 40% CO₂ showed higher dehydrogenase activity (0.133 OD value) compared to the control (0.068 OD value) at the end of ten month storage period (11) (Table 2).

Table 2. Effect of Modified Atmospheric packaging on Biochemical quality and seed Health Activity of seeds

Sl. No.	Seeds and their storage duration	Impact of Modified Atmospheric Packaging	Reference
Biochemical seed quality	1. Pigeon pea (10 months)	Under 40% N ₂ + 60% CO ₂ gas combination, less seed leachate (2.029 dSm ⁻¹) recorded when compared to control (2.207 dSm ⁻¹) at the end of storage period	(31)
	2. Sorghum (10 months)	Less seed leachate (1.001 dSm ⁻¹) compared to control (1.265 dSm ⁻¹) when seeds stored under 40% N ₂ + 60% CO ₂ gas combination	(67)
	3. Groundnut (10 months)	60% N ₂ + 40% CO ₂ combination showed higher dehydrogenase activity (0.133 OD value) compared to the control (0.068 OD value) at the end of ten months storage period	(11)
	4. Onion (6 months)	Storing seeds under CO ₂ conditions recorded higher catalase, peroxidase, alpha amylase activity	(20)
Seed Health activity	5. Green gram (6 months)	45% CO ₂ was effective in reducing bruchid infestation and subsequent progeny production for up to 2 months after storage	(71)
	6. Horse gram (6 months)	Seeds remained free from insect damage for up to six months under CO ₂ (50%)	(56)
	7. Wheat (6 months)	20 percent of CO ₂ was effective in achieving total insect mortality within a two month exposure period	(72)
	8. Maize (17 weeks)	Formation of mycotoxin is prevented by storing seeds under 90% CO ₂ + 5% O ₂ + 5% N ₂	(76)
	9. Groundnut (15 weeks)	Suppress the occurrence of mycobiota and preserve the quality of kernels in terms of color and lipid oxidation and reduce aflatoxin B1 contamination under 100% CO ₂ , 100% N ₂ and vacuum packaging	(77)

Lipase is a fascinating catalytic enzyme that facilitates the hydrolysis of ester bonds in triglycerides into free fatty acids and glycerol. It is negatively correlated with the seed quality. The higher accumulation of lipase enzyme was also evidenced by the higher accumulation of free fatty acids in seeds (60). The EC assesses the condition of a seed's cell membranes by measuring conductivity. During storage, the EC of seed leachates increases primarily due to the loss of membrane integrity at the cellular level, resulting in the release of macromolecules and electrolytes from seed cells. This process involves both passive diffusion of low molecular weight solutes and the release of macromolecules from ruptured cells. The leakage of intracellular substances, particularly enzymes and electrolytes, depends on the loss of testa integrity and subsequent cellular membrane damage due to oxidative stress (65). Seeds of inferior quality typically exhibit a weakened membrane structure, leading to the leakage of essential electrolytes like amino acids and inorganic ions (66). These substances are crucial for the rapid development and emergence of seedlings. Storing seeds under MAP conditions with reduced O₂ levels can decrease oxidative stress on seeds, which helps to preserve membrane integrity and retain nutrients, thereby maintaining seed quality and vigor. The investigation on MAP was conducted with pigeon pea seeds stored in a gaseous combination of 40% N₂, 0% O₂ and 60% CO₂. The results showed that the seed leachate was lower (2.029 dSm⁻¹) in the MAP treatment compared to the control, which recorded a leachate value of 2.207 dSm⁻¹ at the end of a 10-month storage period (31). Similarly, in a study on sorghum seeds stored under MAP, a reduced seed leachate value of 1.001 dSm⁻¹ was recorded in the same gas combination of 40% N₂, 0% O₂ and 60% CO₂, compared to the control, which had a higher leachate value of 1.265 dSm⁻¹ (67) (Table 2). These findings indicate that storing seeds under MAP significantly reduces the leakage of electrolytes and macromolecules from the seeds, as evidenced by the lower

electrical conductivity (EC) values. This suggests that MAP can help preserve membrane integrity by minimizing oxidative stress and reducing metabolic activity, which in turn leads to slower deterioration and better seed quality over time.

Seed health activity

Storage insects: Pests that infest stored grains can result in significant damage to the seeds. The extent of this damage is influenced by factors such as temperature, humidity and moisture levels during storage. Alterations in atmospheric conditions, such as increased CO₂ levels and decreased O₂ concentrations, can lower the metabolic activity of insects to a point where combined with the buildup of toxic byproducts, it leads to their demise (68). Modified atmosphere techniques, which involve adjusting the composition of gases surrounding the stored seeds, serve to diminish both the respiration rate of seeds and the activity of insects or microorganisms within them (69). In the case of groundnuts (11) and wheat (70) the N₂ or CO₂ gases are lethal to insects. Therefore, methods utilizing MA enriched with either N₂ or CO₂ are employed to manage insect infestations in storage. A study on the impact of various concentrations of CO₂ on seed infestation and population growth in green gram was performed and research indicates that green gram seeds infested with bruchids and subjected to CO₂ concentrations of 45% or higher exhibit significant effectiveness in reducing bruchid infestation and subsequent progeny production for up to 2 months after storage. Moreover, these treatments maintain a high germination percentage and seedling vigor (71). In Horse gram, seeds exposed to 40% and 50% CO₂ remained free from insect damage for up to six months. When the CO₂ was less, such as 30% or untreated, it showed higher levels of seed damage, with 3.23% and 15.33%, respectively. MS with elevated CO₂ concentrations effectively prevented the emergence of adult *Callosobruchus chinensis* insects during the six-month storage period (56). In wheat seeds,

storage insects can be controlled by exposure to 20% CO₂, which has proven effective in achieving total insect mortality within a two-month exposure period. If the CO₂ level is low such as 5% and 10%, results in partial reductions in population growth compared to the control. So this study shows the importance of utilizing an optimal CO₂ concentration for efficiently controlling insect populations and reveals the potential of CO₂ as a viable method for pest management (72) (Table 2). MA eradicates insects from stored products without contaminating the atmosphere and is safer than fumigants (73).

Seed pathogens: The moisture content of the seeds influences the biodiversity of fungi contaminating the seeds, which can, in turn affect their storage life and viability (74). The presence of specific mycoflora, such as *Aspergillus terreus*, *A. flavus*, *Penicillium expansum*, *P. frsquentans*, *P. reprens* and *P. patulum*, can lead to degradation of seeds, particularly in the presence of increased relative humidity (75). MAP with elevated levels of CO₂ is effective in reducing infestation and progeny production, as well as maintaining seed quality (56). When the maize seed is exposed to a gas composition of 90% CO₂ + 5% O₂ + 5% N₂ prevents mycotoxin formation. This highlights the potential of MAP, particularly in the context of elevated CO₂ levels, as a strategy to mitigate mycotoxin contamination in stored maize (76). The studies on groundnut suggest that MAP containing 100% CO₂, 100% N₂ and vacuum packaging exhibit the potential to suppress the occurrence of mycobiota and preserve the quality of kernels in terms of color and lipid oxidation and reduce Aflatoxin B1 contamination (77). When radish seed treated with chlorine dioxide and exposed to MA conditions, 5 or 10% O₂ with 10% CO₂ and no *Cronobacter* spp. were detectable on seeds treated with 100 mL/mL aqueous ClO₂. Moreover, 94.3% of the radish seeds germinated successfully after ClO₂ treatment, followed by exposure to MA conditions (78) (Table 2).

Future studies and cost-effective of MAP

Future studies should focus on assessing MAP's effectiveness under varying environmental conditions, including temperature fluctuations, humidity levels and different seed types. Investigating the integration of smart sensors for real-time monitoring of gas compositions and seed viability would further enhance the applicability of MAP. Additionally, research into biodegradable packaging alternatives could improve the sustainability of MAP, making it an even more viable solution for global seed conservation efforts. MAP is a cost-effective solution compared to traditional storage methods, particularly for smallholder farmers. While initial setup costs for MAP packaging may be higher than conventional storage bags, its long-term benefits outweigh the expenses. By reducing post-harvest losses, minimizing dependency on chemical treatments and extending seed viability, MAP offers economic advantages that improve seed accessibility and profitability for farmers operating on limited resources.

Conclusion

Modified atmospheric packaging (MAP) provides a transformative and sustainable solution for addressing the challenges of seed storage and deterioration. Traditional storage methods are limited by their inability to effectively manage factors such as moisture content, O₂ exposure and microbial or pest infestations, which accelerate seed aging and reduce viability. This review has demonstrated that MAP significantly improves seed quality by creating controlled environments with reduced O₂ and elevated N₂ or CO₂ levels. Despite its demonstrated advantages, MAP adoption faces challenges, including optimizing gas mixtures, packaging materials and economic feasibility for different seed varieties. Addressing these limitations through advanced technologies and improved infrastructure will be crucial for broader implementation. Future research should focus on refining MAP techniques, integrating smart sensors and developing cost-effective packaging solutions to maximize its impact. In conclusion, MAP represents a promising advancement in seed storage technology. Its ability to preserve seed quality and extend shelf life aligns with global efforts toward sustainable agricultural practices and food security. Wider application of MAP, supported by continued innovation, will ensure resilient seed systems and enhanced productivity, making it a cornerstone of modern seed management strategies.

Acknowledgements

We extend our gratitude to Tamil Nadu Agricultural University, who offered technical support, writing assistance and overall guidance.

Authors' contributions

PK & KT were involved in the preparation of the article, including data collection, drafting and editing the manuscript. PK & VV originated the idea for the review and developed its structure. BR & KR evaluated the manuscript and provided continuous support in revising and completing it. KS, VV, SPV & HJ contributed by critically reviewing and refining the manuscript.

Compliance with ethical standards

Conflict of interest: Authors do not have any conflict of interests to declare.

Ethical issues: None

References

1. Magray MM, Wani K, Chatto M, Ummyiah H. Synthetic seed technology. International Int J Curr Microbiol Appl Sci. 2017;6 (11):662–74. <https://doi.org/10.20546/jcmas.2017.611.079>
2. Gebeyehu B. Review on: Effect of seed storage period and storage environment on seed quality. IJAAS.2020;6(6):185–90. <https://doi.org/10.11648/j.ijaas.20200606.14>
3. Kiruba S, Sam Manohar Das S, Papadopoulou S. Prospects of traditional seed storage strategies against insect infestation adopted by two ethnic communities of Tamil Nadu, southern peninsular India. Bull Insectology. 2006;59(2):129.

4. Li D-Z, Pritchard HW. The science and economics of ex situ plant conservation. *Trends plant sci.* 2009;14(11):614–21. <https://doi.org/10.1016/j.tplants.2009.09.005>
5. Chala M, Bekana G. Review on seed process and storage condition in relation to seed moisture and ecological factor. *J Nat Sci Res.* 2017;7(9):84–90.
6. Choudhury A, Bordolui SK. Concept of Seed Deterioration: Reason, Factors, Changes During Deterioration And Preventive Measures to Overcome Seed Degradation. *Am Int J Agric Stud.* 2023;7(1):41–56. <https://doi.org/10.46545/aijas.v7i1.291>
7. Ebone LA, Caverzan A, Chavarria G. Physiologic alterations in orthodox seeds due to deterioration processes. *Plant Physiol Biochem.* 2019;145:34–42. <https://doi.org/10.1016/j.plaphy.2019.10.028>
8. Jyoti J, Malik C. Seed deterioration: a review. 2013:374–385.
9. Sujatha M, Uppar D, Biradaeatil N, Mummigatti U, Nagaratna S, Olekar. Biochemical changes in storage of groundnut seeds under different locations, storage containers and storage conditions. *Int J Curr Microbiol Appl Sci.* 2018;7:297–305. <https://doi.org/10.20546/ijcmas.2018.707.036>
10. Gofitshu M, Belete K. Susceptibility of sorghum varieties to the maize weevil *Sitophilus zeamais* Motschulsky (Coleoptera: Curculionidae). *Afr J Agric Res.* 2014;9(31):2419–26. <https://doi.org/10.5897/AJAR2014.8634>
11. Vasudevan S, Shakuntala N, Teli S, Goud S, Gowda B. Studies on effect of modified atmospheric storage condition on storability of groundnut (*Arachis hypogaea* L.) seed kernels. *Int J Res Stud Biosci.* 2014;2(2):25–36.
12. Ziegler V, Paraginski RT, Ferreira CD. Grain storage systems and effects of moisture, temperature and time on grain quality-A review. *J Stored Prod Res.* 2021;91:101770. <https://doi.org/10.1016/j.jspr.2021.101770>
13. Hintlian C, Hotchkiss J. The safety of modified atmosphere packaging: a review. *Food Technology.* 1986;40.
14. Bailly C. Active oxygen species and antioxidants in seed biology. *Seed science research.* 2004;14(2):93–107. <https://doi.org/10.1079/SSR2004159>
15. Narasimha Rao D, Sachindra N. Modified atmosphere and vacuum packaging of meat and poultry products. *Food Reviews International.* 2002;18(4):263–93. <https://doi.org/10.1081/FRI-120016206>
16. Thippareddi H, Phebus RK. Modified atmosphere packaging (MAP): Microbial control and quality. *FACTS National Pork Board.* 2002;4667:1–8.
17. Falagán N, Terry LA. Recent advances in controlled and modified atmosphere of fresh produce. *Johnson Matthey Technology Review.* 2018;62(1):107–17. <https://doi.org/10.1595/205651318X696684>
18. Farber J. Microbiological aspects of modified-atmosphere packaging technology-a review. *Journal of Food protection.* 1991;54(1):58–70. <https://doi.org/10.4315/0362-028X-54.1.58>
19. Zhou W, Chen F, Luo X, Dai Y, Yang Y, Zheng C, Yang W, Shu K. A matter of life and death: Molecular, physiological and environmental regulation of seed longevity. *Plant, Cell & Environment.* 2020;43(2):293–302. <https://doi.org/10.1111/pce.13666>
20. Sivasakthi S, Renugadevi J. Appraisal of storage potential of onion cv. CO (On) 5 seeds under carbon dioxide and ambient storage condition. *J Pharmacogn Phytochem.* 2020;9(5):925–32. <https://doi.org/10.22271/phyto.2020.v9.i5m.12351>
21. Thirumaniselvam K, Kali P, Renganathan U, Ramachandran K, Murugesan B, Venugopal V, Shanmugam K. Unveils the metabolic changes in groundnut CO 7 kernels stored under modified atmospheric conditions. *Plant Science Today.* 2024;11(4):1467–74. <https://doi.org/10.14719/pst.4997>
22. Zagory D, Kader AA. Modified atmosphere packaging of fresh produce. *Food technology (Chicago).* 1988;42(9):70–7.
23. Ghidelli C, Pérez-Gago MB. Recent advances in modified atmosphere packaging and edible coatings to maintain quality of fresh-cut fruits and vegetables. *Critical Reviews in Food Science and Nutrition.* 2018;58(4):662–79. <https://doi.org/10.1080/10408398.2016.1211087>
24. Robertson G. Modified atmosphere packaging. *Food packaging: principles and practice.* 2006:313–29. <https://doi.org/10.1201/9781420056150>
25. Abdelgawad G, Khater E, Bahnasawy AH, Mosa MM. Potato tubers quality as affected by modified atmospheric conditions and package type during storage. *Misr Journal of Agricultural Engineering.* 2023;40(3):227–42. <https://doi.org/10.21608/mjae.2023.213945.1101>
26. Zhang M, Meng X, Bhandari B, Fang Z, Chen H. Recent application of modified atmosphere packaging (MAP) in fresh and fresh-cut foods. *Food Reviews International.* 2015;31(2):172–93. <https://doi.org/10.1080/87559129.2014.981826>
27. Irtwange S. Application of modified atmosphere packaging and related technology in postharvest handling of fresh fruits and vegetables. *Agricultural Engineering International: CIGR Journal.* 2006.
28. Dermiki M, Ntzimani A, Badeka A, Savvaidis IN, Kontominas MG. Shelf-life extension and quality attributes of the whey cheese “Myzithra Kalathaki” using modified atmosphere packaging. *LWT-Food Science and Technology.* 2008;41(2):284–94. <https://doi.org/10.1016/j.lwt.2007.02.014>
29. García-Esteban M, Ansorena D, Astiasarán I. Comparison of modified atmosphere packaging and vacuum packaging for long period storage of dry-cured ham: effects on colour, texture and microbiological quality. *Meat science.* 2004;67(1):57–63. <https://doi.org/10.1016/j.meatsci.2003.09.005>
30. Capilheira AF, Cavalcante JA, Gadotti GI, Bezerra BR, Hornke NF, Villela FA. Storage of soybean seeds: Packaging and modified atmosphere technology. *Revista Brasileira de Engenharia Agrícola e Ambiental.* 2019;23(11):876–82. <https://doi.org/10.1590/1807-1929/agriambi.v23n11p876-882>
31. Manjunatha B, Vasudevan S, Umesha U, Sravani C. Studies on influence of modified atmospheric storage conditions on biochemical parameters in pigeonpea seeds. *Journal of Applied and Natural Science.* 2016;8(3):1249–52. <https://doi.org/10.31018/jans.v8i3.949>
32. Bass L. Seed viability during long-term storage. 1982.
33. Roberts E, Abdalla F. The influence of temperature, moisture and oxygen on period of seed viability in barley, broad beans and peas. *Annals of Botany.* 1968;32(1):97–117. <https://doi.org/10.1093/oxfordjournals.aob.a084202>
34. Davies A. Modified atmospheres and vacuum packaging. *Food Preservatives: Springer;* 2003. p. 218–39. https://doi.org/10.1007/978-0-387-30042-9_11
35. Daniels JA, Krishnamurthi R, Rizvi SS. A review of effects of carbon dioxide on microbial growth and food quality. *Journal of food protection.* 1985;48(6):532–7. <https://doi.org/10.4315/0362-028X-48.6.532>
36. Beaudry R, Luckanatinvong V, Solomos T, editors. Maintaining quality with CA and MAP. IV International Conference on Managing Quality in Chains-The Integrated View on Fruits and Vegetables Quality 712;2006. <https://doi.org/10.17660/ActaHortic.2006.712.26>

37. Gorris LG, Peppelenbos HW. Modified-atmosphere packaging of produce. Handbook of food preservation: CRC Press; 2020. p. 349-62. <https://doi.org/10.1201/9780429091483-26>
38. Mangaraj S, Goswami T, Panda D. Modeling of gas transmission properties of polymeric films used for MA packaging of fruits. Journal of food science and technology. 2015;52:5456-69. <https://doi.org/10.1007/s13197-014-1682-2>
39. Mangaraj S, Goswami T, Mahajan P. Applications of plastic films for modified atmosphere packaging of fruits and vegetables: a review. Food Engineering Reviews. 2009;1:133-58. <https://doi.org/10.1007/s12393-009-9007-3>
40. Sharma C, Dhiman R, Rokana N, Panwar H. Nanotechnology: an untapped resource for food packaging. Frontiers in microbiology. 2017;8:1735. <https://doi.org/10.3389/fmicb.2017.01735>
41. Meena M. Vacuum packaging as novel approach to extend storability and quality of various crop seeds. 2022.
42. Shinde P, Hunje R. Seed viability and seed health as influenced by modified atmospheric gases condition on Kabuli chickpea (*Cicer arietinum* L.) varieties. Journal of Entomology and Zoology Studies. 2019;7(1):713-9.
43. Raghupathi B, Padmasri A, Narayanamma VL, Ramya V. Effect of modified atmosphere with elevated levels of CO₂ on seed quality parameters of greengram during storage. The Pharma Innovation Journal. 2021;10(11):53-9.
44. Lamani K, Deshpande V, Patil N, Shashidhar T. Effect of Modified Atmospheric Packaging on Seed Longevity of Onion (*Allium cepa* L.) cv. Arka Kalyan. International journal of Current Microbiology and Applied Science. 2020;9(3):198-209. <https://doi.org/10.20546/ijcmas.2020.903.024>
45. Punithavathi R, Parameswari K, Umarani R, Eevera T, Anand T, Gomathi V. Exploring the Storage Potential of Groundnut Var. TMV (Gn) 13 under Modified Atmospheric Storage Conditions. International Journal of Environment and Climate Change. 2023;13(10):3982-9. <https://doi.org/10.9734/ijec/2023/v13i103072>
46. Enaru B, Dreţcanu G, Pop TD, Stănilă A, Diaconeasa Z. Anthocyanins: Factors affecting their stability and degradation. Antioxidants. 2021;10(12):1967. <https://doi.org/10.3390/antiox1012196>
47. Gil M, Artes F, Tomas-Barberan F. Minimal processing and modified atmosphere packaging effects on pigmentation of pomegranate seeds. Journal of Food Science. 1996;61(1):161-4. <https://doi.org/10.1111/j.1365-2621.1996.tb14749.x>
48. Daravingas G, Cain R. Thermal degradation of black raspberry anthocyanin pigments in model systems. Journal of Food Science. 1968;33(2):138-42. <https://doi.org/10.1111/j.1365-2621.1968.tb01338.x>
49. Bhattarai B, Walker CK, Wallace AJ, Nuttall JG, Hepworth G, Panozzo JF, Partington DL, Fitzgerald GJ. Modified storage atmosphere prevents the degradation of key grain quality traits in lentil. Agronomy. 2023;13(8):2160. <https://doi.org/10.3390/agronomy13082160>
50. Nasar-Abbas S, Plummer J, Siddique K, White P, Harris D, Dods K. Nitrogen retards and oxygen accelerates colour darkening in faba bean (*Vicia faba* L.) during storage. Postharvest Biology and Technology. 2008;47(1):113-8. <https://doi.org/10.1016/j.postharvbio.2007.06.007>
51. Bailly C, El-Maarouf-Bouteau H, Corbineau F. From intracellular signaling networks to cell death: the dual role of reactive oxygen species in seed physiology. Comptes Rendus Biologies. 2008;331(10):806-14. <https://doi.org/10.1016/j.crv.2008.07.022>
52. Groot SP, de Groot L, Kodde J, van Treuren R. Prolonging the longevity of ex situ conserved seeds by storage under anoxia. Plant Genetic Resources. 2015;13(1):18-26. <https://doi.org/10.1017/S1479262114000586>
53. Vakeswaran V. Assessment of the Pattern of Loss of Viability and Vigour of Ground Nut Seeds Stored as Pod and Kernel in Ambient and Modified Atmospheric Storage. International Journal of Agriculture Sciences, ISSN. 2017:0975-3710.
54. Gayathri M, Regis J, Eevera T, Amuthaselvi G. D-5793 [1-8] Storage Behaviour of Groundnut (*Arachis hypogaea* L.) Seeds under Different Storage Condition. Legume Research - An International Journal. 2023. <https://doi.org/10.18805/ag.D-5793>
55. Doijode S. Seed Germplasm Conservation with Modified Atmosphere Storage in Amaranth (*Amaranthus* spp.). Indian Journal of Plant Genetic Resources. 2001;14(2):288-9.
56. Divya P, Durga KK, Sunil N, Rajasri M, Keshavulu K, Udayababu P. Modified atmosphere storage technique for the management of pulse beetle, *Callosobruchus chinensis* in Horse gram. Legume Research-An International Journal. 2016;39(3):474-8. <https://doi.org/10.18805/lr.v0i0f.9610>
57. Chauhan D, Deswal D, Dahiya O, Punia R. Change in storage enzymes activities in natural and accelerated aged seed of wheat (*Triticum aestivum*). The Indian Journal of Agricultural Sciences. 2011;81(11).
58. Sheidaei S, Hamidi A, Sadeghi H, Oskouei B. Evaluation of initial seed quality and storage conditions on biochemical and physiological changes of soybean seeds. Iranian Journal of Seed Science and Technology. 2020;9(2):101-18.
59. Brar NS, Kaushik P, Dudi BS. Assessment of natural ageing related physio-biochemical changes in onion seed. Agriculture. 2019;9(8):163. <https://doi.org/10.3390/agriculture9080163>
60. Begum MAJ, Balamurugan P, Vanagamudi K, Prabakar K, Ramakrishnan R. Enzyme changes during seed storage in groundnut (*Arachis hypogaea* L.). Journal of Applied and Natural Science. 2014;6(2):748-50. <https://doi.org/10.31018/jans.v6i2.530>
61. Guglielminetti L, Yamaguchi J, Perata P, Alpi A. Amyolytic activities in cereal seeds under aerobic and anaerobic conditions. Plant Physiology. 1995;109(3):1069-76. <https://doi.org/10.1104/pp.109.3.1069>
62. Goel A, Sheoran I. Lipid peroxidation and peroxide-scavenging enzymes in cotton seeds under natural ageing. Biologia plantarum. 2003;46:429-34. <https://doi.org/10.1023/A:1024398724076>
63. Kamaei R, Kafi M, Afshari RT, Shafaroudi SM, Nabati J. Physiological and molecular changes of onion (*Allium cepa* L.) seeds under different aging conditions. BMC Plant Biology. 2024;24(1):85. <https://doi.org/10.1186/s12870-024-04773-7>
64. Fagundes C, Moraes K, Pérez-Gago MB, Palou L, Maraschin M, Monteiro A. Effect of active modified atmosphere and cold storage on the postharvest quality of cherry tomatoes. Postharvest Biology and Technology. 2015;109:73-81. <https://doi.org/10.1016/j.postharvbio.2015.05.017>
65. Duke SH, Kakefuda G, Harvey TM. Differential leakage of intracellular substances from imbibing soybean seeds. Plant Physiology. 1983;72(4):919-24. <https://doi.org/10.1104/pp.72.4.919>
66. Panobianco M, Vieira RD, Perecin D. Electrical conductivity as an indicator of pea seed aging of stored at different temperatures. Scientia Agricola. 2007;64:119-24. <https://doi.org/10.1590/S0103-90162007000200003>
67. Gangadhara T, Vasudevan S, Manjunatha B. Effect of Modified Atmospheric Storage Conditions on Biochemical Parameters of Rabi Sorghum Seeds. Advances. 2016:2912.
68. Donahaye E, Navarro S. Comparisons of energy reserves among strains of *Tribolium castaneum* selected for resistance to hypoxia and hypercarbia and the unselected strain. Journal of Stored Products Research. 2000;36(3):223-34. [https://doi.org/10.1016/S0022-474X\(99\)00044-2](https://doi.org/10.1016/S0022-474X(99)00044-2)

69. Jayas DS, Jeyamkondan S. PH-postharvest technology: modified atmosphere storage of grains meats fruits and vegetables. *Biosystems Engineering*. 2002;82(3):235-51. <https://doi.org/10.1006/bioe.2002.0080>
70. Moncini L, Simone G, Romi M, Cai G, Guerriero G, Whittaker A, Benedettelli S, Berni R. Controlled nitrogen atmosphere for the preservation of functional molecules during silos storage: A case study using old Italian wheat cultivars. *Journal of Stored Products Research*. 2020;88:101638. <https://doi.org/10.1016/j.jspr.2020.101638>
71. Durga KK, Sowmya T, Venkateshwaran K, Keshavulu K. Management of pulse beetle in green gram seed using modified atmosphere. *International Journal of Bio-resource and Stress Management*. 2022;13(11):1176-85. <https://doi.org/10.23910/1.2022.2835>
72. Bera A, Sinha S, Singhal N, Pal R, Srivastava C. Studies on carbon dioxide as wheat seed protectant against storage insects and its effect on seed quality stored under ambient conditions. *Seed science and Technology*. 2004;32(1):159-69. <https://doi.org/10.15258/sst.2004.32.1.16>
73. Cao Y, Xu K, Zhu X, Bai Y, Yang W, Li C. Role of modified atmosphere in pest control and mechanism of its effect on insects. *Frontiers in physiology*. 2019;10:206. <https://doi.org/10.3389/fphys.2019.00206>
74. El-Said A, Goder E. Effect of moisture contents on the biodiversity of fungi contaminating Cuminum cyminum and Pimpinella anisum seeds under storage periods and amylolytic activity of fungal isolates. *International Journal of Current Microbiology and Applied Sciences*. 2014;3(3):969-91.
75. Cîndea M, Ivaşcu A. Evaluation of biochemical changes induced by specific mycoflora in the wheat stored. *Lucrări Ştiinţifice-Universitatea de Ştiinţe Agronomice şi Medicină Veterinară Bucureşti Seria B, Horticultură*. 2009(53):725-8.
76. Riudavets J, Pons MJ, Messeguer J, Gabarra R. Effect of CO₂ modified atmosphere packaging on aflatoxin production in maize infested with *Sitophilus zeamais*. *Journal of stored products research*. 2018;77:89-91. <https://doi.org/10.1016/j.jspr.2018.03.005>
77. Opio P, Photchanachai S. Modified atmosphere influences aflatoxin B1 contamination and quality of peanut (*Arachis hypogaea* L.) kernels cv. Khon Kaen 84-8. *Journal of Stored Products Research*. 2018;78:67-73. <https://doi.org/10.1016/j.jspr.2018.06.005>
78. Kim EG, Ryu JH, Kim H. Effect of Chlorine Dioxide Treatment and Storage in a Modified Atmosphere on the Inactivation of *Cronobacter* spp. on Radish Seeds. *Journal of Food Safety*. 2013;33(2):172-8. <https://doi.org/10.1111/jfs.12037>

Additional information

Peer review: Publisher thanks Sectional Editor and the other anonymous reviewers for their contribution to the peer review of this work.

Reprints & permissions information is available at https://horizonpublishing.com/journals/index.php/PST/open_access_policy

Publisher's Note: Horizon e-Publishing Group remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Indexing: Plant Science Today, published by Horizon e-Publishing Group, is covered by Scopus, Web of Science, BIOSIS Previews, Clarivate Analytics, NAAS, UGC Care, etc
See https://horizonpublishing.com/journals/index.php/PST/indexing_abstracting

Copyright: © The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited (<https://creativecommons.org/licenses/by/4.0/>)

Publisher information: Plant Science Today is published by HORIZON e-Publishing Group with support from Empirion Publishers Private Limited, Thiruvananthapuram, India.